

PRRS pathology and control

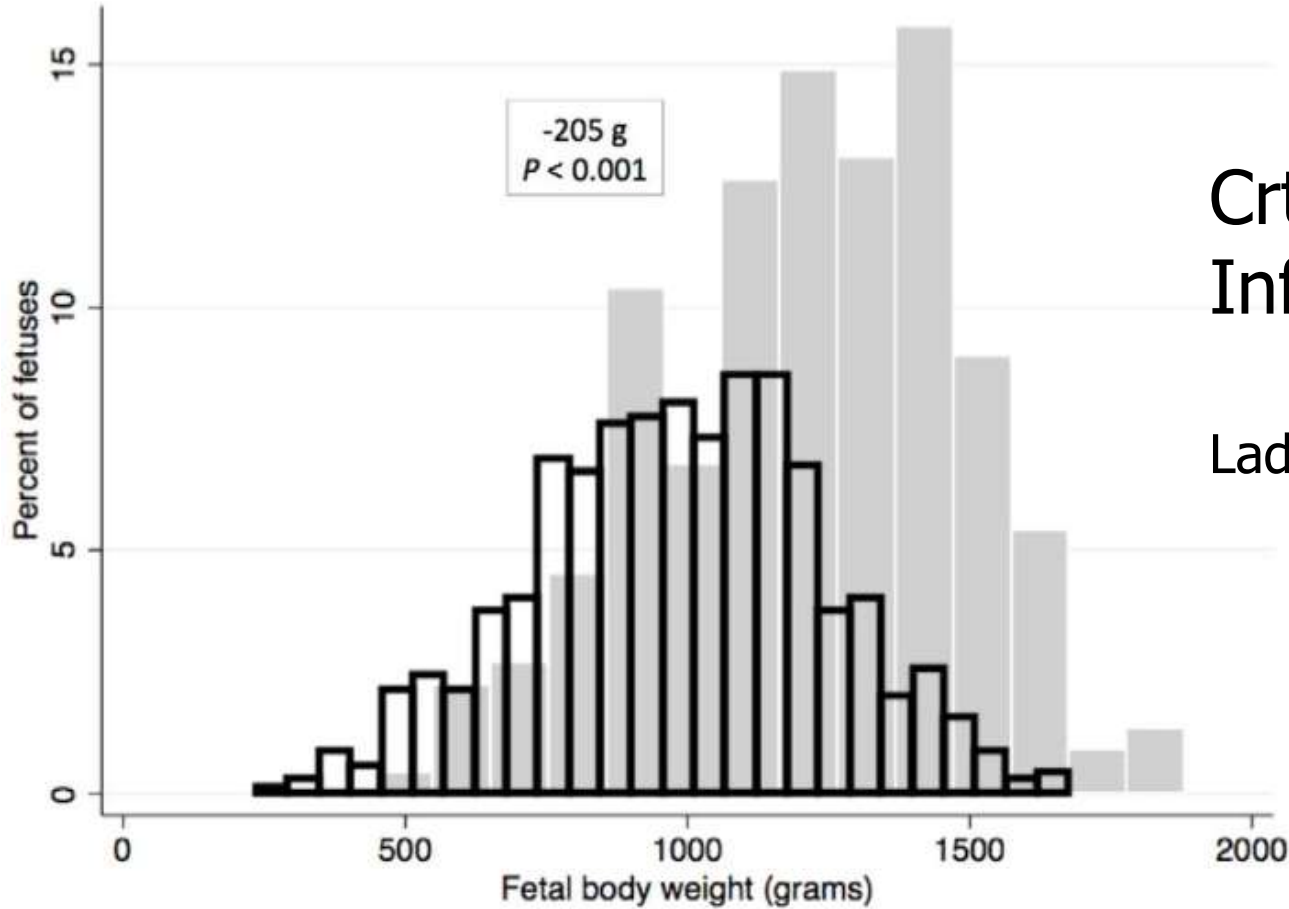
Reproductive pathology

- Early gestation
 - embryonic death, infertility return to estrus
- Late gestation
 - transplacental infection – abortions, early farrowing, fetal death, weak, congenitally infected piglets

Reproductive pathology

- Focal detachment/degeneration of the placenta (Karniychuk et al. 2009, 2013)
 - Sn/CD169 cell numbers increase in the placenta during late gestation
 - Activated NK cells induce placental damage
- Fetal infection – THYMUS!! (Ladinig et al. 2014, 2015)
 - Inoculation of 114 pregnant gilts at 85 DOG – extermination at 21DPI: >95% of dead fetuses were infected

Bodyweight of live piglets



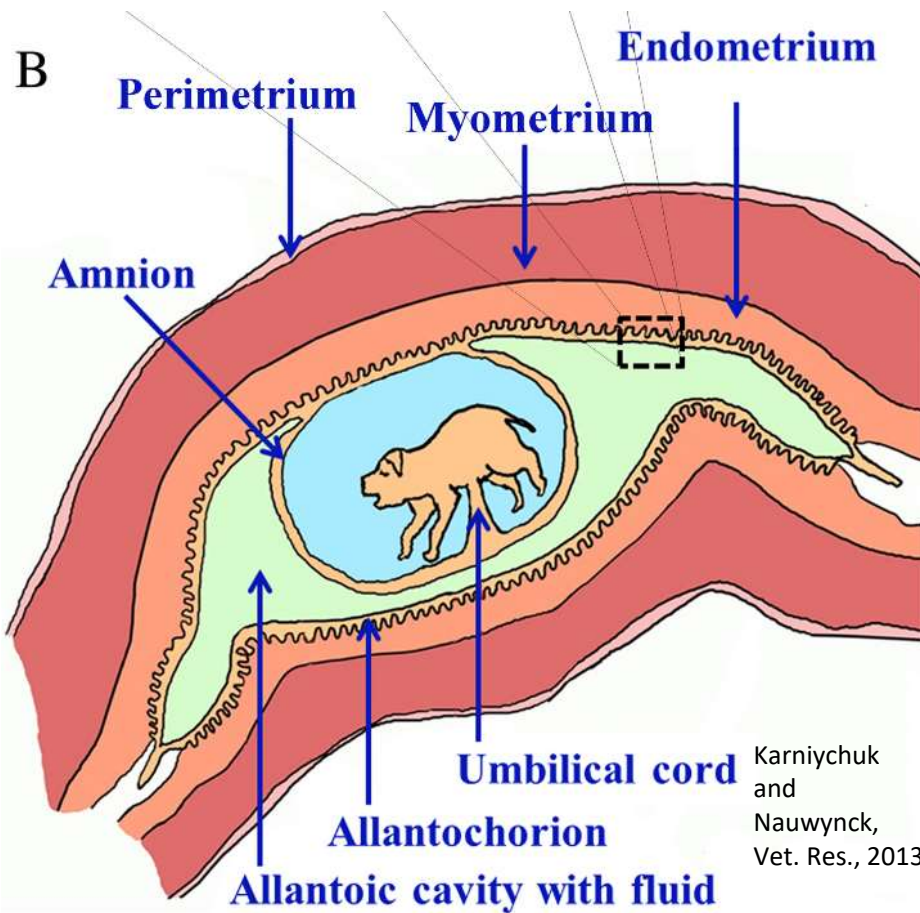
Ctrl: 1204 (± 36) g
Infect: 999 (± 21) g

