NLP phytotoxin, a selective - biological herbicide of dicot weeds

Fatemeh Hashemlou, Morteza Mirzaei, Ali Mohammad Latifi, Nadeali Babaeian and Fatemeh Shakeri

Abstract— NLPs (Necrosis and ethylene inducing proteins) as microbial phytotoxins have been found in extensive range of microorganisms with different necrotic ability that cause hypersensitive immune response in dicotyledones weeds which result in extensive necrosis and death. This protein is not recognized by the immune system of monocot plants. NLPs have been used on dicot plants with various method like spraying, injection, hanging drop method and transferring its corresponding gene by Agrobacterium bacteria to the plant. Also a lot of auxiliary factors have been done such as the use of detergents, fungal or bacterial cell mid NLPs in order to increasing its effectiveness. The purpose of this paper is review of the target plants or host range of NLP, the range of microorganisms having this phytotoxine, how it works, ways to effectively use it as well as factors affecting its functionality as a biological herbicide.

Keywords—Biological control, NLP, dicot weed, monocot weed

I. INTRODUCTION

Weeds has always been a serious problem for farmers. In order to restricting and inhibiting the growth of the weeds, different strategies have been employed. The biological control methods attract attention, due to having no environmental pollution and production residues. NLP protein was isolated first in 1995 from Fusarium oxyporum [10]. After discovering it, NLPs have been used for biological

Manuscript received: February 8, 2014

Fatemeh Hashemlou is with Department of Agronomy, Faculty of Crop Sciences - University of Agricultural Sciences and Natural Resources - Sary -Iran and also is with Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran (e-mail: tanin_65@yahoo.com)

Morteza Mirzaei is with Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran (providephone: +982188617712;fax: +982182482549;e-mail:mraga85@chmail.ir)

Ali Mohammad Latifi is with Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran (e-mail: amlatify@yahoo.com)

Nadeali Babaeian is with Department of Agronomy and Plant Breeding, Faculty of Agricultural Science - University of Agricultural Sciences and Natural Resources - Sary – Iran (e-mail: nbabaeian@yahoo.com)

and Fatemeh Shakeri is with Applied Biotechnology Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran (email:shakery1363@gmail.ir) control of various plants, especially weeds [12], [7], [17]. This protein effectively causes necrosis and induces expression of ethylene hormone, in a large number of dicot plants [7], [8], [24]. Various studies have been done on the mechanism of this protein in plant cells [26], target plants [31], and it producing microorganisms [16]. This protein is stable at ambient temperature for 24 days and since there are a lot of dicot weed, This protein has been introduced as a good candidate in the fields of monocots cultures such as wheat, barley, Zea and so on [15], [12], [5], [8]. It should also be noted that no human toxicity of this protein have been reported [8].

II. TARGET PLANTS:

The table below shows the most important plants that have been presented for biosensing with this protein. With a careful look at the table it can be concluded that the biological assays is done in most cases on the weeds in the tropics of South America and especially in the field of cocoa [6], [11]. Of course a research was conducted by Moghimi in iran that tobacco, tomatoes, Sophora alopecuriodes, sinapis arvensis has been studied [23], [3] and also Mirzaie [2] has worked on Sophora alopecuriodes and triticum aestivum and Shakeri has studied the effect of this protein on Sophora alopecuriodes, sinapis arvensis, Amaranthus retroflexus, Chenopodium album, Rumex crispus, Alisma plantago aquatic and bread wheat [1].

