

# The NAC Protocol

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## A natural fungal mitigation and immunomodulatory protocol

### Introduction

*Fungal infections and their toxins (called mycotoxins) can be a major contributor to inflammation in the body. As a result of that inflammation, it can cause or exacerbate various conditions, including autoimmune diseases.*

*Relevant links to science articles will be presented as we go, listed so you can click on the link for more information. In addition, a more detailed scientific review of the protocol compounds is available at the end of the document for researchers and clinicians alike.*

*Fungal infections and their mycotoxins are seriously under-addressed in symptom management and the treatment of disease. This natural protocol focuses on effective elimination of fungal infections and modulation of immune response to improve outcomes.*

## Common Fungal Infections

Common Fungal Infections include *Candida*, *Aspergillus* and *Cryptococcus* species. Until recently, these fungi have been considered commensal, or a common part of the biome and generally considered harmless.

More recent research indicates that their byproducts (or metabolites) can be carcinogenic, inflammatory and even mutagenic to DNA [1].

These byproducts, or mycotoxins, are commonly measured in the body and regular exposure occurs through both food contamination [2] and inhalation of spores [3] in the home and outside environments.

By reducing fungal infection levels and their mycotoxins, we can reduce the corresponding inflammatory response in the body and promote homeostasis for improved immunity.

## A Natural Anti-Fungal Protocol

There are 3 components to this natural protocol. Those are N-Acetylcysteine (NAC), Oregano Oil (OO) and Black Seed Oil (BSO).

All 3 components work safely and effectively at reducing fungal infection and supporting the body's detoxification process. Part of the process of removing fungal infections in the body is dealing with the cell death of pathogens, which release toxins and byproducts that the body must eliminate through detoxification. This process occurs in the liver and kidneys. During this cleanup process, a **Jarisch-Herxheimer reaction** is common [4].

This reaction can include various symptoms like headaches, tiredness, gastrointestinal symptoms or

even cold like symptoms. This necessary discomfort is required to remove detrimental fungal infections.

Overall, it can take 2 to 4 months to see a significant improvement in health and well-being. Periods of detoxification symptoms will come and go as you progress. It is common to notice small improvements along the way, and it can be a motivator to complete the process.

The process takes longer because depending on the fungi in question, you may have infection in the lungs, gastrointestinal tract, upper respiratory system, spinal fluid, synovial fluid in the joints, various tissues and organs and even the brain itself.

Another thing that makes this process take longer is biofilms. What you normally call plaque on the teeth is an example of biofilms. 80% of the pathogens you are targeting live inside these biofilms. As a result they gain protection from the immune system.

Biofilms can take time to break down, but all 3 components of the protocol serve this purpose. The pathogens targeted by this protocol are both pathogenic fungi and bacteria. Both can be the cause of dysbiosis, or an imbalance in your gut biome, which can lead to various issues [5].

## The NAC Protocol

### Morning

- 600mg NAC
- Oregano Oil (40mg Carvacrol)
- Black Seed Oil (1 teaspoon)

### Night

- 600mg NAC
- Oregano Oil (40mg Carvacrol)
- Black Seed Oil (1 teaspoon)

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\*with contributions from S. C.

Continue daily for a minimum of two months and count out 3 weeks with no die off symptoms prior to discontinuing.

**Fungal die off symptoms may include:** *Tiredness, exhaustion, muscle soreness, increased chest or nasal discharge, cold or flu like symptoms, cold sores, headaches, rash, acne, irritability, change in stool frequency, volume or color; increased urination, bloated stomach, cramps, increased gas.*

### Choosing Your Supplements

Oregano Oil's bioactive ingredient is called **Carvacrol**. You want to purchase your OO in capsule form and purchase a product that has around 40mg of Carvacrol per dose. Read the bottle instructions to determine if 1 or 2 capsules will provide the necessary Carvacrol amount.

NAC can be found easily in 600mg per capsule dosages at various retailers and online shops.

Black Seed Oil should be purchased in the cold pressed, unfiltered oil form. A large 16 ounce bottle is available through **Horbaach** and **SVA Organics**. These products have been tested by users to be effective.

The morning and evening dose can be taken with or without food, but should be taken with food if you have a sensitive stomach. Increase water intake and fiber while on the protocol to ease detoxification.

### #1 Rule: Listen To Your Body

Everybody is different, and various factors can influence how strong your herxheimer reactions are, including level of infection, age and general health. Consult with your doctor before starting.

When you start this natural protocol, die off symptoms may be immediate or may take up to a month to begin. When this detox begins, listen to your body. If at any time the symptoms become too overwhelming, take a few days off, rest and increase your water intake.

You can then continue at lower dosages by reducing the Oregano Oil or taking it once daily, reducing NAC to once daily, and gradually increasing amounts as symptoms improve to reach protocol amounts.

As you advance further, it can be beneficial to scale up your dosages slowly to continue making progress.

If you decide to increase amounts, a general guideline for maximum daily intake based on studies is 400mg for Carvacrol amount, and 1800mg daily for NAC. Black Seed Oil can be maintained at 2 to 4 teaspoons daily.

Once you reach a period where die off stops occurring for 3 to 4 weeks and you are feeling good, you can take a break for a month and see how you react. If any symptoms return quickly, you may need

a more long term solution.

Some people require regular antifungals to stay healthy due to genomic defects [6]. If you find you need to continue with antifungals, the following page details an additional maintenance protocol for regular long-term use.

If you see no return of symptoms after a month off, use The NAC Protocol as necessary to reduce pathogen levels and maintain wellbeing.

## The Maintenance Protocol

After doing The NAC Protocol and taking a month long break, if you see prior symptoms return you may need a more long-term solution to maintain your health and vitality.

The Maintenance Protocol focuses on a more gentle antifungal approach combined with immune modulation to prevent overactive response (autoimmune).

To get technical for a moment, Niacin provides a needed boost in the NAD+ pool [7], which works with Pterostilbene as a SIRT1 activator [8] to promote homeostasis by countering oxidative stress, inflammation and mitochondrial dysfunction.

Pomegranate Extract serves to provide additional antifungal, anti-inflammatory and anti-oxidative support.

### Morning

- 600mg NAC
- 500mg Slo Niacin (nicotinic acid)
- 100mg Pterostilbene
- 250mg Pomegranate Extract (40% Ellagic Acid)
- Black Seed Oil (1 teaspoon)

### Night

- 500mg Slo Niacin
- 100mg Pterostilbene
- 250mg Pomegranate Extract
- Black Seed Oil (1 teaspoon)

After every 3 weeks on the maintenance protocol, take 1 week off. Continue to use black seed oil during the off cycle.

## Safety & Adverse Reactions

Always consult with your doctor before starting this protocol and receive prior approval. They can appropriately address interactions with medications or any current health conditions you have.

In addition, the protocol may lower specific vitamins and minerals including zinc, iron and calcium. A **multi-vitamin** is recommended to address this.

The protocol is known to lower blood sugar, blood pressure and can have a blood thinning effect.

In addition, asthmatics on corticosteroids should consider additional caution with NAC due to potential spasmodic activity. Refer to the science section for more detailed information.

Black seed oil contains thujone derivatives, which may aggravate certain conditions that are prone to seizures.

## Protocol User Feedback

We've shared this protocol online for over a year and a half as we gathered anecdotal reports from users on their experience.

In that time, we've received hundreds of positive anecdotal reports from users of all ages and backgrounds. Various inflammatory issues improved, aches, pains, flexibility issues, mood and well-being increased, various forms of dysbiosis were corrected (including irritable bowel disorder) and general health and well-being improvements were reported.

We hope you experience the wonderful improvements in health and vitality that many have reported. If it helps improve your quality of life, please consider sharing this important information with the people you care about.

**We wish you the best in health and vitality.**

## Advice For Clinicians

Frequent fungal infections like vaginitis (yeast infection), autoimmune skin disorders including seborrheic dermatitis, oral thrush on the tongue, chronic sinus issues, tooth decay and gingivitis, or recurrent infection of the skin, nails or mucous membranes should prompt further diagnostic tests to rule out *Candidiasis*, mold and mycotoxin exposure and genetic predispositions to fungal infections.

*Candidiasis* generally manifests as frequent yeast infections, fungal nail infections, white or yellow tongue thrush, advanced tooth decay, autoimmune skin disorders and infection of the mucous membranes.

*Candida* infections can vary in severity, with most testing done for *Invasive Candidiasis*, which is a more serious late stage infection. *Candida* actively works to penetrate the epithelial barrier of the gut, and if successful can be detected using blood culture tests, however culture tests have been shown to be mostly unreliable.

To rule out *Invasive Candidiasis* using non-culture tests, The Fungitell test (Associates of Cape Cod, East Falmouth, MA), multiplex PCR assays and T2Candida nanodiagnostic panel can be used, with 75% to 98% sensitivity. The Fungitell test should be

confirmed by two consecutive tests (80% sensitivity) and does not detect *Cryptococcal* infection [9].

If there is no positivity for *Invasive Candidiasis* and the patient is not immunosuppressed or compromised, a pattern of recurring infection should prompt further investigation. Frequent vaginitis non-responsive to fluconazole treatment, positive sputum cultures for *Candida* or persistent oral thrush that is non-responsive to fluconazole or nystatin oral suspension should prompt genetic testing if dysbiosis can be ruled out.

*Candida* spp. are associated with a number of specific gene mutations that predispose to fungal infections, impair immune response and improve the likelihood of both chronic infection and disseminated *Candidiasis* [10, 11, 12, 13, 14, 15, 16, 17, 18].

These autosomal dominant traits have been classified under *Familial Candidiasis* [6]. Molecular genetics tests are available to confirm [19].

It is important to stress that if a patient shows positivity for *Familial Candidiasis*, they will need a long-term antifungal solution in addition to an immunomodulatory solution to prevent autoimmune response. The NAC Protocol meets these requirements without the known hepatotoxicity of Amphotericin B or Fluconazole.

The NAC Protocol can be considered as a natural treatment option whenever signs of recurrent fungal infections are presented. The review at the end of this document covers the protocol's anti-fungal, anti-inflammatory, hepatoprotective and restorative benefits.

Additionally, mold exposure at home or in the workplace should be an additional query. *Aspergillus* spp. and their mycotoxins (Gliotoxin, Aflatoxin, Ochratoxin) should be ruled out in cases of chronic inflammation, frequent headaches, chest tightening or asthma, hemoptysis, eye symptoms or lung nodules detected during x-ray or CT scan. Both *Cryptococcus* and *Aspergillus* are frequently misdiagnosed as lung carcinoma [20, 21].

Antibody testing with *Aspergillus*-specific IgG can be used if *Pulmonary Aspergillosis* is suspected. Commercially available tests to detect serum galactomannan (early detection) and 1, 3  $\beta$ -D-glucan can be used as a non-culture based diagnostic.

Urinalysis detecting the primary mycotoxins (aflatoxin, gliotoxin, ochratoxin) can give a good baseline of exposure [22], especially in absence of known environmental exposure, and can indicate an active infection. Baselines would be lower from food contamination [23].

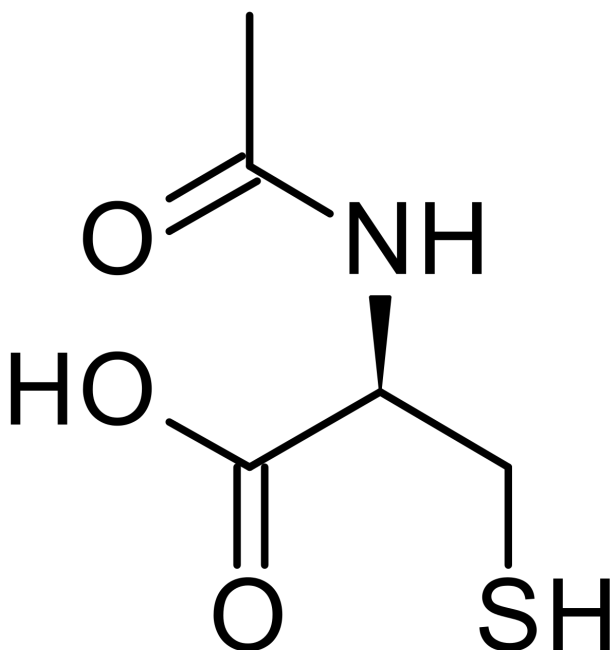


Figure 1: N-Acetyl Cysteine

## Science Behind The NAC Protocol

### N-Acetyl Cysteine

N-Acetyl Cysteine (NAC) is a derivative of amino acid L-cysteine, used clinically to treat acetaminophen overdose and associated hepatic injury. It's commonly used off label in the treatment of lung conditions, including COPD and cystic fibrosis [24]. The sulfhydryl grouping confers antioxidant effect, and NAC acts as a precursor to glutathione (GSH) production [25].