Ladinig et al. 2014



Viable

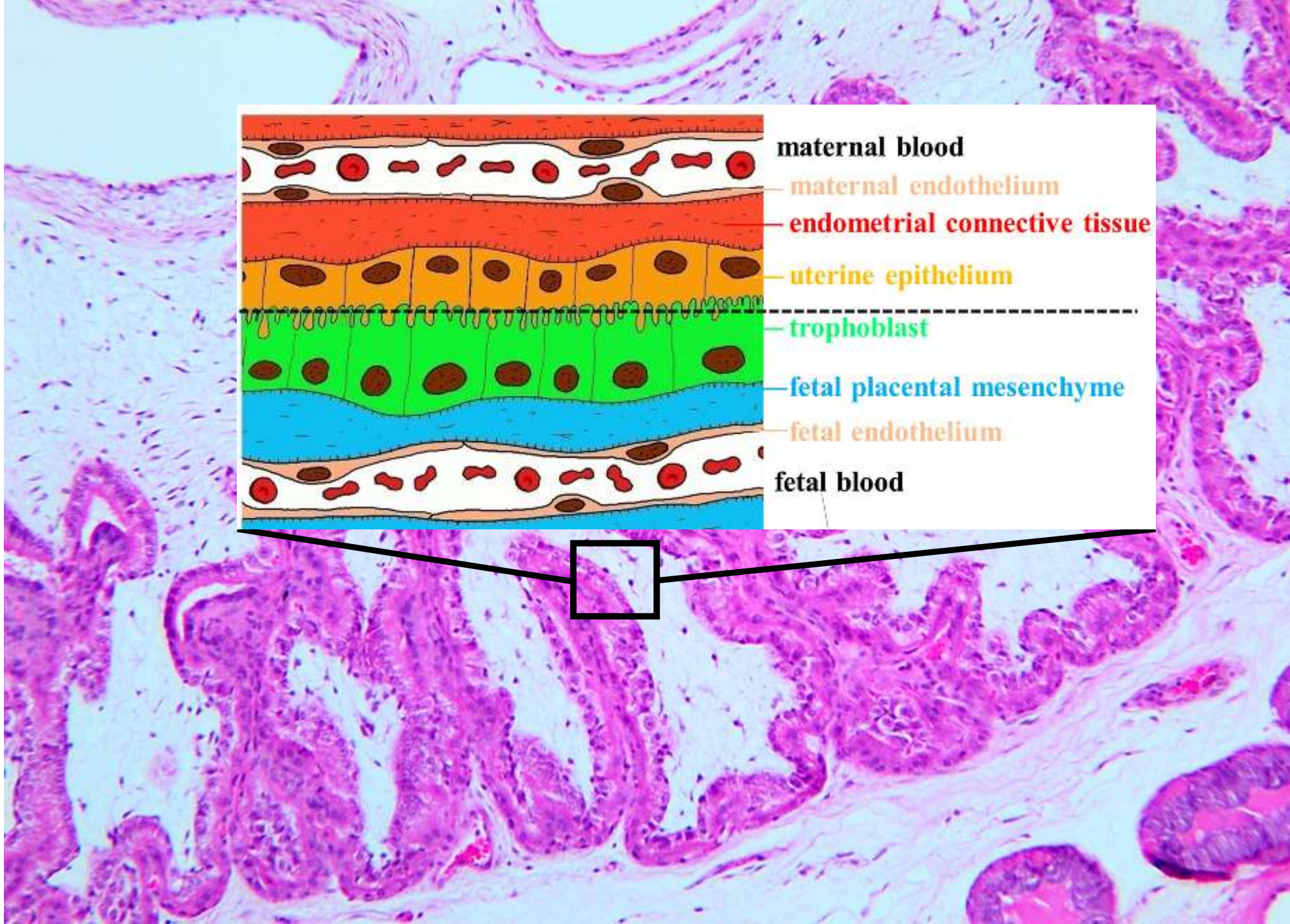
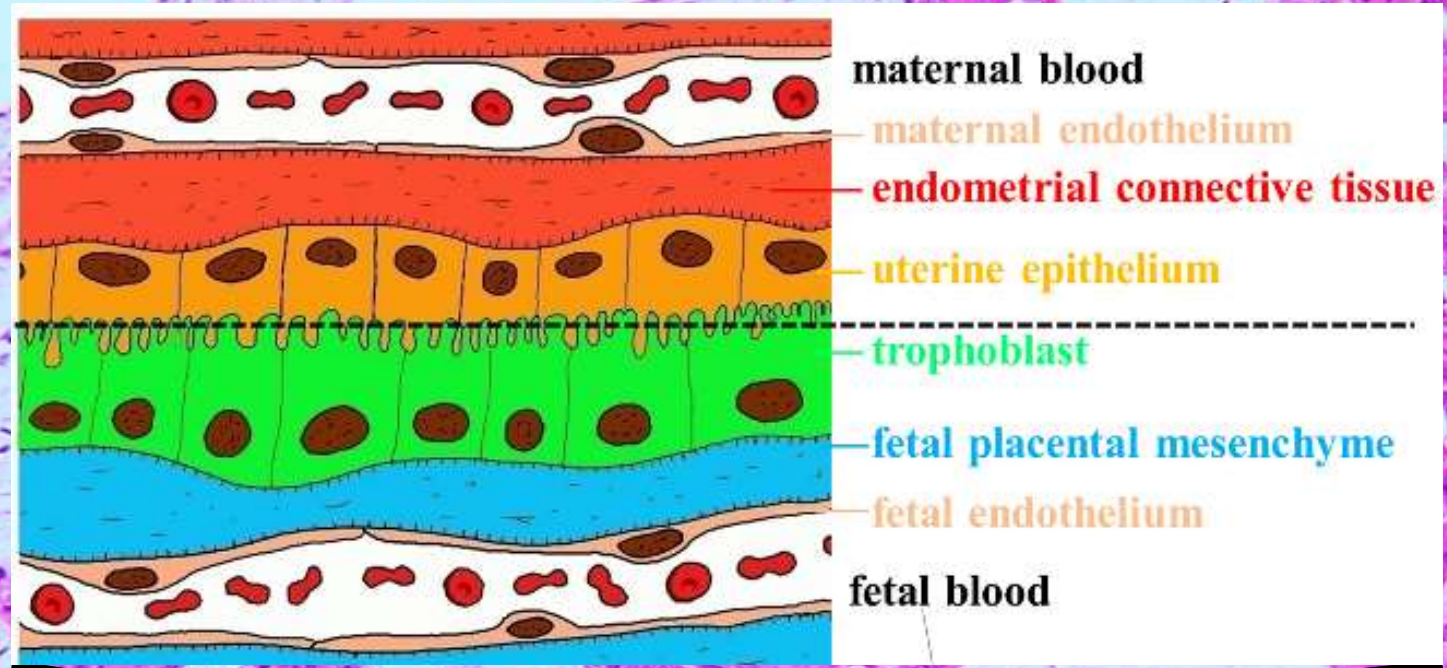
Reproductive PRRS Pathology



Andrea Ladinig,
Susan Detmer,
John Harding et al.
2014, 2015, 2016

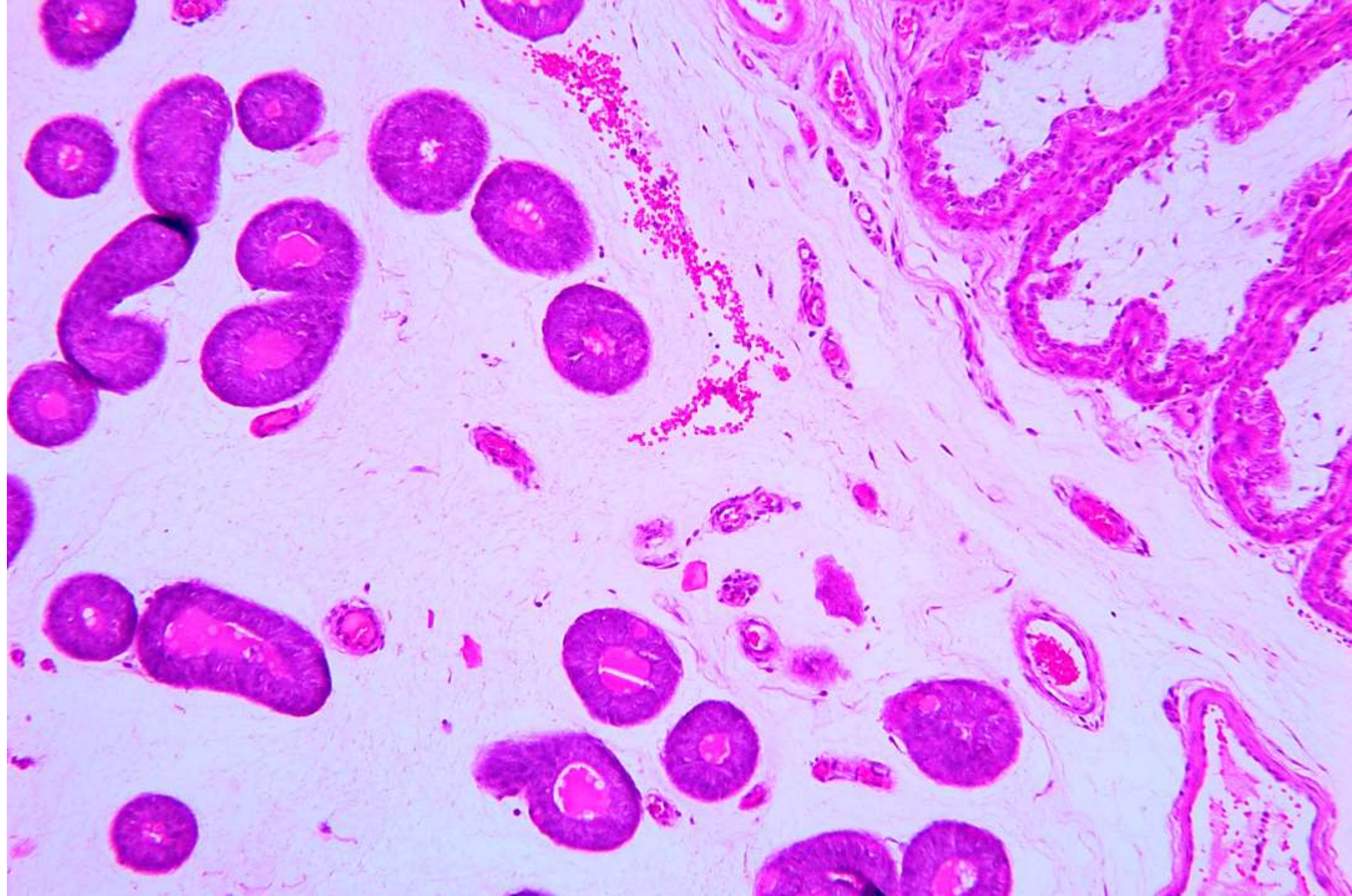
Incomplete diffuse epitheliochorial placenta