	I.	Target plants-	[16],	[17],	[7],	[32],	[12]
--	----	----------------	-------	-------	------	-------	------

Salvia	Ocimim	Opium	Coriandru	Ambrosia	Erythroxylu
officinalis	basilicum	Рорру	m sativum	artemisiifoli	m coca
				a	
Centaurea maculosa	Abutilon theophrasti	Trifolium repens	Diospyros virginiana	Albizia julibrissin	Nicotiana tabacum
Theobrom	Cirsium	Taraxacum	Cannabis	Sasamum	Albizia

a cacao	arvense	officinale	sativa	indicum	julibrissin	pl in
						-

III. RANGE OF MICROORGANISMS HAVING NLP GENE:

Many plant pathogens secrete progressing pathogenic bacteria toxins that its role is performed by cell killing. NLP producers like many bacteria, fungi, and oomysets specious establish a leaf necrosis and mediated immunological responses in many plants [25]. Many of these organisms are plant pathogens [19], so presence of NLP in a variety of herbal microorganisms, indicating the important role of these genes in pathogenecity. NLP as a trigger, causing necrosis of leaves, which are genetically distinct from immune-mediated cell death [27] and stimulates dependent protective levels of immunity in all studied dicot plants, But it is unsuccessful in monocot plants [19]. It is interesting that although NLPs are present in distinct organisms phylogenetically but their sequences remain remarkably conserved during evolution [26]. 7 amino acid of GHRHDWE and some Cysteine roots in all the sequences of these proteins are present in different organisms [30] Although protected areas can be related to the strategy of phatogenecity, However, the homologous gene in non phatogens like Vibrio pommerensi bacteria and ascomyset fungi such as Neurospora crassa is found. Also no homologue of NLP genes in plants, animals or Protista has been reported till now [33].

IV. ACTION MECHANISM OF NLPS IN DICOTYLEDONOUS PLANTS:

About action mechanism of these proteins and the mechanism of necrosis, numerous studies have been conducted. It has been recognized that NLPs are elicitors (compounds activating plant defense mechanisms) [20]. According to various studies it is suggested that this protein induces a series of responses similar to hypersensitivity in the plants. Hypersensitive responses in the plants induced locally infection and eventually cause necrosis of the infected area which results in higher expression of ACC synthase and ACC oxidase enzymes, these enzymes are key products of ethylene biosynthesis. Following the increase in ethylene, activation of MAP kinase, phytoalexine synthesis and intracellular calcium increase is observed [6]. As regards the mechanisms of immune responses is induced by plants to limit the infection process, but the severity of these responses cause the loss of

ant. In the other words, NLPs Log in vascular plants and duce plant defense responses lead to signaling cascade in plant, resulting in extensive tissue necrosis and death [4]. The answers can cause extensive necrosis usually in 24 hours after entry into the vascular system of the plant. Further observation indicates that the toxin-induced interference with the integrity of the cell may be in immunological responses of the plants, when NLP contacts with potential defense of the plant several times, it reaches to its peak [29], [27]. NLP induces several genes that are often associated with different stress- defense responses [12], [6], [29], But as noted NLPs are just effective on dicot plants. The lack of effect on the monocot plants identified that NLPs is not recognized by the immune system of the monocot plants, in other words, there is a receptor protein in the dicot plants that is not exist in the monocot plants and therefore they are not sensetive to any of these proteins. sSo far in studies on host range of NLP no response has been reported despite the use of monocot plants [8]. sSince NLP increases ethylene emission in tobacco leaves [9], [15], [18], It is suggested that necrosis could be an indirect effect of this hormone. However, in some plants necrosis inducing did not occur accompanied by ethylene production [9], [10]. Also NLPs disrupt the homeostasis of ions and membrane potential, leading to the activation of defense responses. Significantly NLP induces falling H⁺ and Ca²⁺ and K⁺ losses [15], [18]. NLP involves in stimulation of virulence -dependent genes in plant, changes in K⁺ channel flow and H⁺, accumulation of reactive oxygen species (ROS) and ethylene, changing in cell respiration, increased response and sensitivity to local death [15], [18], [21]. Due to the absence of stomata in the stems and roots, NLPs are just effective on leaves. In studies to evaluate the biological activity of NLPs, five absorption methods in plants have been utilized:

- 1. Removing leaves from petioles and putting them in Ffungal culture filtrate
- 2. Spraying of Ffungal culture filtrate mid detergent on leaves
- 3. Fungal culture filtrate injected into leaves
- 4. PBI121 vectors for gene transfer by Agrobacterium injection method with agerfilteration
- 5. transfering of *nep1* gene into a suitable expression vector, such as E.coli for obtaining more proteinAnd then extract it and evaluate its effect in different ways such as spraying and etc on the desired plant
- 6. Hanging drop method

Each of the above methods has different benefits; Depending on the purpose of the assay one of these methods can be selected. For example, as in the experiments of Jennings in year 2000, different ways were assayed on the leaf, including spray of the protein, absorption through the leaf petiole and injected into the leaf, resulting in even more destructive effect than other about spraying method Aand even using the spray several times for obtaining further damage has been recommended [18], [10]. Also because the ease use of this method as a biological herbicide and increased treatment areas is recommended. In the difference between extracting protein from producing microorganisms or the expression vector, the second method is recommended because of the increased production of these proteins and ensure that the desired protein is the only factor (and not other metabolites produced by the microorganism) that effects on the plant [3].

V. EFFECTIVE FACTORS ON PERFORMANCE OF NLPS:

A. Temperature:

Previous research indicates that NLP activity is sensitive to temperature and less necrosis ability after exposure to temperatures of 65 ° C for 15 min [8], [18]. The sensitive biological activity of NLP to heat is so that heating this protein for 10 min at 95 ° C for 1 hour or more, causes the protein break down and small molecular weight compounds are made that are observed in SDS-PAGE, and biological activity disappears [10], [14]. Also NLPs are stable for 38 days at 4 ° C and 28 days at 20 ° C [8], [25]

B. The difference in the necrosis ability of NLPs:

NLP producing Microbial Rresources, has different ability with each other, and not all proteins in a functional level [35], [15] for example, BcNEP1 from *Botrytis cinerae* is 5-10 times more potent than this protein in *Fusarium oxyporum* and therefore related symptoms are more severe [35].

C. The effect of NLP concentration on its effectiveness:

In several studies the effects of increasing the concentration of NLP has evaluated and it has been shown that this increasing is more destructive [18] effective concentration has a critical threshold and more and better effect cannot be seen beyond it.

Also the effects of this protein on various plants are different, so that harmful effects on some plants are stronger [21].

VI. INFLUENCING FACTORS ON INCREASING THE EFFECT OF NLPS:

- In different experiments indicated that factors such as detergent enhances the effect of these proteins [18], [21], [1] so by creating turbulence in the cell membrane results in better penetration of this protein and its effects have been increased.
- 2. Jennings also tested the effect of ethylene pretreatment of weed leaves prior to treating with NLP, the level of induced ethylene production was increased approximately threefold in this expriment [18].
- 3. Moreover, several tests on the composition of the protein by fungi or bacteria are used. For example Gronwald et al [2004] examined the effects of purified NLP from its producing fungi and Pseudolonas syringae on 3 weeds of Ambrosia artemisiifolia, Cirsium arvense and Taraxacum officinale [17]. In this study, it was presented that the combination of CFU 10 from Pseudolonas syringae bacteria and 5 μ g / ml of purified protein NLP caused 60 to 80 percent necrosis in leaves of these three weeds [17].
- In another study by Bailey and colleagues in 2000 for the biological control of Opium poppy, fungal spores of Pleospora papaveracea with purified NLP from its producing fungal were used. In this study was cleared that combined use of fungal spores along 10 μg / ml of purified NLP will increase the control of Opium poppy [7].
- In addition to the direct use of NLP, gene transfering into fungi lacking this protein was used in weed control. For example, transfering NLP gene to Colletorrichum coccodes for biological control of velvetleaf weeds, increased its pathogenicity ninefold [15]
- 6. Using NLP proteins from microorganisms that may be more powerful, as in the case mentioned about Botritys [35] Also using of microorganisms that have multi-gene families of NLP and assessment of their power is recommended.