#### Primary Benefit And Methodology

N-Acetyl Cysteine (NAC) features a restorative and protective role in The NAC Protocol, both by ameliorating the genomic damage caused by fungal toxins and restoring excision and chemical repair of DNA.

The specific metabolites studied were Aflatoxin, Gliotoxin, Ochratoxin and Acetaldehyde.

Aflatoxin is a secondary metabolite of *Aspergillus*, specifically *A. flavus* and *A. parasiticus* [26]. Aflatoxin B1 (AFB1) is considered hepatotoxic, teratogenic and immunotoxic in humans [27].

Studies on a human epidermal cell line showed that concentrations of AFB1 >10  $\mu\text{M}$  are toxic to HaCaT cells and induce oxidative stress via ROS<sup>1</sup> and NO<sup>2</sup> generation [27].

Substantial damage to IMR32 neuronal cell lines was also observed, upregulating NOX2 and triggering DNA damage via downregulation of PARP1,

<sup>1</sup> Reactive Oxygen Species

<sup>2</sup> Nitric Oxide

BRCA2, and RAD51 [28].

Gliotoxin is also a toxic metabolite of *Aspergillus*, species *A. fumigatus*, and works via uptake of the disulfide bridge, which cycles between oxidized and reduced state, in turn generating ROS and destroying plasmid DNA [22]. Gliotoxin is also responsible for activating ROS-mediated apoptosis, and disrupting the integrity of the epithelial and endothelial barriers to enhance systemic fungal invasion [22].

Ochratoxin (OTA) is produced by multiple species of *Aspergillus* [29]. It is capable of inducing oxidative DNA damage and apoptosis, starting with glutathione depletion. Animal studies suggest that OTA-dependent oxidative stress is the precursor to cell lysing [30]. OTA concentrations were tested on a human renal proximal tubular epithelial cell line (HK-2), further confirming the role of oxidative stress in genotoxicity [31]. A study of OTA genotoxicity on porcine ovarian granulosa cells showed similar response to Aflatoxin, damaging repair related genes PARP1 and RAD51 [32].

Acetaldehyde is a metabolite of *Candida Albicans* resulting from glycolysis [32]. Both ROS and Ca<sup>2+</sup> pathways are involved in Drp1 phosphorylation and mitochondrial fragmentation. Elevation of Drp1 phosphorylation was partly dependent on ROS-mediated activation of c-Jun-N-terminal kinase (JNK) and p38 mitogen-activated protein kinase (MAPK) [33]. Acetaldehyde chemically induced DNA adducts follow a dose-response relationship, with mutagenicity frequently occurring as aldehyde dehydrogenase reductions become overwhelmed [34, 35].

Guanine is the most frequently oxidized DNA base, causing transversions in DNA replication [36]. O6-methylguanine (O6mG) is a common mispairing, causing GC to AT transversion. Repair of O6mG to guanine is done by O6-alkylguanine-DNA alkyltransferase (AGT), which requires cysteine [37]. 8-Oxo-7,8-dihydroguanine (8-oxoG) is also frequently oxidized, causing GC to TA transversions [36].

A study on mice containing *Ataxia telangiectasia*, which show continuous oxidative stress, showed that Thiol-containing NAC counteracts 8-OH deoxyguanosine, a marker for DNA deletions and genome instability [38]. Further, NAC was also shown to restore O6mG, likely by preventing modification of essential thiol groups [39].

By modulating the intracellular redox state, NAC can directly reduce oxidative-mediated apoptosis and DNA damage, acting as a scavenger for ROS and maintaining reduced glutathione (GSH) production in the liver [40]. Increased levels of GSH from supplementary NAC intake act as a catalyst with Glutathione S-transferases (GSTs) to reduce AFB1 by metabolizing and excreting it [41].

NAC inhibits Gliotoxin-induced apoptosis by blocking activation of caspase-3-like proteases and also scavenging intracellular ROS [42]. With OTA it inhibits apoptosis by preventing glutathione depletion [30].

Finally, NAC binds to Acetaldehyde acting as a scavenger, attenuating ROS and further carcinogenic or genotoxic effect [43].

### Anti-Biofilm Activity

The extracellular matrix of biofilms must be considered a target when eliminating fungal infections due to antimicrobial drug resistance and persistence of infections.

Biofilm formation by fungi and bacteria contribute to various pathogenic processes including gastrointestinal diseases, systemic autoimmune diseases, and neurodegenerative diseases [44]. Until more recently it was assumed that biofilms were formed exclusively by bacteria. Various pathogenic fungi can also form biofilms, including *Candida Albicans*, *Cryptococcus neoformans*, *Cryptococcus gatti*, *Aspergillus fumigatus* and *Saccharomyces cerevisiae*. The persistence of fungal infections is greatly enhanced by its ability to form biofilms [45, 46].

NAC is a powerful mucolytic antioxidant that efficiently inhibits and disrupts biofilms. *Pseudomonas aeruginosa* is an encapsulated bacterium that frequently causes infections in humans that are difficult to treat due to quick formation of biofilm.

At a concentration of 0.5 mg/ml NAC can detach mature *P. aeruginosa* biofilms, and at 10 mg/ml biofilms were completely disrupted [47]. A study on treatment of endodontic multi-species biofilms using NAC showed minimum inhibitory concentration (MIC) of 0.78–3.13 mg/ml. Multi-species culture consisted of *Actinomyces naeslundii*, *Lactobacillus salivarius*, *Streptococcus mutans*, and *Enterococcus faecalis* [48].

A study of NAC on *Candida Albicans* biofilm adhesion and disruption showed that NAC works effectively on mature biofilms (50-95% disruption) but less effectively on adhesion ( $\geq 32.8\%$ ). The study also showed increased efficacy when combined with ketoconazole, an antifungal [49].

*Cryptococcus Neoformans* requires capsular polysaccharide for biofilm formation, which primarily consists of Glucurunoxylomannan (GXM) and is also a constituent of cryptococcal biofilm. These biofilms are composed of 80% GXM which provides a unique challenge [50]. The trans-cell wall vesicular transport system of *Cryptococcus* is dependent on laccase [51], which is susceptible to NAC via a superoxide reaction to copper, converting it to H<sub>2</sub>O<sub>2</sub> [52]. This reaction in the laccase containing vesicle and corresponding membrane disruption appear to prevent

further virulence and tissue adhesion.

A study on wound biofilm formation treated with NAC showed interference with bacterial cellular redox states (NADH) and interference with ECM. Disruption of biofilms was primarily due to the molecular structure of NAC with acetyl and carboxyl groups [53].

### Protocol Synergy

NAC has shown a synergistic effect with many antifungals, decreasing the MIC values significantly [54]. It is believed that this is due to better penetration through membranes and biofilms due to its mucolytic effect, hydrolyzing glycoproteins and lipids via disulfide bonds and decreasing viscosity [55].

Additional benefit is provided by correcting the imbalance between reactive oxygen species (ROS) and glutathione depletion, which offers a protective effect combined with antifungals. As an inhibitor to c-Jun N-terminal kinase (JNK) it can also reduce endothelial dysfunction, inflammation and invasion [56].

The antifungal activity of Carvacrol induces ROS [57] which is ameliorated by NAC, as it is commonly used in clinical settings to identify and test ROS inducers [58].

NAC plays a supportive role, including ROS scavenging, disulfide reduction and glutathione replenishment.

A recent study on NAC further investigated the method of action and proposed an alternative function for antioxidant activity, suggesting that NAC uptake and deacetylation decelerate and prolong Cys delivery, releasing hydrogen sulfide (H<sub>2</sub>S), a product of Cys catabolism. A further product of H<sub>2</sub>S, sulfate sulfur species, is also proposed to contribute to NAC's beneficial effects as a cytoprotective [25].

### Safety Studies

NAC has a well-established safety profile, and its toxicity is rare. Elimination of NAC occurs through the renal system, with approximately 30% excreted through urine. In oral administration the most reported adverse effects are gastrointestinal symptoms such as nausea, vomiting or diarrhea [59].

Intravenous or oral inhalation can cause more serious adverse effects, including anaphylactoid reactions of flushing, itching, and angioedema, and systemic symptoms, such as bronchospasm and hypotension [60].

Oral dosages of 600mg and 1200mg daily showed no significant increase in adverse effects. Dosages as high as 3000mg daily resulted in minor gastrointestinal symptoms [61].

There was 1 fatal case of anaphylactoid reaction in a 40 year old woman with chronic asthma who

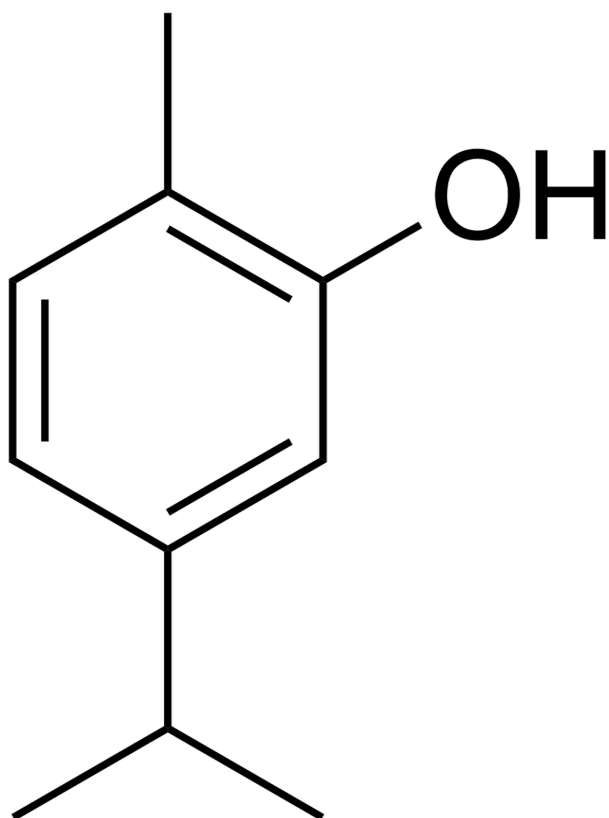


Figure 2: Carvacrol

received intravenous treatment [62]. A potential histamine response with asthmatic patients increases susceptibility to anaphylactoid reactions, and can potentially occur via oral administration [63]. Additionally, review of all available literature found no incidence of sulfa or sulfonamide allergy reactions with administration of NAC.

Reports of NAC preventing apoptosis have been a subject of debate. As an example, NAC can be beneficial to neuronal cells by preventing apoptosis caused by trophic factor deprivation [64] but in other cases can promote tumor progression by downregulating tumor antigen P53 [65]. Thymoquinone provides a counter to this with p53-mediated apoptosis [66], but more importantly is the action of *Origanum Vulgare* (Oregano) as it binds to sterols on the fungal membrane, specifically ergosterols and disrupts the permeability of the membrane leading to apoptosis. The two primary active compounds, Carvacrol [67] and Thymol [68] both contribute to this process.

## Oregano Oil

*Origanum Vulgare* (Oregano) contains two active compounds in abundance, Carvacrol and Thymol. Carvacrol is the primary constituent, a p-menthane monoterpene derived from cymene that provides many benefits to the human body [69].

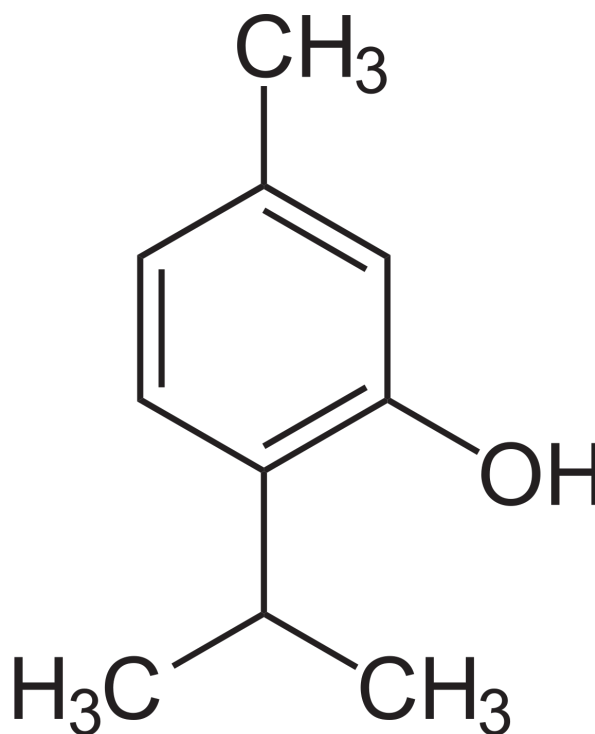


Figure 3: Thymol

### Primary Benefit & Methodology

Oregano serves multiple purposes as part of The NAC Protocol, including antifungal, anti-inflammatory and immunomodulatory roles.