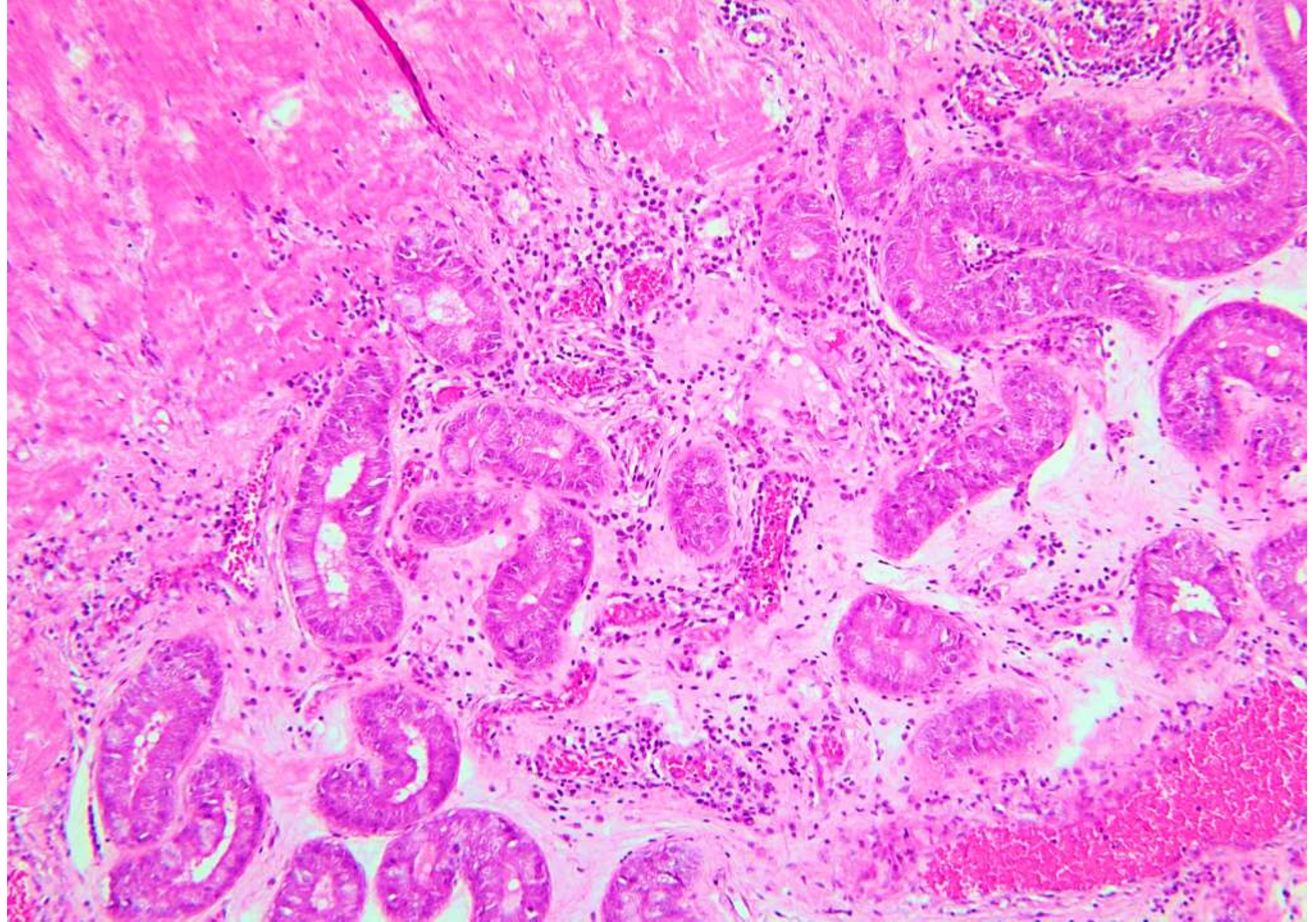
with atrophy at the peripheral tips
without invasion of fetal tissue into
the maternal endometrium, nor
endometrial decidualization