7. combining of NLP with enzymes: Thomas et al [1998] in evaluating the Effect of Fusarium oxyporum in order to control of parasitic broomrape (Orobanche Cumana) found that When seeds are treated by Fusarium cells, compounds such as pectin methyl esterase and trans Almynaz pectin is secreted by the fungi, which can cause wall pectin to dissolve and facilitate the penetration of fungi to the cell walls of the seeds [34].

VII. CONCLUSION

Obviously the use of chemical agents to control weeds is dramatically dangerous because of their environmental damage, destructive effect on human and animals, causing resistant plants. therefore seeking for safer controlling ways are required. Among the controlling ways biological control because of what mentiond above are more economic and safer. Phytotoxins are one of the bioherbicides that produced are by different microorganisms and have different size and chemical structure. Among peptid phytotoxins, NLPs are an good option and can be possible alternative of chemical herbicide in the field of monocot cultures due to their selective effect just on dicot plants, rapid disintegration and not leaving rasidues in environment and therefore having no pollution effect on environment, human and animals, affecting in short time. In order to identifying this protein from other microorganisms, maybe strong proteins could find and so, no other contributing factors on its effect is required which could be more economic.

References

- F. Shakery and A. M. Latify. "The effect of microbial phytotoxins on some dicot weeds of Wheat and Rice", Master of science thesis of payame nour university. 2011. "Submitted for publication".
- [2]. M. Mirzaie and A. M. Latify, "Producing of nlp phytotoxin and asseyiny the effect of it on saphors mollis". Master of science thesis of emam hosseyn university. 2011. "Unpublished".
- [3]. H. Moghimy and J. Hamedi, "Molecular Screening of nlp genes in actynomysetes of iran and cloning of nep1 in agrobacterium tomeficiense and assaying the effect of it on tubacum". Master of science thesis of Tehran university. 2012. "Published".
- [4]. A. Aina, S. May and H. Clare, "The Centralization Phenomenon of Spinal Symptoms-a Symptomatic Review". Manual Therapy. Vol 9, pp 134-143. 2004.