Several additional benefits are obtained by using Oregano over other natural antifungals. One instance is dysbiosis, where imbalances in the mycobiota can influence homeostasis and disease progression [70]. Carvacrol works effectively against pathogenic bacteria and fungi [67, 71, 72, 73] to ameliorate dysbiosis. In one study of mice with *C. difficile* infection, Oregano Oil positively altered the microbiome composition, as revealed by an increased abundance of beneficial bacteria and reduced the proportion of detrimental flora [74].

A similar study on weaned piglets found that Oregano Oil supplemented in chow (25 mg/kg) showed a lowered population of *Escherichia coli* in the jejunum, ileum, and colon. They found that Oregano Oil promotes intestinal barrier integrity by correcting dysbiosis and lowering inflammation by measuring mitogen-activated protein kinase (MAPK), protein kinase B (Akt), and nuclear factor  $\kappa$ B (NF- $\kappa$ B) signaling pathways [75].

A study on oregano oil's effect on the intestinal barrier integrity of Hyland rabbits revealed that oregano essential oil increased significantly the gene expression of junctional adhesion molecule 2 (JAM2) and JAM3 in jejunum ( $p < .05$ ), showing a direct improvement in intestinal barrier function [76].

A study on broiler chickens fed dietary oregano in their feed showed a reduction in *Campylobacter* spp. and *E. coli*, with a significant increase in *Lactobacillus* spp [77]. While another broiler study showed similar results, with *Lactobacilli* raised ( $P < .001$ ) in ileum and cecum of all groups supplemented with Oregano [78].

These additional benefits to dysbiosis and intestinal barrier function were considered when choosing Oregano as the primary antifungal. Altered microbial composition, termed dysbiosis, has been implicated in mucosal barrier dysfunction and inflammatory responses. Restoring the epithelial barrier can potentially prevent autoimmune response and systemic infection [79].

### Antibiofilm Activity

The two primary compounds of *Origanum Vulgare*, Carvacrol and Thymol [69], show both powerful inhibitory and disruptive activity against biofilms. Pathogenic fungi can make their own biofilms [80] or cohabitate in multi-species bacterial biofilms where they arrange micro colonies with distinct features [81].

Therefore it is important to address mixed biofilms to effectively treat fungal infections. In a study on *Staphylococcus aureus* and *Candida albicans* in single and mixed cultures, Carvacrol showed a strong decrease of cell count, biomass, metabolic activity, and vitality of established 24- and 48-h biofilms [82]. A synergy was also shown between Carvacrol and Thymol in a similar study on *Candida albicans* and *Staphylococcus epidermidis*, where this combination killed highly tolerant persister cells of mono-species and mixed-species biofilms and demonstrated less risk of resistance development [83]. Effectiveness against *Salmonella Enteritidis* biofilms also showed Carvacrol and Thymol as effective, showing inhibition of biofilm formation at sub-minimum inhibitory concentration and effectiveness against preformed biofilms [84].

Effectiveness was also found against biofilms produced by pathogenic fungi. In a study on oral candidiasis, carvacrol and thymol significantly reduced both mature biofilm biomass and metabolic activity [85]. A study on the antibiofilm and antifungal activity against *Cryptococcus neoformans* and *Cryptococcus laurentii* compared Oregano oil (Carvacrol), Cinnamon oil (Cinnamaldehyde), Lemongrass oil (Citral), Clove oil (Eugenol), Peppermint oil (Menthol) and Thyme oil (thymol). The top 2 compounds for antibiofilm activity were Thymol and Carvacrol, respectively [86].

Method of inhibitory action on biofilms was elucidated in a *Salmonella typhimurium* biofilm study. Proteomic analysis showed changes in the proteins DsbA

(thiol: disulfide interchange protein DsbA), LuxS (S-ribosylhomocysteine lyase), DksA (RNA polymerase binding transcription factor DksA), and SODs (superoxide dismutases) A, B and C showed inhibited synthesis [87].

### Antifungal Activity

*Origanum Vulgare* (Oregano) and its primary constituents Carvacrol and Thymol have shown anti-oxidant, antiseptic, anticarcinogenic, anti-inflammatory, antidiabetic, immunomodulatory, antimicrobial, antispasmodic and antibacterial benefits. Effectiveness against a wide variety of pathogenic fungi and bacteria have been observed [88]. Carvacrol and Thymol are effective antifungal compounds that directly disrupt membrane integrity and ergosterol synthesis against *Candida* isolates [88].

Inhibitory activity against *Candida globosa*, *Candida albicans*, *Cryptococcus laurentii*, *Trichosporon asahii*, *Kodamaea ohmeri* and *Saccharomyces* using an Oregano ethanolic extract showed a MIC value of 1.56 mg/mL [89].

A study of Oregano against *Aspergillus flavus* and *Penicillium commune* as possible alternatives for food preservation showed a MIC of 4 mg/mL [90]. Effectiveness against *Aspergillus niger* and *Aspergillus flavus* was compared between oregano (*Origanum vulgare*), thyme (*Thymus vulgaris*) and clove (*Syzygium aromaticum*), with Oregano showing the highest inhibitory levels [91].

Tests of essential oils against heat resistant molds *Aspergillus fumigatus* and *Paecilomyces variotii* using citrus (*Citrus sinensis* L. Osbeck), laurel (*Laurus nobilis* L.), myrtle (*Myrtus communis* L.), oregano (*Origanum vulgare* L.), and savory (*Satureja thymbra* L.) showed Oregano as the most effective inhibitor of growth [92]. Another study on *Aspergillus niger*, *Aspergillus carbonarius*, and *Aspergillus wentii* showed inhibitory effect of 95.6%, 45.6%, and 100% at 2.5mL/100mL, respectively [93].

Effectiveness of essential oils tested against *Cryptococcus neoformans* and *Cryptococcus laurentii* showed Carvacrol and Thymol as most effective (16 and 32  $\mu\text{g/mL}$ ) as planktonic inhibitors, compared to Cinnamon oil (Cinnamaldehyde), Lemongrass oil (Citral), Clove oil (Eugenol), Peppermint oil (Menthol) and Thyme oil (Thymol) [86].

### Protocol Synergy

There are a number of likely synergies between NAC, Oregano Oil and Black Seed Oil based on available studies.

Thymoquinone (TQ) is the active compound in Black Seed Oil (*Nigella Sativa*). In a study of oral candidiasis, TQ was tested against *Candida albicans*, *Candida tropicalis*, *Candida glabrata* and *Candida krusei*

strains and the synergistic antifungal activity of these strains in combination with nystatin. With TQ alone *C. albicans* was significantly inhibited at 7.5 µg/mL. Nystatin showed inhibition against *C. albicans* at 1.875 µg/mL, but when combined with TQ it lowered MIC to 0.234 µg/mL showing a strong synergy [94]. TQ has also shown a synergistic effect against multi-drug resistant bacteria and fungi when combined with antibiotics [95] or antifungal treatments [96]. We believe there will be a similar synergy between Carvacrol, Thymol, and Black Seed Oil compounds, decreasing inhibitory concentrations and increasing effectiveness against multi-drug resistant fungi.

Determining chemical composition of *N. sativa* shows many potential synergies. Some of the additional active compounds found by GC and GC/MS analysis were trans-anethole, p-cymene and limonene [97]. Carvacrol and p-cymene have shown synergy as compounds, reducing the minimum inhibitory concentration of Carvacrol [98]. Studies on Limonene-Carcacrol (Lim-Car) have also shown synergy in inhibitory concentrations [99].

Synergy between NAC and antifungals has been shown previously with Fluconazole and Caspofungin [100]. Data infers that the mucolytic activity of NAC combined with the antifungal activity of Oregano provide an effective treatment against eukaryotic and sessile forms of pathogenic fungi.

### Immune Modulation

Oregano as part of The NAC Protocol acts as an immunomodulatory compound through several mechanisms. Both Carvacrol and Thymol play a role, with Thymol suppressing expression of iNOS and COX-2, blocking the phosphorylation of IκBα, NF-κB p65, ERK, JNK, and p38 MAPK [101]. Carvacrol showed similar effect against pro inflammatory IL-1b, COX-1 and COX-2, while upregulating IL-10 [102, 103] and demonstrating tissue healing ability against gastric ulcers and remodeling ability in a skin disease study. OEO significantly inhibited several inflammatory biomarkers, including monocyte chemoattractant protein 1 (MCP-1), vascular cell adhesion molecule 1 (VCAM-1), intracellular cell adhesion molecule 1 (ICAM-1), interferon gamma-induced protein 10 (IP-10), interferon-inducible T-cell alpha chemoattractant (I-TAC), and monokine induced by gamma interferon (MIG) [104, 105].

Oregano oil supports the immune system overall by also reducing mycotoxin burden via fungicidal action against susceptible pathogens like *Aspergillus* and *Candida*, as shown in **Oregano Oil Antifungal Activity** section

Additional immune modulatory benefit was demonstrated in **Oregano Oil Primary Benefit & Methodology** section, showing how Oregano can

ameliorate dysbiosis issues which disrupt homeostasis and promote disease progression.

Finally, the repair of the intestinal epithelial barrier by reducing inflammation and stabilizing dysbiosis promotes further immune enhancing effect. In all, Oregano is a powerful tool against pathogen-derived disruption of homeostasis and acute inflammatory response.

**Safety Studies** Oregano is one of the most widely studied natural antimicrobials, with animal studies in vitro and in vivo, as well as human clinical trials, approval as a food additive by the FDA, and used extensively as a food preservative to prevent spoilage.

A Phase I clinical study on the safety of Carvacrol studied 1mg/kg and 2mg/kg groups in a human trial for one month, finding all post-treatment measured parameters were within normal range. The results of this phase I study regarding carvacrol effects on healthy subjects, showed clinical safety and tolerability [106].

A Phase II clinical trial on the possible therapeutic effect of Carvacrol on asthmatic patients also showed no adverse outcomes [107].

Due to strong interest by the food industry for natural options for food preservatives, animal studies are also plentiful. A study on in vivo genotoxic effects produced in rats orally exposed to 81, 256 or 810 mg cavacrol/kg body weight (bw) at 0, 24 and 45 h found that carvacrol (81-810 mg/kg bw) did not induce in vivo genotoxicity or oxidative DNA damage in any of the tissues investigated [108].

Studies on Oregano Oil (OO) and Oregano Essential Oil (OEO) showed similar safety profiles. A study on OEO's oxidant effect (DPPH and ABTS assays) and cytotoxicity found OEO to be nontoxic [109]. A similar study on Wistar rats tested for genotoxicity over a 90 day trial, using 50, 100 and 200 mg/kg administered daily. Results obtained in the genotoxicity assays indicated lack of effect in micronucleus and standard comet assay under the conditions tested, showing no genotoxicity or oxidative damage to tissue [110].

The evidence currently suggests that Oregano Oil is safe for more long-term use, showing no indicators of oxidative damage, genotoxicity, mitochondrial dysfunction or morphological changes in healthy cells. Oregano Oil and its active compounds do show cytotoxicity against cancerous cells, however.

In a study on Acute myeloid leukemia cell lines (AML) carvacrol and thymol showed powerful synergy, inducing tumor cell death with low toxicity on normal cells. Cell death induced by the carvacrol and thymol combination is caspase-dependent in the HL60 cell line and caspase-independent in the other cell lines tested [111]. Furthermore, a study on



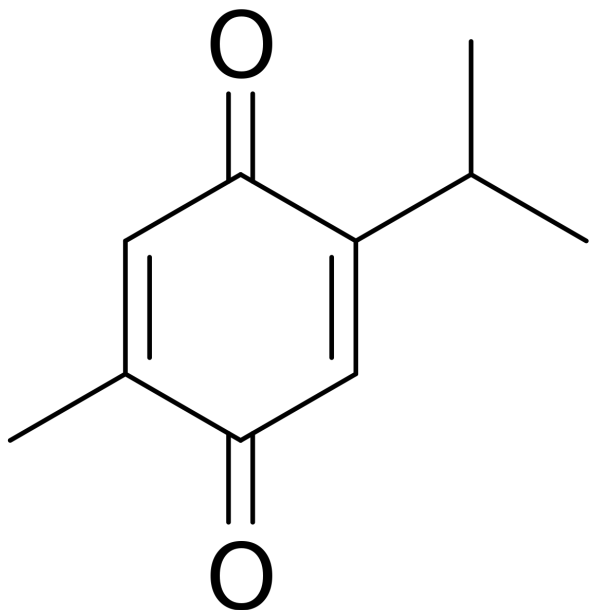


Figure 4: Thymoquinone

F1 DBA C57 Black hybrid mice studied OEO effect on Lewis carcinoma tumor engraftment. Mice were fed a low uptake dose of oregano essential oil with drinking water for three months, showing a tumor engraftment decreased by 1.8 times, its size decreased by 1.5 times, and the development of tumor was significantly suppressed. Interestingly, activity of antioxidant enzymes was found to increase after three months of essential oil uptake (by 1.5–3 times) as compared to the control group [112].