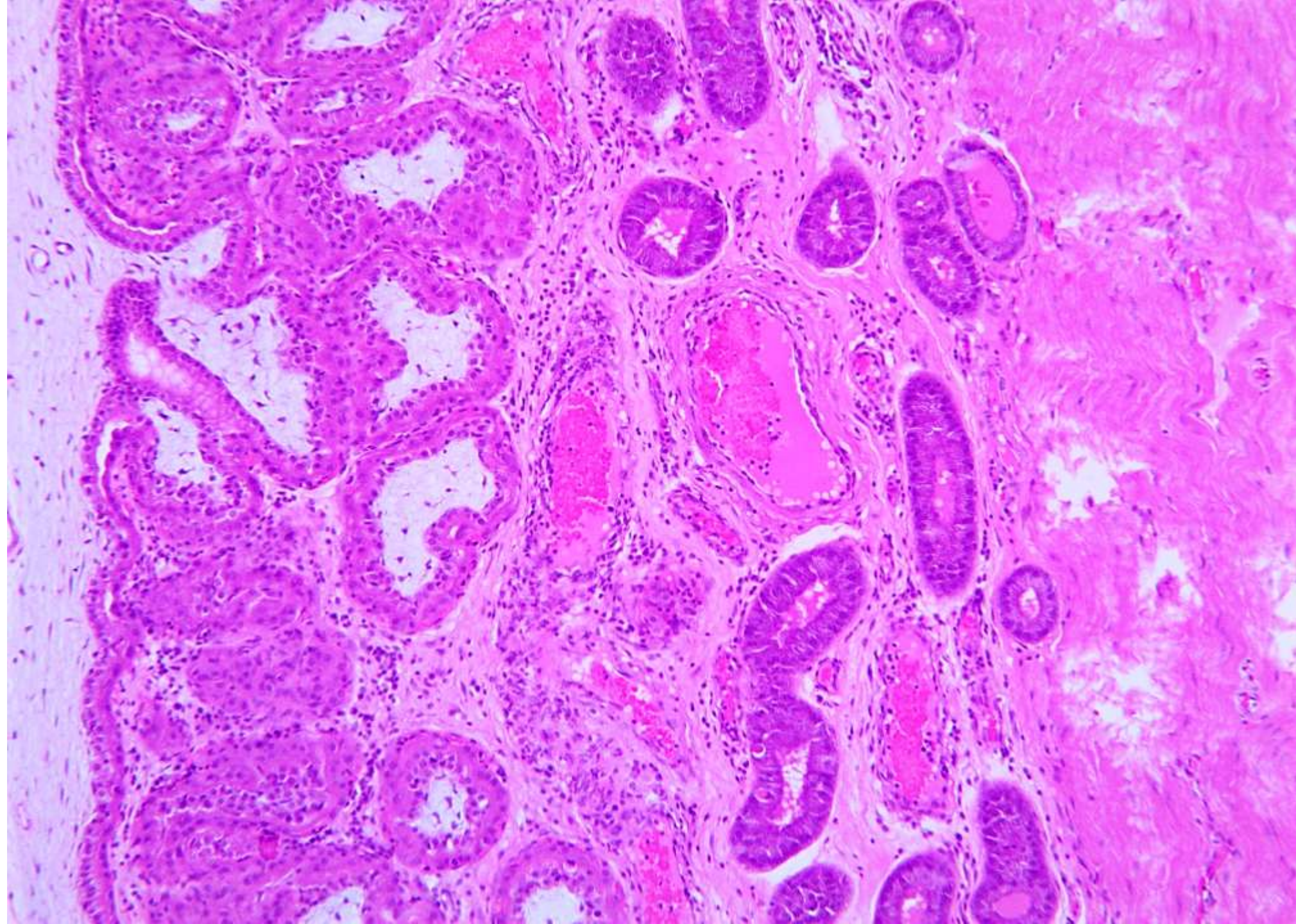


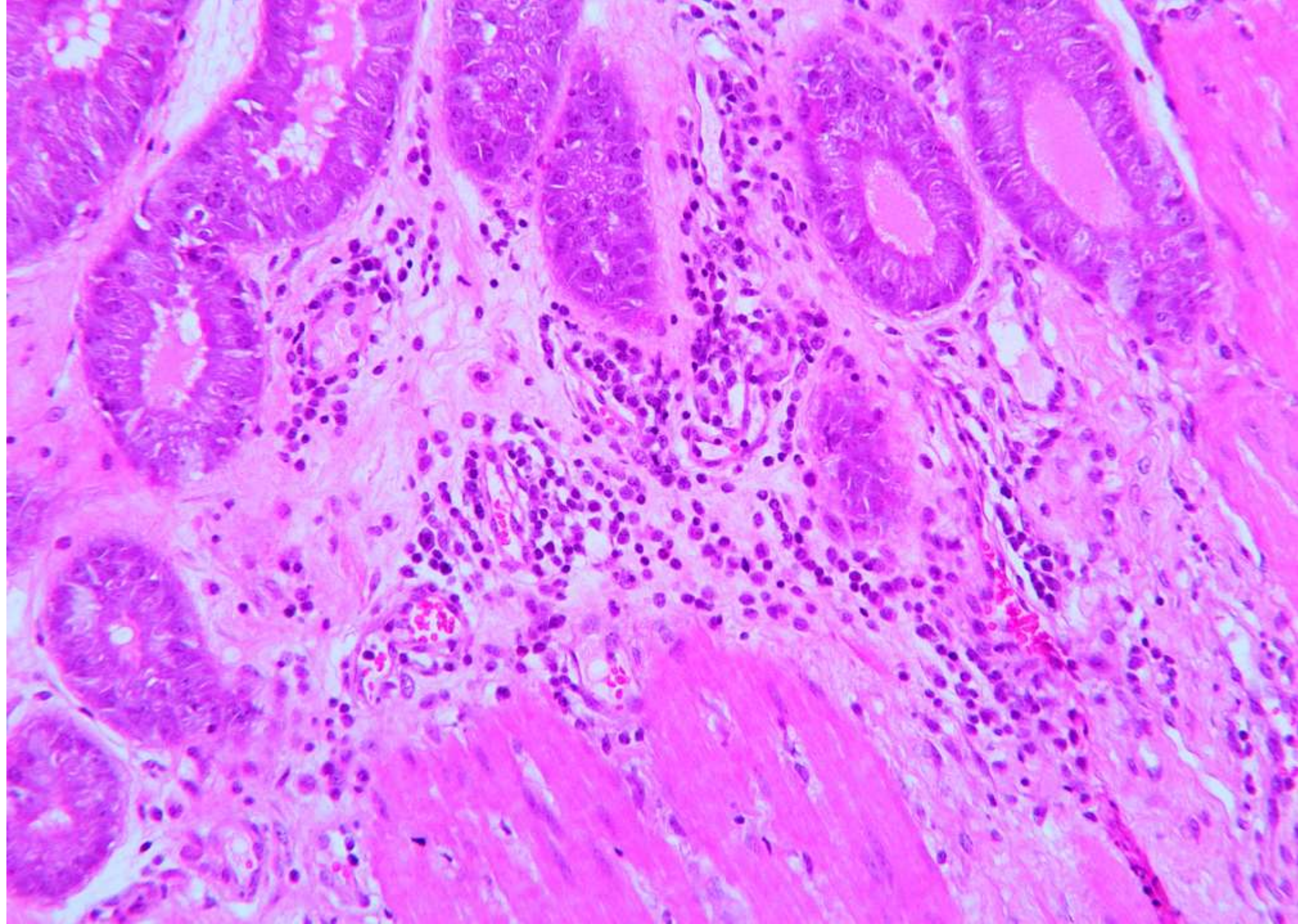
Fetal evaluation

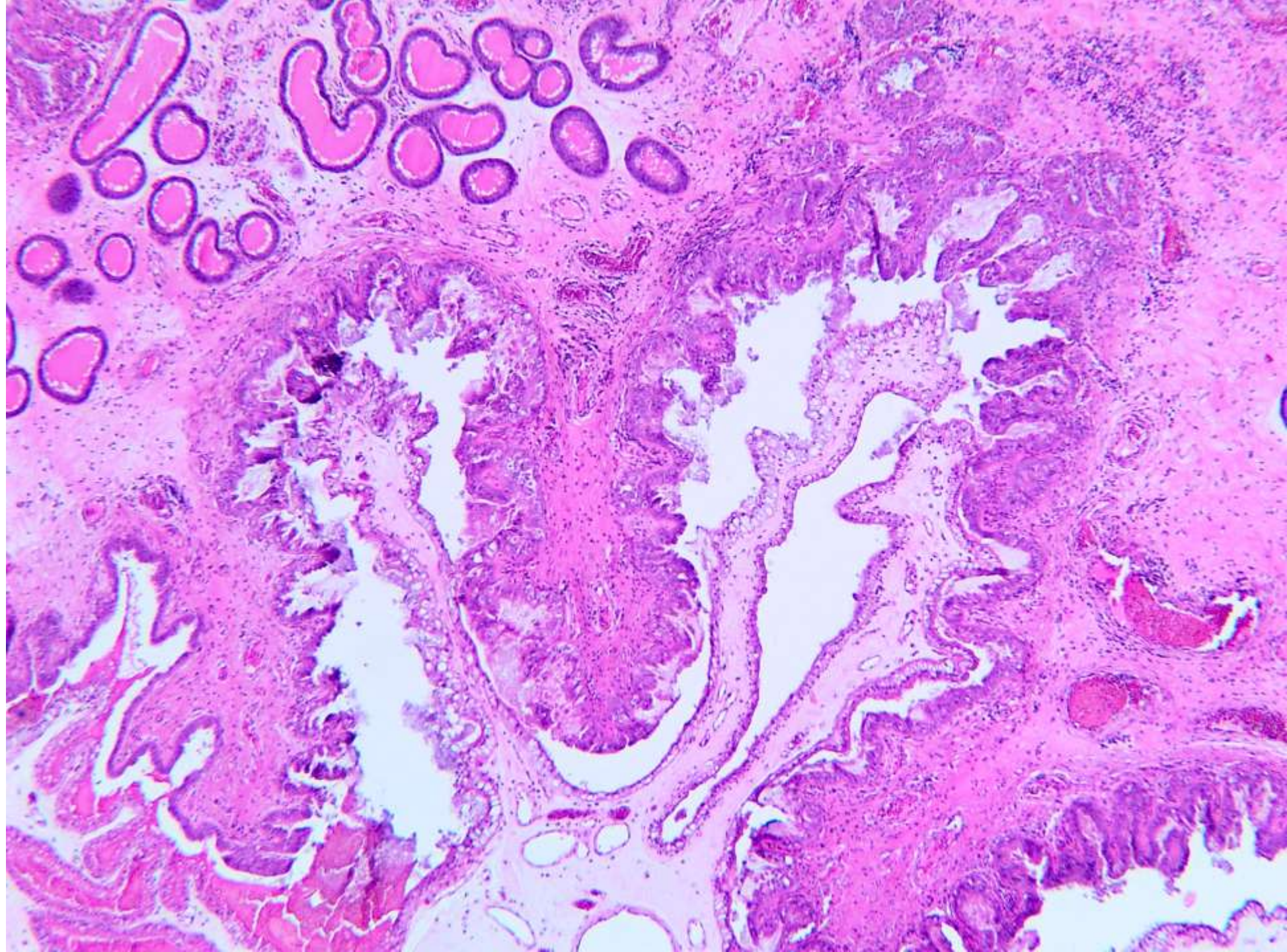


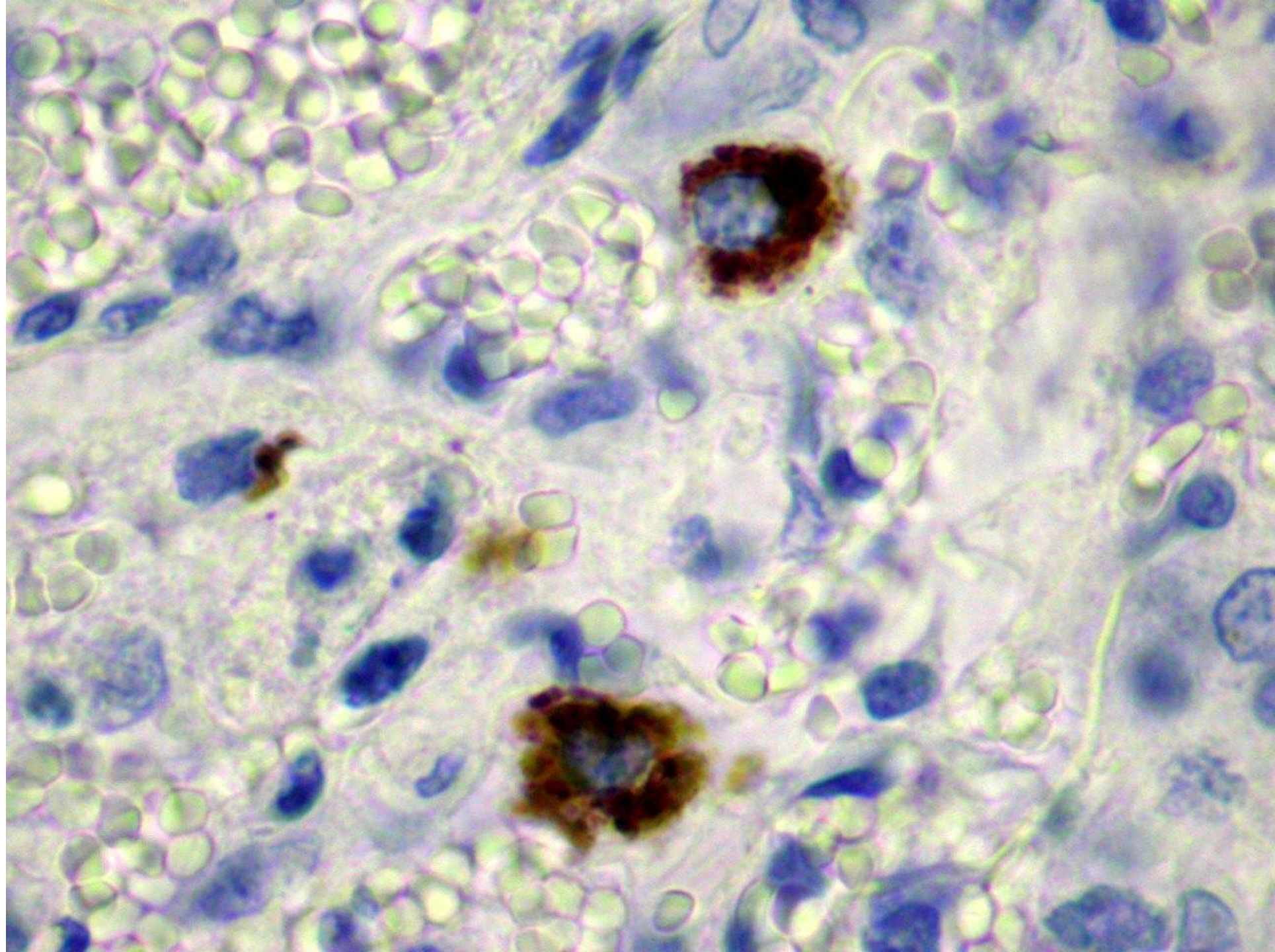









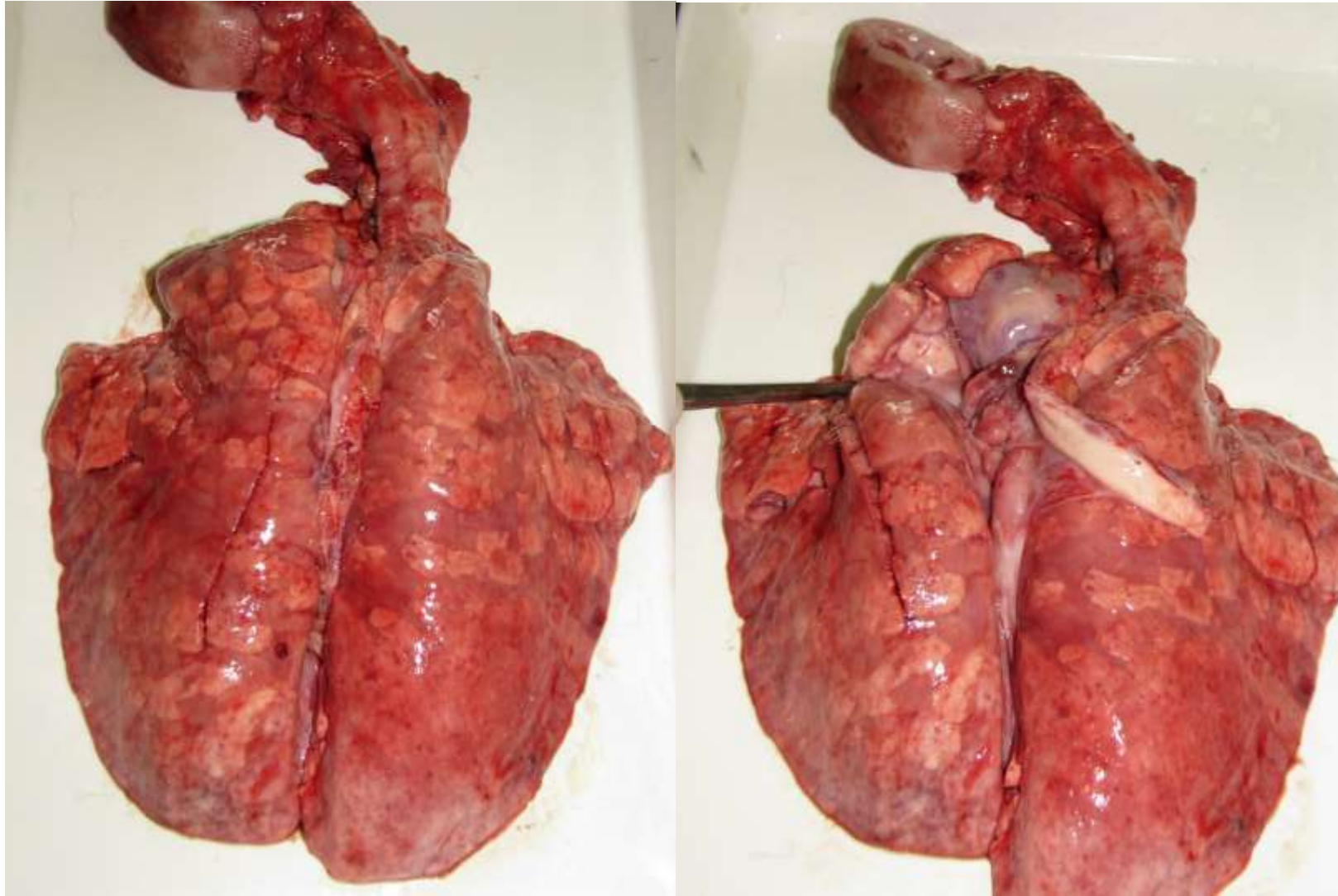