- [5]. Z. Amsellem, B. A. Cohen, and J. Gressel, "Engineering Hypervirulence in an Inundative Mycoherbicidal Fungus for Efficient Weed Control", Nature Biotech. Vol 20, pp 1035-1039. 2002.
- [6]. B. A. Bailey, H. Bae, M. D. Strem, C. Antunez, G. Mayolo, M. J. Guiltinan, J. A. Verica, S. N. Maximova and J. H. Bowers, "Developmental Expression of Stress Response Genes in Theobroma cacao leaves and Their Response to Nep1 Treatment and a Compatible Infection by Phytophthora megakarya". Plant Physiology and Biochemistry, Vol 43, pp 611- 622, 2005.
- [7]. B. A. Bailey, P. C. Apel-Birkhold, O. O. Akingbe, J. L. Ryan, N. R. O'Neill, J. D. Anderson."Nep1 protein from Fusarium oxysporum enhances biological control of opium poppy by Pleospora papaveracea". Phytopathology. Vol 90, pp812–818. 2000.
- [8]. B. A. Bailey, R. Collins and J. D. Anderson.. "Factors influencing the herbicidal activity of Nep1, a fungal protein that induces the hypersensitive response in Centaurea maculosa. Weed". Sci vol 48, pp 776-785. 2000.
- [9]. B. A. Bailey, J. C. Jennings and J. D. Anderson, "The 24-kDa Protein from Fusarium oxysporum f. sp. Erythroxyli, Occurrence in Related Fungi and the Effect of Growth Medium on its Production". Can Journal Microbiology, Vol 43, pp 45- 55. 1997.
- [10]. B. A.Bailey, "Purification of a Protein from Culture Filtrates of Fusarium oxysporum that Induces Ethylene and Necrosis in Leaves of Erythroxylum coca". Phytopathology, Vol 85, pp 1250-1255. 1995.
- [11]. H. Bae, S. H. Kim, M. S. Kim, R. C. Sicher, D. Lary, M. D. Strem, S. Natarajan, B. A. Bailey, "The drought response of Theobroma cacao (cacao) and the regulation of genes involved in polyamine biosynthesis by drought and other stresses". Plant Physiol Bioch, vol 46, pp174-188, 2008.
- [12]. B. J. Bang-Jun Zhou, P. S. Jia, F. Feng Gao and H. S. Guo, "Molecular Characterization and Functional Analysis of a Necrosis- and Ethylene-Inducing, Protein Encoding Gene Family from Verticillium dahlia Molecular Plant-Microbe Intractions". Vol 25, pp 964-975, 2012.
- [13]. L. Clare, G. Pemberton and G. P. C. Salmond, ^{ic}The Nep1-like proteins a growing family of microbial elicitors of plant necrosis. Molecular" Plant Phsiol. Vol 5, pp353–359, 2004.
- [14]. S. Dong, G. Guanghui Kong, X. Qutob D Yu, J. Junli Tang, J. Jixiong Kang, T. Dai, H. Wang, M. Gijzen and Y. Wang, "The NLP Toxin Family in Phytophthora sojae Includes Rapidly Evolving Groups That Lack Necrosis-Inducing Activity". Molecular Plant-Microbe Intractions. Vol 25, pp 896-909, 2012.
- [15]. G. Fellbrich, A. Romanski, A. Varet, B. Blume, F. Brunner, S. Engelhardt, G. Felix, B. Kemmerling, M. Krzymowska and T. Nurnberger, "NPP1, a Phytophthora-Associated Trigger of Plant Defense in Parsley and Arabidopsis". Plant Journal. Vol 32, pp 375-390, 2002.
- [16]. M. Gijzen and T. Nurnberger, "Nep1-like proteins from plant pathogens: Recruitment and diversification of the NPP1 domain across taxa Phytochemistry". Phytochem, vol 67, pp 1800–1807, 2006.
- [17]. W. Gronwald, L. Kathryn, Plaisance and B. A. Bailey, "Effects of the Fungal Protein Nep1 and Pseudomonas syringae on Growth of Canada Thistle (Cirsium arvense), Common Ragweed (Ambrosia artemisiifolia), and Common Dandelion (Taraxacum officinale)«.Weed Science, vol 52, pp 98-104, 2004.
- [18]. J. C. Jennings, P. C. Birkhold, N. M. Mock, C. J. Baker, J. D. Anderson and B. A. Bailey, "Induction of defense Responses in Tobacco by the Protein Nep1 from Fusarium oxysporum". Plant Science, Vol 161, pp 891-899, 2001.
- [19]. S. Kamoun, "A Catalogue of the Effector Secretome of Plant Pathogenic Oomyce Tes" Annual ReviewPhytopathology vol 44, pp 41–60, 2006.
- [20]. T. Koch, S. Schulz, H. Schröder, R. Wolf, E. Raulf and V. Höllt, "Carboxyl- Terminal Splicing of the Rat μ Opioid Receptor Modulates Agonist- Mediated Internalization and Receptor Resensitization". Biology Chemical, Vol 273, pp 13652-13657, 20,1998.