Oregano oil as part of The NAC Protocol is recommended at 40mg Carvacrol twice daily, or 80mg total intake per day. Safe levels were tested up to 600mg per day in the above referenced Phase I trial (2mg/kg), and up to 800mg/kg daily in animal trials showed no cytotoxic effects.

## Black Seed Oil

Thymoquinone, derived naturally from *Nigella Sativa*, is a natural compound with widespread protective effects, including anti-oxidative, anti-inflammatory, immunomodulatory, anti-cancer, and anti-microbial [113].

### Primary Benefit & Methodology

Black seed oil is typically produced using a cold press process, extracting the active compounds from the *Nigella Sativa* seed. A GC-MS analysis revealed more than 30 active compounds, including thymoquinone, fenchone, p-cymene, trans-anethole, limonene, carvone, carvacrol, longifolene and many additional active compounds [114].

The effect of Black seed oil (BSO) as part of The

NAC Protocol is multi-purpose, serving as hepatoprotective, a potentiator for antifungal activity, a biofilm disruptor, immune modulator, and a restorative which can increase t-cell count and differentiation [115, 98, 116, 117, 118].

A study of doxorubicin-induced cardiotoxicity in rats using 10mg/kg daily TQ in drinking water showed amelioration of induced cardiotoxicity. TQ proved to be a potent superoxide radical scavenger, with scavenging power being as effective as superoxide dismutase against superoxide [119]. A reduction of TQ in the liver to dihydrothymoquinone is part of this antioxidant mechanism, and combined they appear to mediate this protective action [120] and also act as effective OH radical scavenging agents [121]. TQ is known as a scavenger for hydroxyl and carbon centered radicals. It also shortens ROS-facilitated stress by yielding glutathionylated-dihydrothymoquinone via non-enzymatic reaction [122].

Antioxidant and Anti-inflammatory actions of TQ are the primary mechanisms that protect hepatocytes from injury. Myeloperoxidase activity in the liver tissue is an aggravating factor by increasing lipid peroxidation and free radical formation [123].

The hepatoprotective role of BSO is crucial as part of The NAC Protocol.

### Antibiofilm Activity

As part of The NAC Protocol BSO acts primarily as hepatoprotective, immune modulatory and an antifungal potentiator. Anti-Biofilm activity is also robust due to the abundant monoterpenes and sesquiterpenes [114]. Minimum biofilm inhibitory concentration (MBIC) for Thymoquinone ranges from 25-100  $\mu\text{g}/\text{mL}$ , with *Candida Albicans* being highly susceptible using in vitro assays [124]. *Staphylococcus aureus* and *Staphylococcus epidermidis* minimum biofilm inhibition concentration (BIC50) was reached with 22 and 60  $\mu\text{g}/\text{ml}$ , respectively. TQ also prevented cell adhesion [125]. *N. sativa* oil (BSO) showed highest microbial activity when compared to aqueous and methanolic extracts. BSO was also shown to reduce preformed biofilms of multi-drug resistant MRSA 1294, MRSA 1295 and MRSE 1297 effectively [126].

The complexity of bioactive ingredients plays a major role. In a study testing BSO against *Listeria monocytogenes*, a common food contaminant, 30 ligands were tested.  $\alpha$ -longipinene was selected based on in silico docking studies. Further in vitro studies demonstrated the anti-biofilm activity of  $\alpha$ -longipinene [127]. The complexity of terpenes in the volatile oil likely contributes to its broad spectrum effectiveness. This complexity leads to many potential synergies. p-cymene, a major constituent of BSO based on GC-MS analysis [114] has shown to

have a synergistic effect with  $\gamma$ -terpinene, carvacrol and other active compounds in BSO to increase anti-biofilm activity [128].

Studies on the individual active compounds in BSO show several unique anti-biofilm qualities. Limonene interferes with *C. albicans* biofilm adhesion, while trans-Anethole shows synergy with biofilm inhibition against *S. aureus* [129, 130].

### Antifungal Activity

*Nigella sativa* has been studied extensively for its pharmacological benefits, but antifungal research is limited. In a study of *N. sativa* in a methanolic extract, it was found effective against 20 different strains of *Candida* [131].

A further study on candidiasis of mice using an aqueous extract of *N. sativa* (6.6 mL/kg) showed significant inhibitory effect, only 24 hours after inoculation. A 5-fold decrease in *Candida* in kidneys, 8-fold in liver and 11-fold in spleen was observed [132]. Inhibitory effect on *Aspergillus parasiticus* (CBS 921.7) and *Aspergillus flavus* (SQU 21) was also demonstrated (1-3mg/100ml) using *N. sativa* oil (BSO) with potential metabolic effects on biosynthesis pathways for aflatoxin [133].

Investigating the composition of *N. sativa* volatile oil yields several active compounds, including thymoquinone, p-cymene,  $\alpha$ -thujene, limonene, trans-anethole, fenchone and carvacrol [114].

Thymol, thymoquinone (TQ) and thymohydroquinone (THQ), all constituents of *N. sativa*, were tested against 30 pathogens acquired from patients at a concentration of 1mg/mL. 100% inhibition was demonstrated against eight dermatophyte, five yeast and five mold isolates. TQ was found to be the strongest antifungal compound against dermatophytes and yeasts. Thymol was the most effective against molds [134].

A study on human infection by *Fusarium solani*, a filamental fungi from the *Nectriaceae* family, was performed comparing Thymoquinone to Amphotericin B. A 10 day inhibition test using 1mg/mL was performed. TQ demonstrated 100% inhibition by day 10, however Amphotericin B only inhibited 72.4% of growth in the same time range [135].

P-cymene has shown to be effective against drug-resistant forms of *Candida*, showing synergy when combined with Thymol [136].

Trans-anethole also has strong antifungal properties. Fennel is known as a strong antifungal, which is composed primarily of trans-anethole [137]. Trans-anethole has demonstrated effect with other drugs as it exhibits synergistic activity against several fungi [136].

Fenchone was also shown to inhibit fungal growth (32-64  $\mu$ g/mL) testing against *Candida albicans* ATCC-

76645 and LM-05, *Candida tropicalis* ATCC-13803 and LM-20, and *Candida Krusei* ATCC-6258 [138]. Limonene was also shown effective against *C. tropicalis* (20-40  $\mu$ L/mL) using potato dextrose broth [139].

### Protocol Synergy

*N. sativa* oil (BSO) has shown synergy with both antifungals and antibacterials as a potentiator [95, 96]. Active compounds in BSO have also shown direct synergy with Carvacrol, including p-cymene and limonene [98, 99]. Carvacrol and Thymol, the primary active compounds in Oregano are also featured in BSO [114]. BSO has been found effective against multi-drug resistant *S. aureus*, *P. aeruginosa* and *C. albicans*, and Carvacrol performs similarly [140, 141].

Carvacrol and Thymol concentrations in BSO are in lower concentrations [114] but when adding Oregano, which contains higher levels of Carvacrol and Thymol, two distinct methods of action are present [69]. Thymoquinone has been observed to disrupt *C. albicans* cell wall synthesis, disintegrate the cytoplasm and act as a pro oxidant inducing oxidative stress via ROS generation [142, 143]. Differentially, Oregano disrupts the cell membrane by interrupting ergosterol synthesis [88].

N-Acetylcysteine (NAC) was studied on chronic wound biofilms using mice with a 20 day maturation period. *Pseudomonas*, *Staphylococcus*, *Acinetobacter*, and *Enterobacter* were identified in the wound biofilm. NAC demonstrated effectiveness in disrupting the extracellular matrix of the biofilm, penetrating the bacterial cell membrane, inducing oxidative stress and disrupting protein synthesis [53].

We believe the mechanism of NAC combined with the antifungal compounds in *N. sativa* and Oregano provides a specific advantage when treating fungal and bacterial infections. This combination is crucial with up to 80% of the targeted pathogens residing in biofilms [144].

### Immune Modulation

*Nigella Sativa* has been used in Middle Eastern folk medicine since biblical times, with modern research showing *N. sativa* has effect on respiratory problems, dyspepsia, metabolic syndrome, *diabetes mellitus*, inflammatory diseases, and various types of cancer [145, 146].

The primary purpose of Black Seed Oil (BSO) as part of The NAC Protocol is both hepatoprotective and immunomodulatory. BSO has potent antioxidant effect over several pathways, modulating NF- $\kappa$ B, inhibiting iron-dependent lipid peroxidation, elevation in total thiol content and (GSH) level, radical scavenging, increasing the activity of quinone reductase, catalase, superoxide dismutase (SOD) and glutathione transferase (GST) and inhibiting COX/LOX

[147, 148].

As an anti-inflammatory, Thymoquinone (TQ) inhibits JNK, ERK and P38 phosphorylation and PI3K/mTOR signaling activation. In addition, BSO was shown to decrease lipid profiles (TG, TC, LDL, VLDL), liver enzymes (AST and ALT), hs-CRP inflammatory marker, IL-6 and TNF- $\alpha$  [149, 150, 151].

As an immunomodulator, BSO can directly improve immune response to fighting infection. A study of immunostimulation on a murine macrophage cell line showed that *N. sativa* ethanolic extract directly increased macrophage count in a cell proliferation assay, showing up to an 138% increase (25  $\mu$ g/ml) [152]. An additional study using ethanolic extract on blood derived, splenic and peritoneal macrophages showed a remarkable increase in phagocytic activity [153].

Immunostimulatory effect has also been demonstrated with peripheral blood mononuclear cells (PBMCs), LPS-induced doubling in phagocytic activity and upregulation of p-I $\kappa$ B $\alpha$  and p-NF- $\kappa$ B p65 [154, 155].

*N. sativa* can also enhance survivability in CD8-Positive T Cells by enhancing cytokine interferon- $\gamma$  (IFN $\gamma$ ) production. A study on the immunomodulatory effect of BSO on rheumatoid arthritis found a positive Modulation of T lymphocytes as well [156, 157].

BSO strengthens immune response, T Cell proliferation and function, supporting the body's response against infection.

### Safety Studies

*N. sativa* preparations have shown to provide gastro-protective, neuroprotective, anti-cancer, anti-diabetic, cardioprotective, bone regenerative and anti-arthritis effect [158, 159, 160, 161, 162, 163].

Several acute and subchronic toxicity tests have been carried out on *N. sativa*. Acute oral administration (LD50) was measured in mice (2.4g/kg) with signs of toxicity being difficulty in respiration and hypoactivity. Acute and sub-acute toxicity was measured in Sprague Dawley rats showing LD50 of 2000mg/kg, with sub-acute dosage of 500mg/kg showing a decrease in AST enzymes. No lethality was observed in all dosage groups (100, 500, 1000 and 2000 mg/kg). Analysis of liver and kidney observed no adverse morphology and BSO was considered safe and non-toxic [164, 165].

A Phase I human clinical trial on the safety of Thymoquinone (TQ) with Patients with Advanced Refractory Malignant Disease. 21 patients received 1 to 20 weeks treatment (median 3.7 weeks) with no side effects reported. No maximum tolerated dose was identified (75mg/day to 2600mg/day) [166].

An additional randomised, double-blinded

placebo-controlled Phase I human clinical trial was carried out on 70 individuals for a period of 90 days. Blood and serum collection tests were performed. Liver function parameters included alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP). Lipid profiles included Total cholesterol (TC), Low density lipoprotein (LDL), High density lipoprotein (HDL), Very low-density lipoprotein (VLDL) and Triglycerides (TG). Renal function markers (creatinine) were also tested. Recruited participants did not exhibit any clinical signs of toxicity or adverse effects. Liver toxicity and kidney function markers showed no change, however, lipid profiles showed significant decrease but were within safe limits. Change in TC, TG, LDL, VLDL and HDL were 12.1%, 19.66%, 16.33%, 12.76%, 8.21% and 15.27%, respectively [167].

## References

- [1] Anna Kowalska et al. "Aflatoxins: characteristics and impact on human health". In: *Postępy higieny i medycyny doświadczalnej* 71.0 (May 2017), pp. 315–327. URL: <https://phmd.pl/resources/html/article/details?id=152471&language=en>.
- [2] Mari Eskola et al. "Worldwide contamination of food-crops with mycotoxins: Validity of the widely cited 'FAO estimate' of 25%". In: *Critical reviews in food science and nutrition* 60.16 (2020), pp. 2773–2789. URL: <https://doi.org/10.1080/10408398.2019.1658570>.
- [3] John Mullins. "Aspergillus and Aerobiology". In: *The Genus Aspergillus: From Taxonomy and Genetics to Industrial Application*. Ed. by Keith A. Powell, Annabel Renwick, and John F. Peberdy. Boston, MA: Springer US, 1994, pp. 351–359. ISBN: 978-1-4899-0981-7. URL: [https://doi.org/10.1007/978-1-4899-0981-7\\_27](https://doi.org/10.1007/978-1-4899-0981-7_27).
- [4] Zawn Villines and Deborah Weatherspoon. *What to know about Candida die-off*. Feb. 2020. URL: <https://www.medicalnewstoday.com/articles/candida-die-off>.
- [5] Charisse Petersen and June L. Round. "Defining dysbiosis and its influence on host immunity and disease". In: *Cellular Microbiology* 16.7 (2014), pp. 1024–1033. URL: <https://doi.org/10.1111/cmi.12308>.
- [6] MedlinePlus Genetics. *Familial candidiasis*. Sept. 2016. URL: <https://medlineplus.gov/genetics/condition/familial-candidiasis/>.