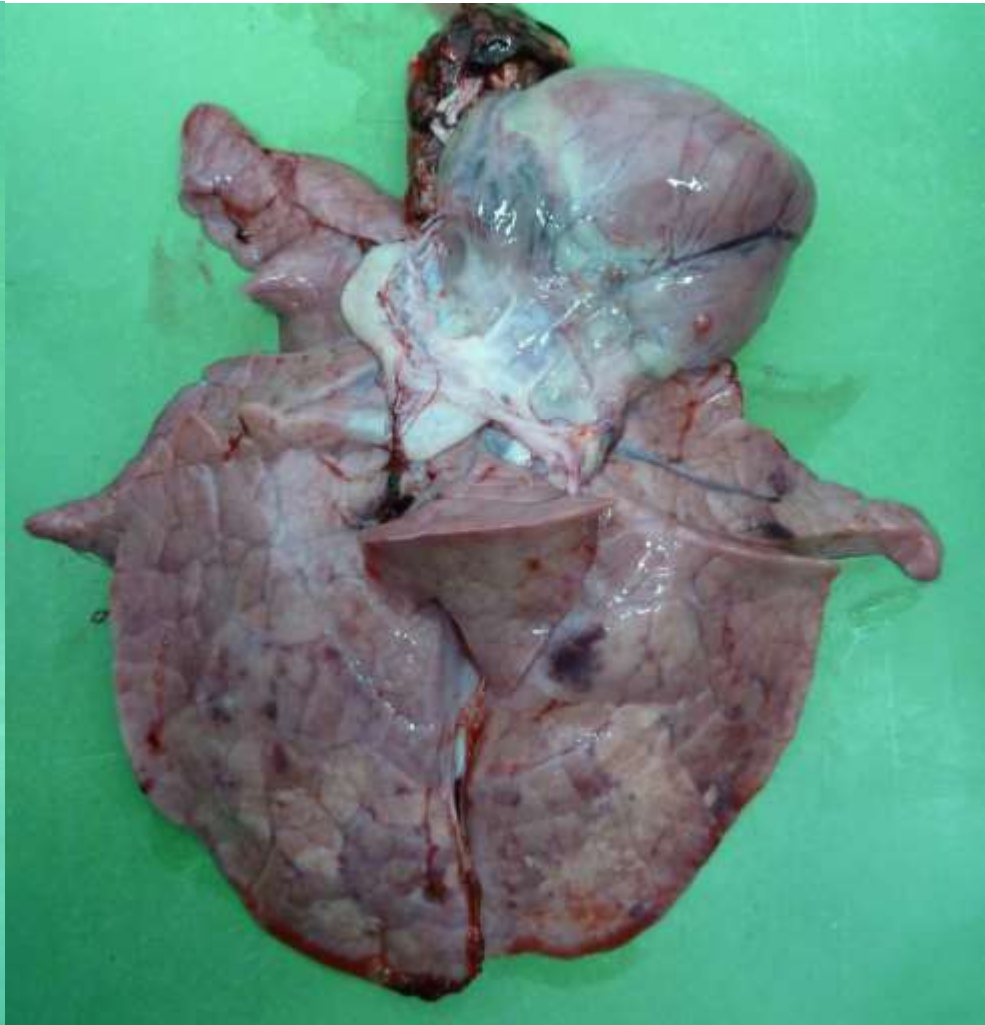
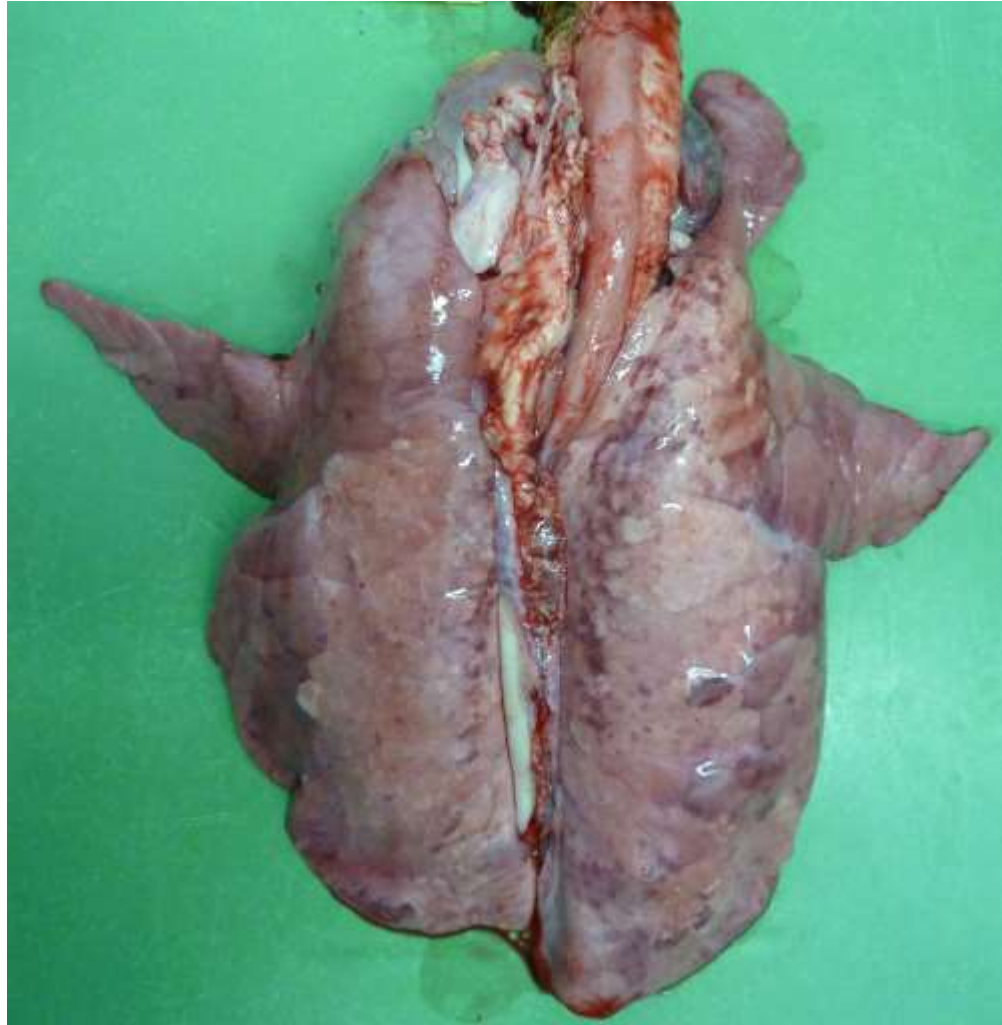


Immunopathogenesis in the lungs

(Gómez-Laguna et al., 2013)

- Pro inflammatory cytokine upregulation (TNF- α , INF- β)
- Anti inflammatory cytokine downregulation (IL-10, IL-4)
- Haptoglobin levels increase
 - CD163 receptor upregulation helps virus internalization, replication
 - Decrease the bactericidal activity of macrophages  **SECONDARY INFECTIONS**