- [21]. S. E. Keates, T. A. Kostman, J. D. Anderson and B. A. Bailey, "Altered Gene Expression in Three Plant Species in Response to Treatment with Nep1, a Fungal Protein that Causes Necrosis". Plant Physiology, Vol 132, pp 1610-1622, 2003.
- [22]. Y. Q. Li, Z. L. Sun, X. F. Zhuang, L. Xu, S. F. Chen and M. Z. Li, "Research progress on microbial herbicides". Crop Prot. Vol 22, pp 247-252, 2003.
- [23].Moghimi H, Hamedi J, Sepehrizadeh Z, Ofoghi H.2012.Overexpression of recombinant Nep1 in Escherichia coli and its use as a biological agent for control of Sinapis arvensis. Ann Microbiol1-7.
- [24]. J. Motteram, I. Küfner, S. Deller, F. Brunner, K. E. Hammond-Kosack, T. Nürnberger and J. J. Rudd, "Molecular characterization and functional analysis of MgNLP, the sole NPP1 domain-containing protein, from the fungal wheat leaf pathogen Mycosphaerella graminicola". Mol Plant Microbe Interact, vol 22, pp790-799, 2009.
- [25]. C. Ottmann, B. Luberacki, I. Kufner, W. Koch, F. Brunner, M. Weyand and L. Mattinen, "Pirhonen, M. Anderluh, G.A. and Seitz, H.U. "Common Toxin Fold Mediates Microbial Attack and Plant Defense", PNAS, vol 106, pp10359-10364, June 23. 2009.
- [26]. C. L. Pemberton, G. P. C. Salmond, "The Nep1- Like Proteins: A Growing Family of Microbial Elicitors of Plant Necrosis" Plant Pathology, vol 5, pp 353.359, 2004.
- [27]. D. Qutob, B. Kemmerling, F. R. Brunner, I. Kufner, B. F. Engelhardt, A. A. Gust, B. Luberacki, B. Hanns Ulrich Seitz, B. Dietmar Stahl, T. Rauhut, E. Glawischnig, G. Schween, B. Lacombe, N. Watanabe, G. Dodt, D. Hubert, M. Gijzen, T. Nurnbergerb, "Phytotoxicity and Innate Immune Responses Induced by Nep1-Like Proteins" The Plant Cell, vol 18, pp 3721–3744, 2006.
- [28]. S. M. Singh, A. K. Panda, "Solubilization and refolding of bacterial inclusion body proteins". J Biosci Bioeng, vol 99, pp 303-310. 2005.
- [29]. K. Slot and W. Knogge, "A Dual Role for Microbial Pathogen- Derived Effector Proteins in Plant Disease and Resistance" Crit ReviewPlant Science, vol 21, pp229- 271, 2002.
- [30]. A. Schouten, P.V. Baarlen and A. L. Jan, "Blackwell Publishing Ltd Phytotoxic Nep1-like proteins from the necrotrophic fungus Botrytis cinerea associate with membranes and the nucleus of plant cells" Laboratory of Phytopathology, Wageningen University. 2007.
- [31]. J. A. Verica, S. N. Maximova, M. D. Strem, J. E. Carlson, B. A. Bailey, M. J. Guiltinan, "Isolation of ESTs from cacao(Theobroma cacao L.) Leaves Treated with Inducers of the Defense Response". Plant Cell Reports. Vol 23, pp404-413. 2004.
- [32] S. Veit, J. M. Worle, T. Nurnberger, W. Koch, and H. U. Seitz, "A Novel Protein Elicitor (PaNie) from Pythium Aphanidermatum Induces Multiple Defense Responses in Carrot, Arabidopsis, and tobacco". Plant Physiology, Vol 127, pp 832- 841. 2001.
- [33]. J. Win, T. D. Kanneganti, T. Torto- Alalibo and S. Kamoun, "Computational and Comparative Analyses of 150 Full- Length cDNA Sequences from the Oomycete Plant Pathogen Phytophthora infestans" Fungal Genetics and Biology, vol 43, pp20–33. 2006.
- [34]. H. Thomas, J. Sauerborn, D. Muller-stover, A. Ziegler, J. S. Bedi and J. Kroschel, "The potenthial of Fusarium oxysporumf. sp. orthocerasas a biological control agent for Orobanche cumanain sunflower". Journal of Biological Control, vol 13, pp 41-48, 1998.
- [35]. A. schuten, K. J. Arnoldus, L. Van, "Novel necrosis and ethylene inducing proteins from botrytis", PCT/NL2005/050058,2006