- [7] Eija Pirinen et al. "Niacin Cures Systemic NAD<sup>+</sup> Deficiency and Improves Muscle Performance in Adult-Onset Mitochondrial Myopathy". In: *Cell Metabolism* 31.6 (2020), 1078–1090.e5. URL: <https://doi.org/10.1016/j.cmet.2020.04.008>.
- [8] Anu Kauppinen et al. "Antagonistic crosstalk between NF- $\kappa$ B and SIRT1 in the regulation of inflammation and metabolic disorders". In: *Cellular Signalling* 25.10 (2013), pp. 1939–1948. URL: <https://doi.org/10.1016/j.cellsig.2013.06.007>.
- [9] Cornelius J. Clancy and M. Hong Nguyen. "Diagnosing Invasive Candidiasis". In: *Journal of Clinical Microbiology* 56.5 (2018), e01909–17. URL: <https://doi.org/10.1128/JCM.01909-17>.
- [10] Paul J. Converse and Ada Hamosh. *INTERLEUKIN 17 RECEPTOR C; IL17RC*. Sept. 2020. URL: <https://www.omim.org/entry/610925>.
- [11] Ada Hamosh. *CANDIDIASIS, FAMILIAL, 6; CANDF6*. June 2015. URL: <https://www.omim.org/entry/613956>.
- [12] Cassandra L. Kniffin. *CANDIDIASIS, FAMILIAL, 9; CANDF9*. July 2015. URL: <https://www.omim.org/entry/616445>.
- [13] Victor A. McKusick and Marla J. F. O'Neill. *CANDIDIASIS, FAMILIAL, 1; CANDF1*. Dec. 2016. URL: <https://www.omim.org/entry/114580>.
- [14] Victor A. McKusick and Marla J. F. O'Neill. *AUTOIMMUNE POLYENDOCRINE SYNDROME, TYPE I, WITH OR WITHOUT REVERSIBLE METAPHYSEAL DYSPLASIA; APS1*. July 2022. URL: <https://www.omim.org/entry/240300>.
- [15] Marla J. F. O'Neill. *CANDIDIASIS, FAMILIAL, 4; CANDF4*. Dec. 2016. URL: <https://www.omim.org/entry/613108>.
- [16] Marla J. F. O'Neill. *CANDIDIASIS, FAMILIAL, 8; CANDF8*. Nov. 2013. URL: <https://www.omim.org/entry/615527>.
- [17] Victor A. McKusick and Marla J. F. O'Neill. *CANDIDIASIS, FAMILIAL, 3; CANDF3*. June 2011. URL: <https://www.omim.org/entry/607644>.
- [18] Patricia A. Hartz and Paul J. Converse. *TRAF3-INTERACTING PROTEIN 2; TRAF3IP2*. Mar. 2015. URL: <https://www.omim.org/entry/607043>.
- [19] Genetic Testing Registry. *Familial chronic mucocutaneous candidiasis*. URL: <https://www.ncbi.nlm.nih.gov/gtr/conditions/C0341024/>.
- [20] Wei Luo et al. "Pulmonary sequestration with Aspergillus infection presenting as massive hemoptysis and hemothorax with highly elevated carcinoembryonic antigen in pleural effusion that mimics advanced lung malignancy". In: *European Journal of Medical Research* 26.1 (2021), p. 48. URL: <https://doi.org/10.1186/s40001-021-00519-5>.
- [21] Sheng Li et al. "One deep learning local-global model based on CT imaging to differentiate between nodular cryptococcosis and lung cancer which are hard to be diagnosed". In: *Computerized Medical Imaging and Graphics* 94 (2021), p. 102009. URL: <https://doi.org/10.1016/j.compmedimag.2021.102009>.
- [22] Ting Yu Wu et al. "Prevalence of Aspergillus-Derived Mycotoxins (Ochratoxin, Aflatoxin, and Gliotoxin) and Their Distribution in the Urinalysis of ME/CFS Patients". In: *International Journal of Environmental Research and Public Health* 19.4 (2022), p. 2052. URL: <https://doi.org/10.3390/ijerph19042052>.
- [23] Alessandra V. Jager et al. "Assessment of aflatoxin exposure using serum and urinary biomarkers in São Paulo, Brazil: A pilot study". In: *International Journal of Hygiene and Environmental Health* 219.3 (2016), pp. 294–300. URL: <https://doi.org/10.1016/j.ijheh.2015.12.003>.
- [24] Muhammed Ershad, Abdullah Naji, and David Vearrier. *N Acetylcysteine*. StatPearls [Internet]. Feb. 2023. URL: <https://www.ncbi.nlm.nih.gov/books/NBK537183/>.
- [25] Brandán Pedre et al. "The mechanism of action of N-acetylcysteine (NAC): The emerging role of H<sub>2</sub>S and sulfane sulfur species". In: *Pharmacology & Therapeutics* 228 (2021), p. 107916. URL: <https://doi.org/10.1016/j.pharmthera.2021.107916>.
- [26] Hassan Gourama and Lloyd B. Bullerman. "Aspergillus flavus and Aspergillus parasiticus: Aflatoxigenic Fungi of Concern in Foods and Feedst: A Review". In: *Journal of Food Protection* 58.12 (1995), pp. 1395–1404. URL: <https://doi.org/10.4315/0362-028X-58.12.1395>.
- [27] Debasish Kumar Dey and Sun Chul Kang. "Aflatoxin B1 induces reactive oxygen species-dependent caspase-mediated apoptosis in normal human cells, inhibits Allium cepa root cell division, and triggers inflammatory response in zebrafish larvae". In: *Science of The Total Environment* 737 (2020), p. 139704. URL: <https://doi.org/10.1016/j.scitotenv.2020.139704>.

- <https://doi.org/10.1016/j.scitotenv.2020.139704>.
- [28] Boyan Huang et al. "Aflatoxin B1 Induces Neurotoxicity through Reactive Oxygen Species Generation, DNA Damage, Apoptosis, and S-Phase Cell Cycle Arrest". In: *International Journal of Molecular Sciences* 21.18 (2020), p. 6517. URL: <https://doi.org/10.3390/ijms21186517>.
- [29] J. Varga et al. "Ochratoxin production by *Aspergillus* species". In: *Applied and Environmental Microbiology* 62.12 (1996), pp. 4461–4464. URL: <https://doi.org/10.1128/aem.62.12.4461-4464.1996>.
- [30] Hennicke G. Kamp et al. "Ochratoxin A: induction of (oxidative) DNA damage, cytotoxicity and apoptosis in mammalian cell lines and primary cells". In: *Toxicology* 206.3 (2005), pp. 413–425. URL: <https://doi.org/10.1016/j.tox.2004.08.004>.
- [31] Leire Arbillaga et al. "Oxidative DNA damage induced by Ochratoxin A in the HK-2 human kidney cell line: evidence of the relationship with cytotoxicity". In: *Mutagenesis* 22.1 (2007), pp. 35–42. URL: <https://doi.org/10.1093/mutage/ge1049>.
- [32] E. Marttila et al. "Fermentative 2-carbon metabolism produces carcinogenic levels of acetaldehyde in *Candida albicans*". In: *Molecular Oral Microbiology* 28.4 (2013), pp. 281–291. URL: <https://doi.org/10.1111/omi.12024>.
- [33] Tingting Yan and Yan Zhao. "Acetaldehyde induces phosphorylation of dynamin-related protein 1 and mitochondrial dysfunction via elevating intracellular ROS and Ca<sup>2+</sup> levels". In: *Redox Biology* 28 (2020), p. 101381. URL: <https://doi.org/10.1016/j.redox.2019.101381>.
- [34] H el ena Alamil et al. "Genotoxicity of aldehyde mixtures: profile of exocyclic DNA-adducts as a biomarker of exposure to tobacco smoke". In: *Toxicology Letters* 331 (2020), pp. 57–64. URL: <https://doi.org/10.1016/j.toxlet.2020.05.010>.
- [35] Rachel A. Montel et al. "Can gene therapy be used to prevent cancer? Gene therapy for aldehyde dehydrogenase 2 deficiency". In: *Cancer Gene Therapy* 29.7 (2022), pp. 889–896. URL: <https://doi.org/10.1038/s41417-021-00399-1>.
- [36] Katsuhito Kino et al. "Generation, repair and replication of guanine oxidation products". In: *Genes and Environment* 39.1 (2017), p. 21. URL: <https://doi.org/10.1186/s41021-017-0081-0>.
- [37] P. K. Shukla and P. C. Mishra. "Repair of O6-methylguanine to guanine by cysteine in the absence and presence of histidine and by cysteine thiolate anion: a quantum chemical study". In: *Physical Chemistry Chemical Physics* 11.37 (2009), pp. 8191–8202. URL: <https://doi.org/10.1039/B908295F>.
- [38] Ramune Reliene, Elvira Fischer, and Robert H. Schiestl. "Effect of N-Acetyl Cysteine on Oxidative DNA Damage and the Frequency of DNA Deletions in Atm-Deficient Mice". In: *Cancer Research* 64.15 (2004), pp. 5148–5153. URL: <https://doi.org/10.1158/0008-5472.CAN-04-0442>.
- [39] Anja G oder et al. "Lipoic acid inhibits the DNA repair protein O6-methylguanine-DNA methyltransferase (MGMT) and triggers its depletion in colorectal cancer cells with concomitant autophagy induction". In: *Carcinogenesis* 36.8 (2015), pp. 817–831. URL: <https://doi.org/10.1093/carcin/bgv070>.
- [40] Xiehong Liu et al. "N-acetylcysteine alleviates H<sub>2</sub>O<sub>2</sub>-induced damage via regulating the redox status of intracellular antioxidants in H9c2 cells". In: *International Journal of Molecular Medicine* 43.1 (2019), pp. 199–208. URL: <https://doi.org/10.3892/ijmm.2018.3962>.
- [41] A. E. Salinas and M. G. Wong. "Glutathione S-transferases—a review". In: *Current medicinal chemistry* 6.4 (1999), pp. 279–309. URL: <https://pubmed.ncbi.nlm.nih.gov/10101214/>.
- [42] S. Vasdev et al. "N-acetyl cysteine attenuates ethanol induced hypertension in rats". In: *Artery* 21.6 (1995), pp. 312–316. URL: <https://pubmed.ncbi.nlm.nih.gov/8833231/>.
- [43] Anatoly Zhitkovich. "N-Acetylcysteine: Antioxidant, Aldehyde Scavenger, and More". In: *Chemical Research in Toxicology* 32.7 (2019), pp. 1318–1319. URL: <https://doi.org/10.1021/acs.chemrestox.9b00152>.
- [44] Amanda L. Miller et al. "Microbiome or Infections: Amyloid-Containing Biofilms as a Trigger for Complex Human Diseases". In: *Frontiers in Immunology* 12 (2021), p. 638867. URL: <https://doi.org/10.3389/fimmu.2021.638867>.