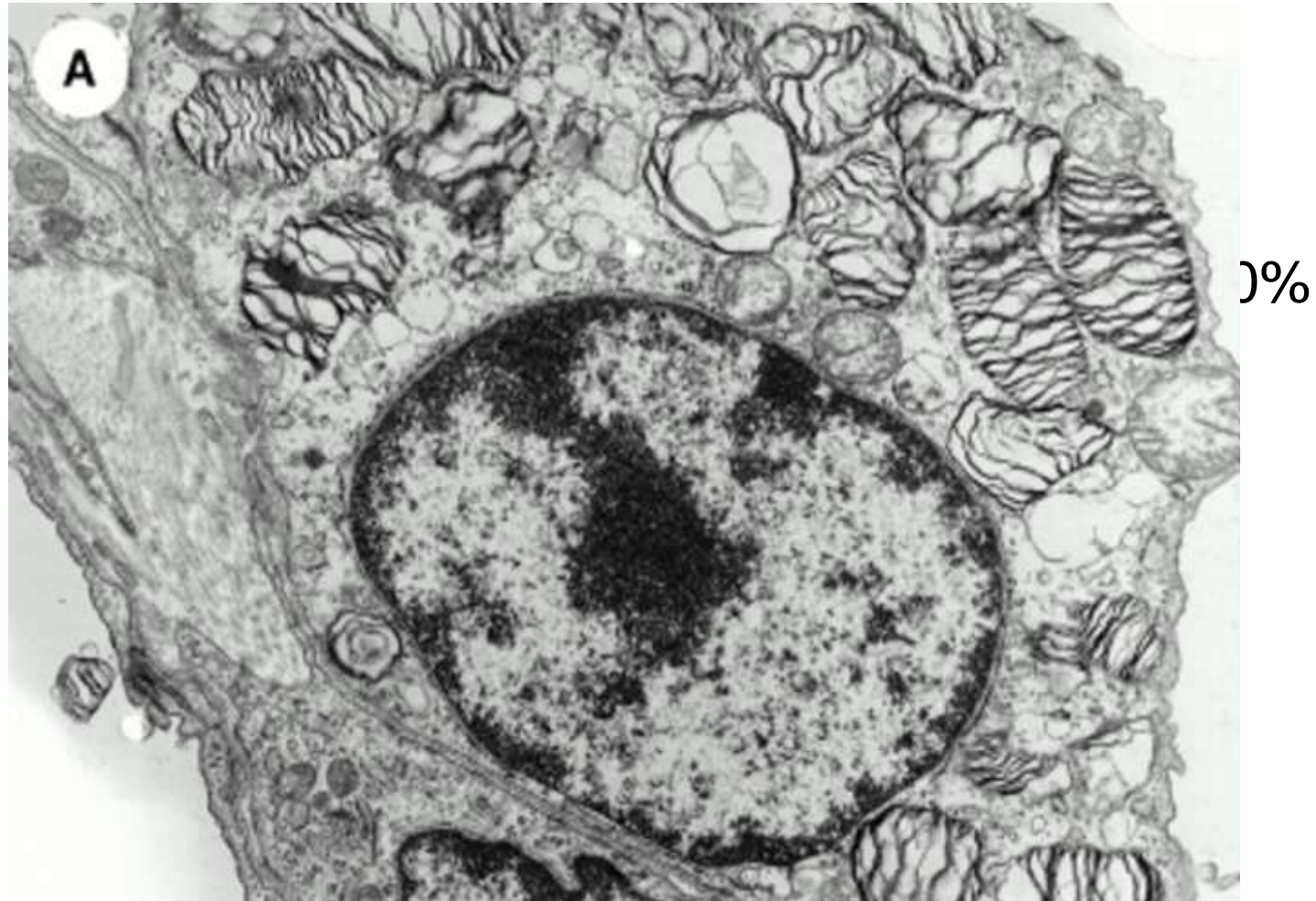




Microscopic lung lesions in PRRS

- Intralobular interstitial pneumonia
- Septal infiltration by mononuclear cells
- Intraalveolar necrotic debris
- Intraalveolar cellular infiltration
- Type 2 pneumocyte hypertrophy and hyperplasia

Type 2 pneumocytes



Aims

- Identification of type 2 pneumocytes in pigs
- Characterize PRRSV induced lesions
 - H&E
 - IHC
- Establish a new scoring system
- Find a more objective scoring method by counting IHC positive cell

PRRSV infection model

- 9-week-old piglets
- 2.2×10^5 TCID₅₀/ml, subtype 1 „wild” isolate, from Germany
- Euthanasia at 10 DPI (n = 7) and 21 DPI (n = 5), parallelly from PBS treated control group
- Necropsy and histopathology from all 7 lung lobes
- FFPE
- H&E
- IHC (left middle lobe only): TTF-1, Ki67, MAC387, panCK
- Blinded analysis

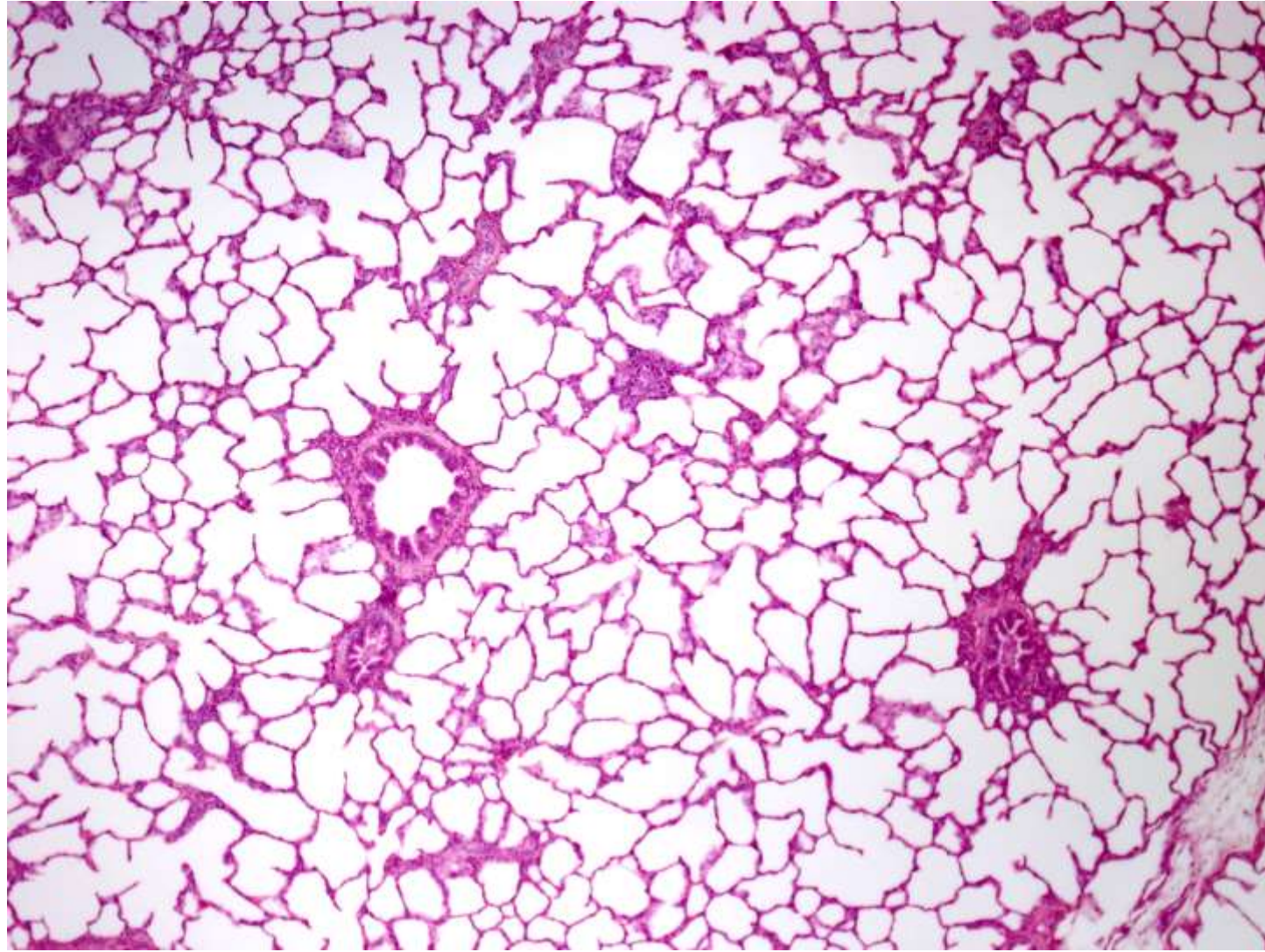
Scoring of H&E slides

Severity (0–3) and extent (0–3)