- [45] Janaina De Cássia Orlandi Sardi et al. "Highlights in pathogenic fungal biofilms". In: *Revista Iberoamericana de Micología* 31.1 (2014), pp. 22–29. URL: <https://doi.org/10.1016/j.riam.2013.09.014>.
- [46] Luis R. Martinez and Bettina C. Fries. "Fungal Biofilms: Relevance in the Setting of Human Disease". In: *Current Fungal Infection Reports* 4.4 (2010), pp. 266–275. URL: <https://doi.org/10.1007/s12281-010-0035-5>.
- [47] Tiemei Zhao and Youning Liu. "N-acetylcysteine inhibit biofilms produced by *Pseudomonas aeruginosa*". In: *BMC Microbiology* 10.1 (2010), p. 140. URL: <https://doi.org/10.1186/1471-2180-10-140>.
- [48] Ji-Hoi Moon et al. "Antibacterial effects of N-acetylcysteine against endodontic pathogens". In: *Journal of Microbiology* 54.4 (2016), pp. 322–329. URL: <https://doi.org/10.1007/s12275-016-5534-9>.
- [49] Rehab Mahmoud Abd El-Baky, Dalia Mohamed Mohamed Abo El Ela, and Gamal Fadl Mamoud Gad. "N-acetylcysteine Inhibits and Eradicates *Candida albicans* Biofilms". In: *American Journal of Infectious Diseases and Microbiology* 2.5 (2014), pp. 122–130. URL: <https://doi.org/10.12691/ajidm-2-5-5>.
- [50] Luis R. Martinez and Arturo Casadevall. "Cryptococcus neoformans Biofilm Formation Depends on Surface Support and Carbon Source and Reduces Fungal Cell Susceptibility to Heat, Cold, and UV Light". In: *Applied and Environmental Microbiology* 73.14 (2007), pp. 4592–4601. URL: <https://doi.org/10.1128/AEM.02506-06>.
- [51] Marcio L. Rodrigues et al. "Vesicular Trans-Cell Wall Transport in Fungi: A Mechanism for the Delivery of Virulence-Associated Macromolecules?" In: *Lipid Insights* 2 (2008), LPI.S1000. URL: <https://doi.org/10.4137/LPI.S1000>.
- [52] Jie Zheng et al. "N-Acetylcysteine interacts with copper to generate hydrogen peroxide and selectively induce cancer cell death". In: *Cancer Letters* 298.2 (2010), pp. 186–194. URL: <https://doi.org/10.1016/j.canlet.2010.07.003>.
- [53] Xin Li et al. "N-Acetyl-cysteine and Mechanisms Involved in Resolution of Chronic Wound Biofilm". In: *Journal of Diabetes Research* 2020 (2020), p. 9589507. URL: <https://doi.org/10.1155/2020/9589507>.
- [54] Mónica Homa et al. "In vitro susceptibility of *Scedosporium* isolates to N-acetyl-L-cysteine alone and in combination with conventional antifungal agents". In: *Medical Mycology* 54.7 (2016), pp. 776–779. URL: <https://doi.org/10.1093/mmy/myw029>.
- [55] Srabani Banerjee and Suzanne McCormack. *Acetylcysteine for Patients Requiring Mucous Secretion Clearance: A Review of Clinical Effectiveness and Safety*. CADTH Rapid Response Reports. Ottawa (ON): Canadian Agency for Drugs and Technologies in Health, June 2019. URL: <https://pubmed.ncbi.nlm.nih.gov/31503431/>.
- [56] M. Zafarullah et al. "Molecular mechanisms of N-acetylcysteine actions". In: *Cellular and Molecular Life Sciences CMLS* 60.1 (2003), pp. 6–20. URL: <https://doi.org/10.1007/s000180300001>.
- [57] Chao Niu et al. "Carvacrol Induces *Candida albicans* Apoptosis Associated With Ca<sup>2+</sup>/Calcineurin Pathway". In: *Frontiers in Cellular and Infection Microbiology* 10 (2020), p. 192. URL: <https://doi.org/10.3389/fcimb.2020.00192>.
- [58] Marianna Halasi et al. "ROS inhibitor N-acetyl-L-cysteine antagonizes the activity of proteasome inhibitors". In: *Biochemical Journal* 454.2 (2013), pp. 201–208. URL: <https://doi.org/10.1042/BJ20130282>.
- [59] Micaely Cristina dos Santos Tenório et al. "N-Acetylcysteine (NAC): Impacts on Human Health". In: *Antioxidants* 10.6 (2021), p. 967. URL: <https://doi.org/10.3390/antiox10060967>.
- [60] Mark Yarema et al. "Anaphylactoid Reactions to Intravenous N-Acetylcysteine during Treatment for Acetaminophen Poisoning". In: *Journal of Medical Toxicology* 14.2 (2018), pp. 120–127. URL: <https://doi.org/10.1007/s13181-018-0653-9>.
- [61] Peter Calverley, Paola Rogliani, and Alberto Papi. "Safety of N-Acetylcysteine at High Doses in Chronic Respiratory Diseases: A Review". In: *Drug Safety* 44.3 (2021), pp. 273–290. URL: <https://doi.org/10.1007/s40264-020-01026-y>.
- [62] A. V. Appelboom, P. I. Dargan, and J. Knighton. "Fatal anaphylactoid reaction to N-acetylcysteine: caution in patients with asthma". In: *Emergency Medicine Journal* 19.6 (2002), pp. 594–595. URL: <https://doi.org/10.1136/emj.19.6.594>.

- [63] E. A. Sandilands and D. N. Bateman. "Adverse reactions associated with acetylcysteine". In: *Clinical Toxicology* 47.2 (2009), pp. 81–88. URL: <https://doi.org/10.1080/15563650802665587>.
- [64] G. Ferrari, C. Y. Yan, and L. A. Greene. "N-acetylcysteine (D- and L-stereoisomers) prevents apoptotic death of neuronal cells". In: *Journal of Neuroscience* 15.4 (1995), pp. 2857–2866. URL: <https://doi.org/10.1523/JNEUROSCI.15-04-02857.1995>.
- [65] Volkan I. Sayin et al. "Antioxidants Accelerate Lung Cancer Progression in Mice". In: *Science Translational Medicine* 6.221 (2014), 221ra15. URL: <https://doi.org/10.1126/scitranslmed.3007653>.
- [66] Mehdi Dastjerdi et al. "Effect of thymoquinone on P53 gene expression and consequence apoptosis in breast cancer cell line". In: *International Journal of Preventive Medicine* 7.1 (2016), p. 66. URL: <https://doi.org/10.4103/2008-7802.180412>.
- [67] Rafaela de Oliveira Nóbrega et al. "Investigation of the antifungal activity of carvacrol against strains of *Cryptococcus neoformans*". In: *Pharmaceutical Biology* 54.11 (2016), pp. 2591–2596. URL: <https://doi.org/10.3109/13880209.2016.1172319>.
- [68] A. Ahmad et al. "Fungicidal activity of thymol and carvacrol by disrupting ergosterol biosynthesis and membrane integrity against *Candida*". In: *European Journal of Clinical Microbiology & Infectious Diseases* 30.1 (2011), pp. 41–50. URL: <https://doi.org/10.1007/s10096-010-1050-8>.
- [69] National Center for Biotechnology Information. *PubChem Compound Summary for CID 10364, Carvacrol*. Apr. 2023. URL: <https://pubchem.ncbi.nlm.nih.gov/compound/Carvacrol>.
- [70] Iliyan D. Iliiev and Irina Leonardi. "Fungal dysbiosis: immunity and interactions at mucosal barriers". In: *Nature Reviews Immunology* 17.10 (2017), pp. 635–646. URL: <https://doi.org/10.1038/nri.2017.55>.
- [71] Mohammad Y. Memar et al. "Carvacrol and thymol: strong antimicrobial agents against resistant isolates". In: *Reviews and Research in Medical Microbiology* 28.2 (2017), pp. 63–68. URL: <https://doi.org/10.1097/MRM.000000000000100>.
- [72] Ilham Abass Bnyan, Aumaima Tariq Abid, and Hamid Naji Obied. "Antibacterial Activity of Carvacrol against Different Types of Bacteria". In: *Journal of Natural Sciences Research* 4.9 (2014), pp. 13–16. URL: <https://www.iiste.org/Journals/index.php/JNSR/article/viewFile/13191/13559>.
- [73] Hicham Ferhout et al. "Antifungal Activity of Selected Essential Oils, Cinnamaldehyde and Carvacrol against *Malassezia furfur* and *Candida albicans*". In: *Journal of Essential Oil Research* 11.1 (1999), pp. 119–129. URL: <https://doi.org/10.1080/10412905.1999.9701086>.
- [74] Shankumar Mooyottu et al. "Protective Effect of Carvacrol against Gut Dysbiosis and *Clostridium difficile* Associated Disease in a Mouse Model". In: *Frontiers in Microbiology* 8 (2017), p. 625. URL: <https://doi.org/10.3389/fmicb.2017.00625>.
- [75] Yi Zou et al. "Oregano Essential Oil Improves Intestinal Morphology and Expression of Tight Junction Proteins Associated with Modulation of Selected Intestinal Bacteria and Immune Status in a Pig Model". In: *BioMed Research International* 2016 (2016), p. 5436738. URL: <https://doi.org/10.1155/2016/5436738>.
- [76] Chenyang Li et al. "The effects of oregano essential oil on production performance and intestinal barrier function in growing Hyla rabbits". In: *Italian Journal of Animal Science* 20.1 (2021), pp. 2165–2173. URL: <https://doi.org/10.1080/1828051X.2021.2005471>.
- [77] Carmel Kelly et al. "The In Vitro and In Vivo Effect of Carvacrol in Preventing *Campylobacter* Infection, Colonization and in Improving Productivity of Chicken Broilers". In: *Foodborne Pathogens and Disease* 14.6 (2017), pp. 341–349. URL: <https://doi.org/10.1089/fpd.2016.2265>.
- [78] Maria Pia Franciosini et al. "Effects of oregano (*Origanum vulgare* L.) and rosemary (*Rosmarinus officinalis* L.) aqueous extracts on broiler performance, immune function and intestinal microbial population". In: *Journal of Applied Animal Research* 44.1 (2016), pp. 474–479. URL: <https://doi.org/10.1080/09712119.2015.1091322>.
- [79] Yusuke Kinashi and Koji Hase. "Partners in Leaky Gut Syndrome: Intestinal Dysbiosis and Autoimmunity". In: *Frontiers in Immunology* 12 (2021), p. 673708. URL: <https://doi.org/10.3389/fimmu.2021.673708>.

- [80] Patrícia Pimentel de Barros et al. "Candida Biofilms: An Update on Developmental Mechanisms and Therapeutic Challenges". In: *Mycopathologia* 185.3 (2020), pp. 415–424. URL: <https://doi.org/10.1007/s11046-020-00445-w>.
- [81] Sivan Elias and Ehud Banin. "Multi-species biofilms: living with friendly neighbors". In: *FEMS Microbiology Reviews* 36.5 (2012), pp. 990–1004. URL: <https://doi.org/10.1111/j.1574-6976.2012.00325.x>.
- [82] Roberto Scaffaro et al. "Efficacy of poly(lactic acid)/carvacrol electrospun membranes against *Staphylococcus aureus* and *Candida albicans* in single and mixed cultures". In: *Applied Microbiology and Biotechnology* 102.9 (2018), pp. 4171–4181. URL: <https://doi.org/10.1007/s00253-018-8879-7>.
- [83] Thirukannamangai Krishnan Swetha et al. "Synergistic antimicrobial combination of carvacrol and thymol impairs single and mixed-species biofilms of *Candida albicans* and *Staphylococcus epidermidis*". In: *Biofouling* 36.10 (2020), pp. 1256–1271. URL: <https://doi.org/10.1080/08927014.2020.1869949>.
- [84] Ivana Čabarkapa et al. "Anti-biofilm activities of essential oils rich in carvacrol and thymol against *Salmonella Enteritidis*". In: *Biofouling* 35.3 (2019), pp. 361–375. URL: <https://doi.org/10.1080/08927014.2019.1610169>.
- [85] Katherine Miranda-Cadena et al. "In vitro activities of carvacrol, cinnamaldehyde and thymol against *Candida* biofilms". In: *Biomedicine & Pharmacotherapy* 143 (2021), p. 112218. URL: <https://doi.org/10.1016/j.biopha.2021.112218>.
- [86] Poonam Kumari et al. "Antifungal and Anti-Biofilm Activity of Essential Oil Active Components against *Cryptococcus neoformans* and *Cryptococcus laurentii*". In: *Frontiers in Microbiology* 8 (2017), p. 2161. URL: <https://doi.org/10.3389/fmicb.2017.02161>.
- [87] Daliah Alves Coelho Trevisan et al. "Action of carvacrol in *Salmonella Typhimurium* biofilm: A proteomic study". In: *Journal of Applied Biomedicine* 18.4 (2020), pp. 106–114. URL: <https://doi.org/10.32725/jab.2020.014>.
- [88] Muhammad Imran et al. "Therapeutic application of carvacrol: A comprehensive review". In: *Food Science & Nutrition* 10.11 (2022), pp. 3544–3561. URL: <https://doi.org/10.1002/fsn3.2994>.
- [89] Daiane Einhardt Blank et al. "Bioactive Compounds and Antifungal Activities of Extracts of Lamiaceae Species". In: *Journal of Agricultural Chemistry and Environment* 9.3 (2020), pp. 85–96. URL: <https://doi.org/10.4236/jacen.2020.93008>.
- [90] Priscilla de Almeida et al. "Antioxidant and antifungal properties of essential oils of oregano (*Origanum vulgare*) and mint (*Mentha arvensis*) against *Aspergillus flavus* and *Penicillium commune* for use in food preservation". In: *Food Science and Technology* 42 (2022), e64921. URL: <https://doi.org/10.1590/fst.64921>.
- [91] M. Viuda-Martos et al. "Antifungal activities of thyme, clove and oregano essential oils". In: *Journal of Food Safety* 27.1 (2007), pp. 91–101. URL: <https://doi.org/10.1111/j.1745-4565.2007.00063.x>.
- [92] Tuncay Gumus et al. "Inhibition of heat resistant molds: *Aspergillus fumigatus* and *Paecilomyces variotii* by some plant essential oils". In: *Food Science and Biotechnology* 19.5 (2010), pp. 1241–1244. URL: <https://doi.org/10.1007/s10068-010-0177-9>.
- [93] Sunčica Kocić-Tanackov et al. "The inhibitory effect of oregano extract on the growth of *Aspergillus* spp. and on sterigmatocystin biosynthesis". In: *LWT* 49.1 (2012), pp. 14–20. URL: <https://doi.org/10.1016/j.lwt.2012.04.013>.
- [94] Özge Özdal Zincir et al. "Synergistic effect of thymoquinone and nystatin in the treatment of oral candidiasis; an in vitro study". In: *Odontology* 110.2 (2022), pp. 330–337. URL: <https://doi.org/10.1007/s10266-021-00667-4>.
- [95] Ayed A. Dera et al. "Synergistic efficacies of thymoquinone and standard antibiotics against multi-drug resistant isolates". In: *Saudi Medical Journal* 42.2 (2021), pp. 196–204. URL: <https://doi.org/10.15537/smj.2021.2.25706>.
- [96] Masood Alam Khan et al. "Liposomal thymoquinone effectively combats fluconazole-resistant *Candida albicans* in a murine model". In: *International Journal of Biological Macromolecules* 76 (2015), pp. 203–208. URL: <https://doi.org/10.1016/j.ijbiomac.2015.02.015>.
- [97] Bahman Nickavar et al. "Chemical Composition of the Fixed and Volatile Oils of *Nigella sativa* L. from Iran". In: *Zeitschrift für Naturforschung C* 58.9-10 (2003), pp. 629–631. URL:



- <https://doi.org/10.1515/znc-2003-9-1004>.
- [98] Gabriella Kiskó and Sibel Roller. "Carvacrol and p-cymene inactivate Escherichia coli O157:H7 in apple juice". In: *BMC Microbiology* 5.1 (2005), p. 36. URL: <https://doi.org/10.1186/1471-2180-5-36>.
- [99] Rita de Cássia Viana de Carvalho et al. "Limonene-carvacrol: A combination of monoterpenes with enhanced antileishmanial activity". In: *Toxicology in Vitro* 74 (2021), p. 105158. URL: <https://doi.org/10.1016/j.tiv.2021.105158>.
- [100] Thaís Soares Bezerra Santos Nunes et al. "Fungistatic Action of N-Acetylcysteine on Candida albicans Biofilms and Its Interaction with Antifungal Agents". In: *Microorganisms* 8.7 (2020), p. 980. URL: <https://doi.org/10.3390/microorganisms8070980>.
- [101] Dejie Liang et al. "Thymol Inhibits LPS-Stimulated Inflammatory Response via Down-Regulation of NF- $\kappa$ B and MAPK Signaling Pathways in Mouse Mammary Epithelial Cells". In: *Inflammation* 37.1 (2014), pp. 214–222. URL: <https://doi.org/10.1007/s10753-013-9732-x>.
- [102] Milena da Silva Lima et al. "Anti-inflammatory effects of carvacrol: Evidence for a key role of interleukin-10". In: *European Journal of Pharmacology* 699.1 (2013), pp. 112–117. URL: <https://doi.org/10.1016/j.ejphar.2012.11.040>.
- [103] Premysl Landa et al. "In vitro anti-inflammatory activity of carvacrol: Inhibitory effect on COX-2 catalyzed prostaglandin E2 biosynthesis". In: *Archives of Pharmacal Research* 32.1 (2009), pp. 75–78. URL: <https://doi.org/10.1007/s12272-009-1120-6>.
- [104] Francilene V. Silva et al. "Anti-Inflammatory and Anti-Ulcer Activities of Carvacrol, a Monoterpene Present in the Essential Oil of Oregano". In: *Journal of Medicinal Food* 15.11 (2012), pp. 984–991. URL: <https://doi.org/10.1089/jmf.2012.0102>.
- [105] Xuesheng Han and Tory L. Parker. "Anti-inflammatory, tissue remodeling, immunomodulatory, and anticancer activities of oregano (*Origanum vulgare*) essential oil in a human skin disease model". In: *Biochimie Open* 4 (2017), pp. 73–77. URL: <https://doi.org/10.1016/j.biopen.2017.02.005>.
- [106] Vahideh Ghorani et al. "Safety and tolerability of carvacrol in healthy subjects: a phase I clinical study". In: *Drug and Chemical Toxicology* 44.2 (2021), pp. 177–189. URL: <https://doi.org/10.1080/01480545.2018.1538233>.
- [107] Azam Alavinezhad, Mohammad Reza Khazdair, and Mohammad Hossein Boskabady. "Possible therapeutic effect of carvacrol on asthmatic patients: A randomized, double blind, placebo-controlled, Phase II clinical trial". In: *Phytotherapy Research* 32.1 (2018), pp. 151–159. URL: <https://doi.org/10.1002/ptr.5967>.
- [108] María Llana-Ruiz-Cabello et al. "Genotoxicity evaluation of carvacrol in rats using a combined micronucleus and comet assay". In: *Food and Chemical Toxicology* 98 (2016), pp. 240–250. URL: <https://doi.org/10.1016/j.fct.2016.11.005>.
- [109] Fatiha El Babili et al. "Oregano: Chemical Analysis and Evaluation of Its Antimalarial, Antioxidant, and Cytotoxic Activities". In: *Journal of Food Science* 76.3 (2011), pp. C512–C518. URL: <https://doi.org/10.1111/j.1750-3841.2011.02109.x>.
- [110] María Llana-Ruiz-Cabello et al. "Use of micronucleus and comet assay to evaluate evaluate the genotoxicity of oregano essential oil (*Origanum vulgare* L. Virens) in rats orally exposed for 90 days." In: *Journal of Toxicology and Environmental Health, Part A* 81.12 (2018), pp. 525–533. URL: <https://doi.org/10.1080/15287394.2018.1447522>.
- [111] Fatima Bouhtit et al. "New Anti-Leukemic Effect of Carvacrol and Thymol Combination through Synergistic Induction of Different Cell Death Pathways". In: *Molecules* 26.2 (2021), p. 410. URL: <https://doi.org/10.3390/molecules26020410>.
- [112] T. A. Misharina et al. "Effect of oregano essential oil on the engraftment and development of Lewis carcinoma in F1 DBA C57 black hybrid mice". In: *Applied Biochemistry and Microbiology* 49.4 (2013), pp. 432–436. URL: <https://doi.org/10.1134/S0003683813040091>.
- [113] Alireza Tavakkoli et al. "Review on Clinical Trials of Black Seed (*Nigella sativa*) and Its Active Constituent, Thymoquinone." In: *Journal of pharmacopuncture* 20.3 (2017), pp. 179–193. URL: <https://doi.org/10.3831/KPI.2017.20.021>.

- [114] Saptha Jyothi Gerige et al. "GC-MS analysis of *Nigella sativa* seeds and antimicrobial activity of its volatile oil". In: *Brazilian Archives of Biology and Technology* 52.5 (2009), pp. 1189–1192. URL: <https://doi.org/10.1590/S1516-89132009000500016>.
- [115] Osaretin Albert Taiwo Ebuehi et al. "Nigella sativa (black seed) oil ameliorates CCl<sub>4</sub>-induced hepatotoxicity and mediates neurotransmitter levels in male Sprague Dawley albino rats". In: *Journal of Food Biochemistry* 44.2 (2020), e13108. URL: <https://doi.org/10.1111/jfbc.13108>.
- [116] Hina (Ed.) Younus. *Molecular and Therapeutic actions of Thymoquinone*. Singapore: Springer Singapore, Apr. 2018. URL: <https://doi.org/10.1007/978-981-10-8800-1>.
- [117] Opeyemi Oluwafemi Ojueromi, Ganiyu Obob, and Ayokunle Olubode Ademosun. "Effect of black seeds (*Nigella sativa*) on inflammatory and immunomodulatory markers in Plasmodium berghei-infected mice". In: *Journal of Food Biochemistry* 46.11 (2022), e14300. URL: <https://doi.org/10.1111/jfbc.14300>.
- [118] Gamal Badr et al. "Perinatal supplementation with thymoquinone improves diabetic complications and T cell immune responses in rat offspring". In: *Cellular Immunology* 267.2 (2011), pp. 133–140. URL: <https://doi.org/10.1016/j.cellimm.2011.01.002>.
- [119] Mahmoud N. Nagi and Mahmoud A. Mansour. "Protective effect of thymoquinone against doxorubicin-induced cardiotoxicity in rats: A possible mechanism of protection". In: *Pharmacological Research* 41.3 (2000), pp. 283–289. URL: <https://doi.org/10.1006/phrs.1999.0585>.
- [120] Mahmoud N. Nagi et al. "Thymoquinone protects against carbon tetrachloride hepatotoxicity in mice via an antioxidant mechanism". In: *IUBMB Life* 47.1 (1999), pp. 153–159. URL: <https://doi.org/10.1080/15216549900201153>.
- [121] M. Burits and F. Bucar. "Antioxidant activity of *Nigella sativa* essential oil". In: *Phytotherapy Research* 14.5 (2000), pp. 323–328. URL: [https://doi.org/10.1002/1099-1573\(200008\)14:5%3C323::aid-pt621%3E3.0.co;2-q](https://doi.org/10.1002/1099-1573(200008)14:5%3C323::aid-pt621%3E3.0.co;2-q).
- [122] Ferah Armutcu, Sumeyya Akyol, and Omer Akyol. "The interaction of glutathione and thymoquinone and their antioxidant properties". In: *Electronic Journal of General Medicine* 15.4 (2018), em59. URL: <https://doi.org/10.29333/ejgm/89493>.
- [123] Osama A. Badary et al. "The influence of thymoquinone on doxorubicin-induced hyperlipidemic nephropathy in rats". In: *Toxicology* 143.3 (2000), pp. 219–226. URL: [https://doi.org/10.1016/S0300-483X\(99\)00179-1](https://doi.org/10.1016/S0300-483X(99)00179-1).
- [124] Kamal A. Qureshi et al. "In Vitro and In Silico Approaches for the Evaluation of Antimicrobial Activity, Time-Kill Kinetics, and Anti-Biofilm Potential of Thymoquinone (2-Methyl-5-propan-2-ylcyclohexa-2,5-diene-1,4-dione) against Selected Human Pathogens". In: *Antibiotics* 11.1 (2022), p. 79. URL: <https://doi.org/10.3390/antibiotics11010079>.
- [125] Kamel Chaieb et al. "Antibacterial activity of Thymoquinone, an active principle of *Nigella sativa* and its potency to prevent bacterial biofilm formation". In: *BMC Complementary and Alternative Medicine* 11.1 (2011), p. 29. URL: <https://doi.org/10.1186/1472-6882-11-29>.
- [126] Fatima A. Saleh et al. "Phytochemical Analysis of *Nigella sativa* L. Utilizing GC-MS Exploring its Antimicrobial Effects against Multidrug-Resistant Bacteria". In: *Pharmacognosy Journal* 10.1 (2018), pp. 99–105. URL: <https://doi.org/10.5530/pj.2018.1.18>.
- [127] Ramar Vanajothi et al. "In silico and In vitro Analysis of *Nigella sativa* Bioactives Against Chorismate Synthase of *Listeria monocytogenes*: a Target Protein for Biofilm Inhibition". In: *Applied Biochemistry and Biotechnology* 195.1 (2023), pp. 519–533. URL: <https://doi.org/10.1007/s12010-022-04157-3>.
- [128] Hanene Miladi et al. "Synergistic effect of eugenol, carvacrol, thymol, p-cymene and  $\gamma$ -terpinene on inhibition of drug resistance and biofilm formation of oral bacteria". In: *Microbial Pathogenesis* 112 (2017), pp. 156–163. URL: <https://doi.org/10.1016/j.micpath.2017.09.057>.
- [129] Archana Thakre et al. "Limonene inhibits *Candida albicans* growth by inducing apoptosis". In: *Medical Mycology* 56.5 (2017), pp. 565–578. URL: <https://academic.oup.com/mmy/article/56/5/565/4372441>.
- [130] Paweł Kwiatkowski et al. "The Effect of Subinhibitory Concentrations of trans-Anethole on Antibacterial and Antibiofilm Activity of Mupirocin Against Mupirocin-Resistant *Staphylococcus aureus* Strains". In: *Microbial Drug Resistance* 25.10 (2019), pp. 1424–1429. URL: <https://doi.org/10.1089/mdr.2019.0101>.

- [131] A. Bitá et al. "An alternative treatment for Candida infections with *Nigella sativa* extracts". In: *BMJ Specialist Journals* 19.2 (2012), p. 162. URL: <https://doi.org/10.1136/ejhp-2012-000074>. 203.
- [132] M. A. U. Khan et al. "The in vivo antifungal activity of the aqueous extract from *Nigella sativa* seeds". In: *Phytotherapy Research* 17.2 (2003), pp. 183–186. URL: <https://doi.org/10.1002/ptr.1146>.
- [133] Saifeldin Ahmed Faragalla El-Nagerabi et al. "Effect of *Hibiscus sabdariffa* extract and *Nigella sativa* oil on the growth and aflatoxin B1 production of *Aspergillus flavus* and *Aspergillus parasiticus* strains". In: *Food Control* 25.1 (2012), pp. 59–63. URL: <https://doi.org/10.1016/j.foodcont.2011.09.033>.
- [134] M. Taha, A. Azeiz, and W. Saudi. "Antifungal effect of thymol, thymoquinone and thymohydroquinone against yeasts, dermatophytes and non-dermatophyte molds isolated from skin and nails fungal infections". In: *Egyptian Journal of Biochemistry and Molecular Biology* 28.2 (2010). URL: <https://doi.org/10.4314/ejbmb.v28i2.60802>.
- [135] Naeem Akhtar et al. "Comparison of Antifungal Activity of Thymoquinone and Amphotericin B Against *Fusarium solani* in-vitro". In: *Scientific Journal of King Faisal University* 8.2 (2007), pp. 137–145. URL: [https://www.academia.edu/63461871/Comparison\\_of\\_Antifungal\\_Activity\\_of\\_Thymoquinone\\_and\\_Amphotericin\\_B\\_Against\\_Fusarium\\_solani\\_in\\_vitro](https://www.academia.edu/63461871/Comparison_of_Antifungal_Activity_of_Thymoquinone_and_Amphotericin_B_Against_Fusarium_solani_in_vitro).
- [136] Fengli Chen et al. "Insight into the essential oil isolation from *Foeniculum vulgare* Mill. fruits using double-condensed microwave-assisted hydrodistillation and evaluation of its antioxidant, antifungal and cytotoxic activity". In: *Industrial Crops and Products* 144 (2020), p. 112052. URL: <https://doi.org/10.1016/j.indcrop.2019.112052>.
- [137] Y. Tsukuda et al. "Structure–activity relationships of antifungal phenylpropanoid derivatives and their synergy with n-dodecanol and fluconazole". In: *Letters in Applied Microbiology* 74.3 (2022), pp. 377–384. URL: <https://doi.org/10.1111/lam.13613>.
- [138] Michelle Liz de Souza Pessoa et al. "Antifungal activity and anti-diarrheal activity via antimotility mechanisms of (-)-fenchone in experimental models". In: *World Journal of Gastroenterology* 26.43 (2020), pp. 6795–6809. URL: <https://doi.org/10.3748/wjg.v26.i43.6795>.
- [139] Hao Yu et al. "Antifungal activity and mechanism of d-limonene against foodborne opportunistic pathogen *Candida tropicalis*". In: *LWT* 159 (2022), p. 113144. URL: <https://doi.org/10.1016/j.lwt.2022.113144>.
- [140] Nasser Vahdati-Mashhadian and Hassan Rakhshandeh. "Antibacterial and antifungal effects of *Nigella sativa* extracts against *S. aureus*, *P. aeruginosa* and *C. albicans*". In: *Pakistan Journal of Medical Sciences* 21.1 (2005), pp. 47–52. URL: [https://www.researchgate.net/publication/279551703\\_Antibacterial\\_and\\_antifungal\\_effects\\_of\\_Nigella\\_sativa\\_extract\\_against\\_S\\_aureus\\_P\\_aeruginosa\\_and\\_C\\_albicans](https://www.researchgate.net/publication/279551703_Antibacterial_and_antifungal_effects_of_Nigella_sativa_extract_against_S_aureus_P_aeruginosa_and_C_albicans).
- [141] Cristina Rodrigues dos Santos Barbosa et al. "Effect of Carvacrol and Thymol on NorA efflux pump inhibition in multidrug-resistant (MDR) *Staphylococcus aureus* strains". In: *Journal of Bioenergetics and Biomembranes* 53.4 (2021), pp. 489–498. URL: <https://doi.org/10.1007/s10863-021-09906-3>.
- [142] Gökalp İşcan, Arzu İşcan, and Fatih Demirci. "Anticandidal Effects of Thymoquinone: Mode of Action Determined by Transmission". In: *Natural product communications* 11.7 (2016), pp. 977–978. URL: <https://pubmed.ncbi.nlm.nih.gov/30452175/>.
- [143] Hala Almshawit and Ian Macreadie. "Fungicidal effect of thymoquinone involves generation of oxidative stress in *Candida glabrata*". In: *Microbiological Research* 195 (2017), pp. 81–88. URL: <https://doi.org/10.1016/j.micres.2016.11.008>.
- [144] Anahit Penesyan et al. "Three faces of biofilms: a microbial lifestyle, a nascent multicellular organism, and an incubator for diversity". In: *npj Biofilms and Microbiomes* 7.1 (2021), p. 80. URL: <https://doi.org/10.1038/s41522-021-00251-2>.
- [145] Sadaf Dabeer et al. "Chapter 1 - History and traditional uses of black seeds (*Nigella sativa*)". In: *Black Seeds (Nigella Sativa)*. Ed. by Andleeb Khan and Muneeb Rehman. Elsevier, 2022, pp. 1–28. ISBN: 978-0-12-824462-3. URL: <https://doi.org/10.1016/B978-0-12-824462-3.00016-0>.

- [146] Krishnapura Srinivasan. "Cumin (*Cuminum cyminum*) and black cumin (*Nigella sativa*) seeds: traditional uses, chemical constituents, and nutraceutical effects". In: *Food Quality and Safety* 2.1 (2018), pp. 1–16. URL: <https://doi.org/10.1093/fqsafe/fyx031>.
- [147] Mohammad-Foad Noorbakhsh et al. "An Overview of Hepatoprotective Effects of Thymoquinone". In: *Recent Patents on Food, Nutrition & Agriculture* 9.1 (2018), pp. 14–22. URL: <https://doi.org/10.2174/2212798410666180221105503>.
- [148] Heena Tabassum, Asad Ahmad, and Iffat Z. Ahmad. "Nigella sativa L. and Its Bioactive Constituents as Hepatoprotectant: A Review". In: *Current Pharmaceutical Biotechnology* 19.1 (2018), pp. 43–67. URL: <https://doi.org/10.2174/1389201019666180427110007>.
- [149] Ben-Wen Cui et al. "Thymoquinone Attenuates Acetaminophen Overdose-Induced Acute Liver Injury and Inflammation Via Regulation of JNK and AMPK Signaling Pathway". In: *The American Journal of Chinese Medicine* 47.03 (2019), pp. 577–594. URL: <https://doi.org/10.1142/S0192415X19500307>.
- [150] Mohammad Rashidmayvan et al. "The effect of Nigella sativa oil on serum levels of inflammatory markers, liver enzymes, lipid profile, insulin and fasting blood sugar in patients with non-alcoholic fatty liver". In: *Journal of Diabetes & Metabolic Disorders* 18.2 (2019), pp. 453–459. URL: <https://doi.org/10.1007/s40200-019-00439-6>.
- [151] Mina Darand et al. "Nigella sativa and inflammatory biomarkers in patients with non-alcoholic fatty liver disease: Results from a randomized, double-blind, placebo-controlled, clinical trial". In: *Complementary Therapies in Medicine* 44 (2019), pp. 204–209. URL: <https://doi.org/10.1016/j.ctim.2019.04.014>.
- [152] Sheik Noor Mohamed and John Wyson. "In vitro immunostimulation activity of Nigella sativa Linn. And psoralea Corylifolia Linn. seeds using a murine macrophage cell line". In: 2017. URL: <https://www.semanticscholar.org/paper/IN-VITRO-IMMUNOSTIMULATION-ACTIVITY-OF-NIGELLA-AND-Mohamed-Wyson/114f14cc437d06e59b6dd8677aee6c179451afff>.
- [153] Ashraf S. Hakim et al. "Assessment of immunomodulatory effects of black cumin seed (*Nigella sativa*) extract on macrophage activity in vitro". In: *International Journal of Veterinary Science* 8.4 (2019), pp. 385–389. URL: <http://www.ijvets.com/pdf-files/Volume-8-no-4-2019/385-389.pdf>.
- [154] Ali A. Alshatwi. "Bioactivity-guided identification to delineate the immunomodulatory effects of methanolic extract of Nigella sativa seed on human peripheral blood mononuclear cells". In: *Chinese Journal of Integrative Medicine* (2014), pp. 1–6. URL: <https://doi.org/10.1007/s11655-013-1534-3>.
- [155] Yun Niu et al. "Nigella sativa: A Dietary Supplement as an Immune-Modulator on the Basis of Bioactive Components". In: *Frontiers in Nutrition* 8 (2021), p. 722813. URL: <https://doi.org/10.3389/fnut.2021.722813>.
- [156] M. L. Salem, F. Q. Alenzi, and W. Y. Attia. "Thymoquinone, the active ingredient of Nigella sativa seeds, enhances survival and activity of antigen-specific CD8-positive T cells in vitro". In: *British Journal of Biomedical Science* 68.3 (2011), pp. 131–137. URL: <https://doi.org/10.1080/09674845.2011.11730340>.
- [157] Sorayya Kheirouri, Vahid Hadi, and Mohammad Alizadeh. "Immunomodulatory Effect of Nigella sativa Oil on T Lymphocytes in Patients with Rheumatoid Arthritis". In: *Immunological Investigations* 45.4 (2016), pp. 271–283. URL: <https://doi.org/10.3109/08820139.2016.1153649>.
- [158] Faaiza Shahid et al. "Oral Nigella sativa oil and thymoquinone administration ameliorates the effect of long-term cisplatin treatment on the enzymes of carbohydrate metabolism, brush border membrane, and antioxidant defense in rat intestine". In: *Naunyn-Schmiedeberg's Archives of Pharmacology* 391.2 (2018), pp. 145–157. URL: <https://doi.org/10.1007/s00210-017-1444-6>.
- [159] Saeed Samarghandian, Farkhondeh Tahereh, and Samini Fariborz. "A Review on Possible Therapeutic Effect of Nigella sativa and Thymoquinone in Neurodegenerative Diseases". In: *CNS & Neurological Disorders - Drug Targets* 17.6 (2018), pp. 412–420. URL: <https://doi.org/10.2174/1871527317666180702101455>.
- [160] Yasmina K. Mahmoud and Heba Mohammed Ahmed Abdelrazek. "Cancer: Thymoquinone antioxidant/pro-oxidant effect as potential anticancer remedy". In: *Biomedicine & Pharmacotherapy* 115 (2019), p. 108783. URL: <https://doi.org/10.1016/j.biopha.2019.108783>.

- [161] Abbasali Abbasnezhad et al. "Nigella Sativa Improve Redox Homeostasis in Heart and Aorta of Diabetic Rat". In: *Current Nutrition & Food Science* 12.1 (2016), pp. 35–41. URL: <https://www.ingentaconnect.com/content/ben/cnf/2016/00000012/00000001/art00009>.
- [162] Md. Quamrul Hassan et al. "Nigella sativa protects against isoproterenol-induced myocardial infarction by alleviating oxidative stress, biochemical alterations and histological damage". In: *Asian Pacific Journal of Tropical Biomedicine* 7.4 (2017), pp. 294–299. URL: <https://doi.org/10.1016/j.apjtb.2016.12.020>.
- [163] Seref Ezirganli et al. "The Effects of Nigella Sativa Seed Extract on Bone Healing in an Experimental Model". In: *Journal of Craniofacial Surgery* 27.7 (2016), pp. 1905–1909. URL: <https://doi.org/10.1097/SCS.0000000000002986>.
- [164] Osama A. Badary et al. "Acute and subchronic toxicity of thymoquinone in mice". In: *Drug Development Research* 44.2-3 (1998), pp. 56–61. URL: [https://doi.org/10.1002/\(SICI\)1098-2299\(199806/07\)44:2/3%3C56::AID-DDR2%3E3.0.CO;2-9](https://doi.org/10.1002/(SICI)1098-2299(199806/07)44:2/3%3C56::AID-DDR2%3E3.0.CO;2-9).
- [165] Wong Pei Lou et al. "Sub-acute toxicity of black seed (Nigella sativa) and honey mixture". In: *Malaysian Applied Biology* 47.6 (2018), pp. 11–18. URL: <https://core.ac.uk/reader/195387730>.
- [166] Ali M. Al-Amri and Abdullah O. Bamosa. "Phase I Safety and Clinical Activity Study of Thymoquinone in Patients with Advanced Refractory Malignant Disease". In: *Shiraz E-Medical Journal* 10.3 (2009), pp. 107–111. URL: <https://brieflands.com/articles/semj-76453.html>.
- [167] Jestin V. Thomas et al. "A phase I clinical trial to evaluate the safety of thymoquinone-rich black cumin oil (BlaQmax®) on healthy subjects: Randomized, double-blinded, placebo-controlled prospective study". In: *Toxicology Reports* 9 (2022), pp. 999–1007. URL: <https://doi.org/10.1016/j.toxrep.2022.04.020>.