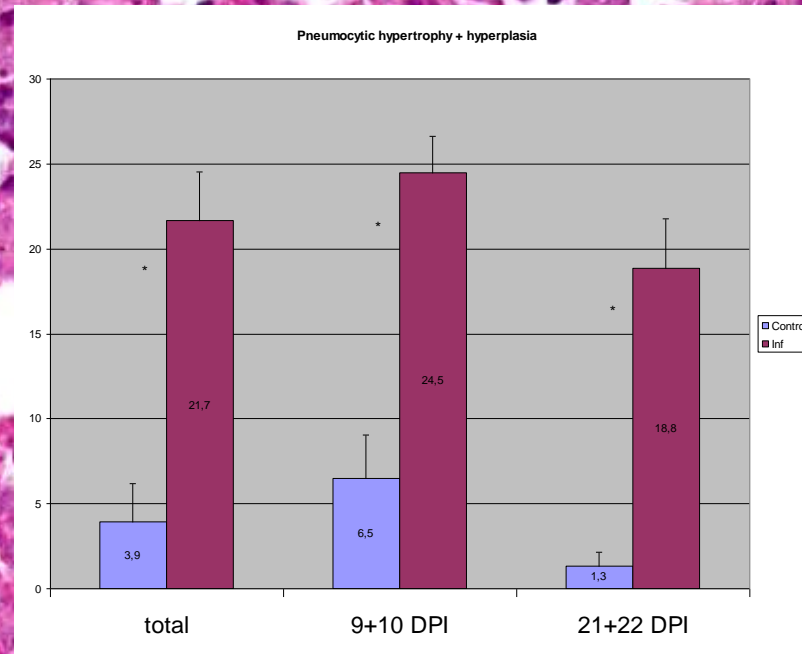
- pneumocyte hypertrophy and hyperplasia
- septal mononuclear infiltration
- intraalveolar necrotic debris accumulation
- intraalveolar inflammatory cells
- perivascular inflammatory cells

Scoring IHC slides

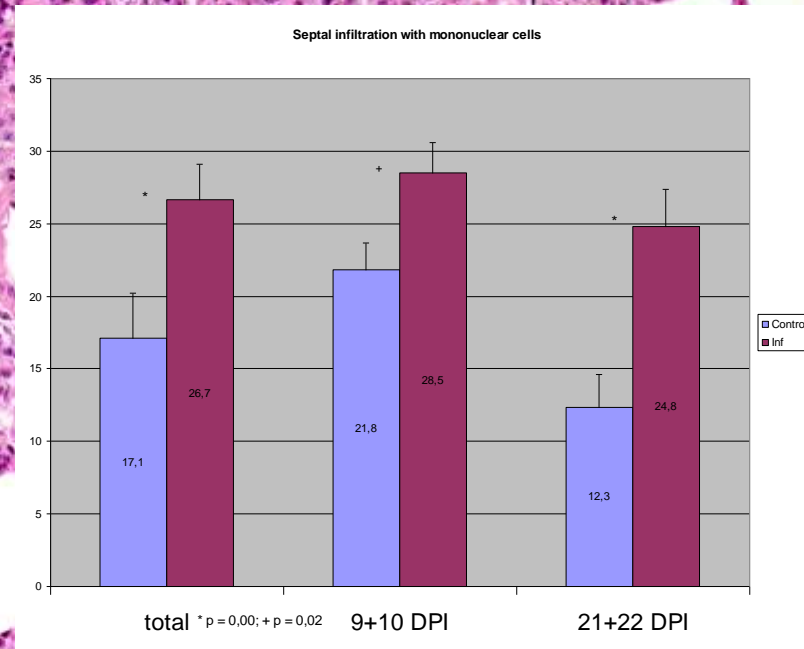
- TTF-1, Ki67, MAC387, (panCK)
- IHC positive cells on 50, non overlapping 0.20 mm² fields
- SPSS:
 - Significance calculation: Student's T test
 - Correlation analysis IHC and H&E scores: Pearson's X²-test



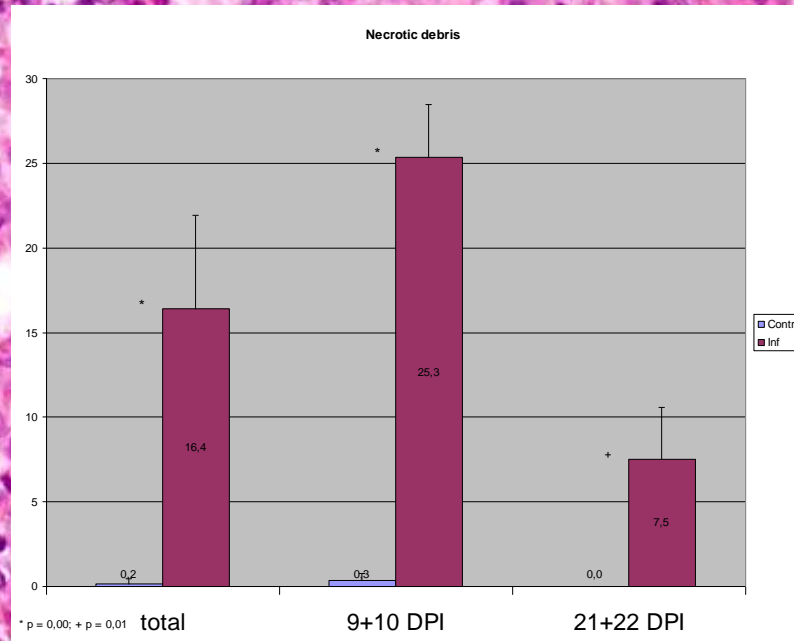
pneumocyte hypertrophy and hyperplasia



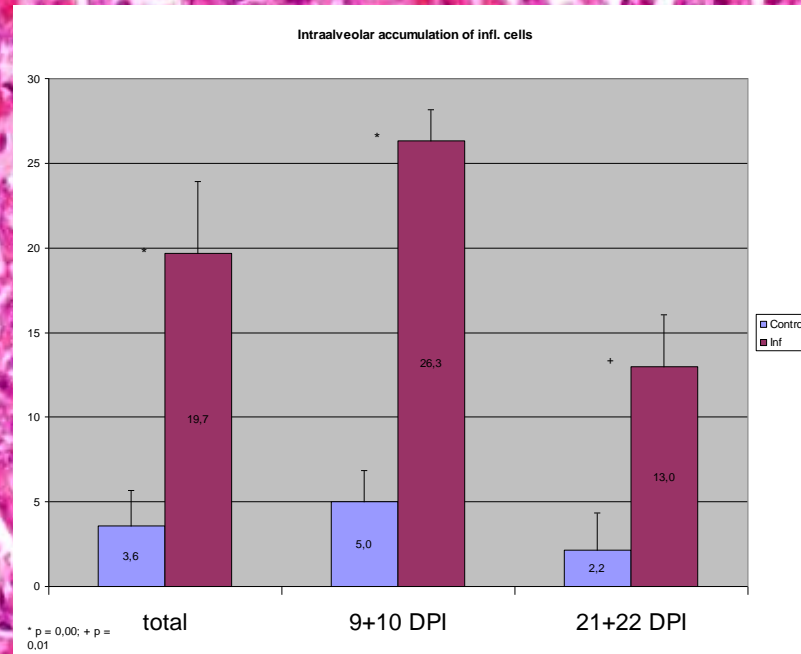
septal mononuclear infiltration



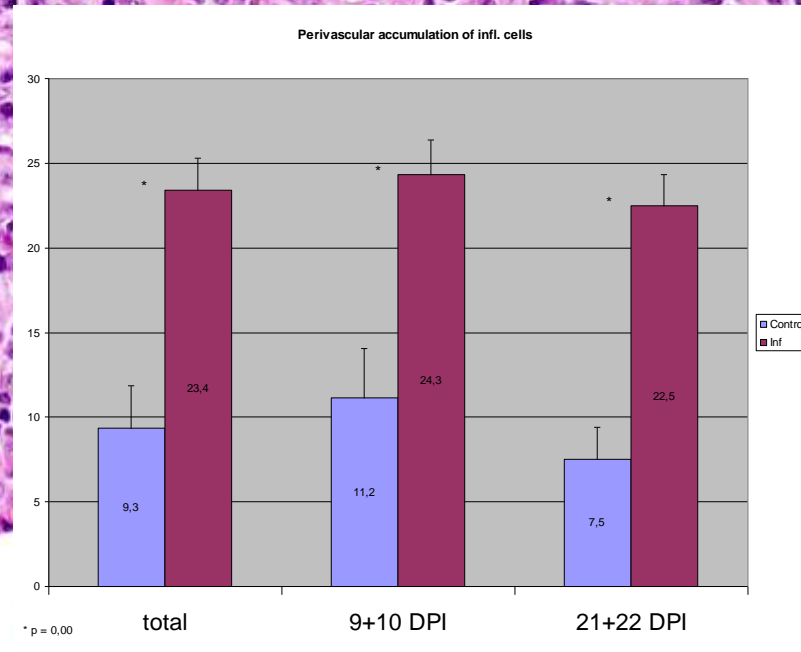
intraalveolar necrotic debris accumulation

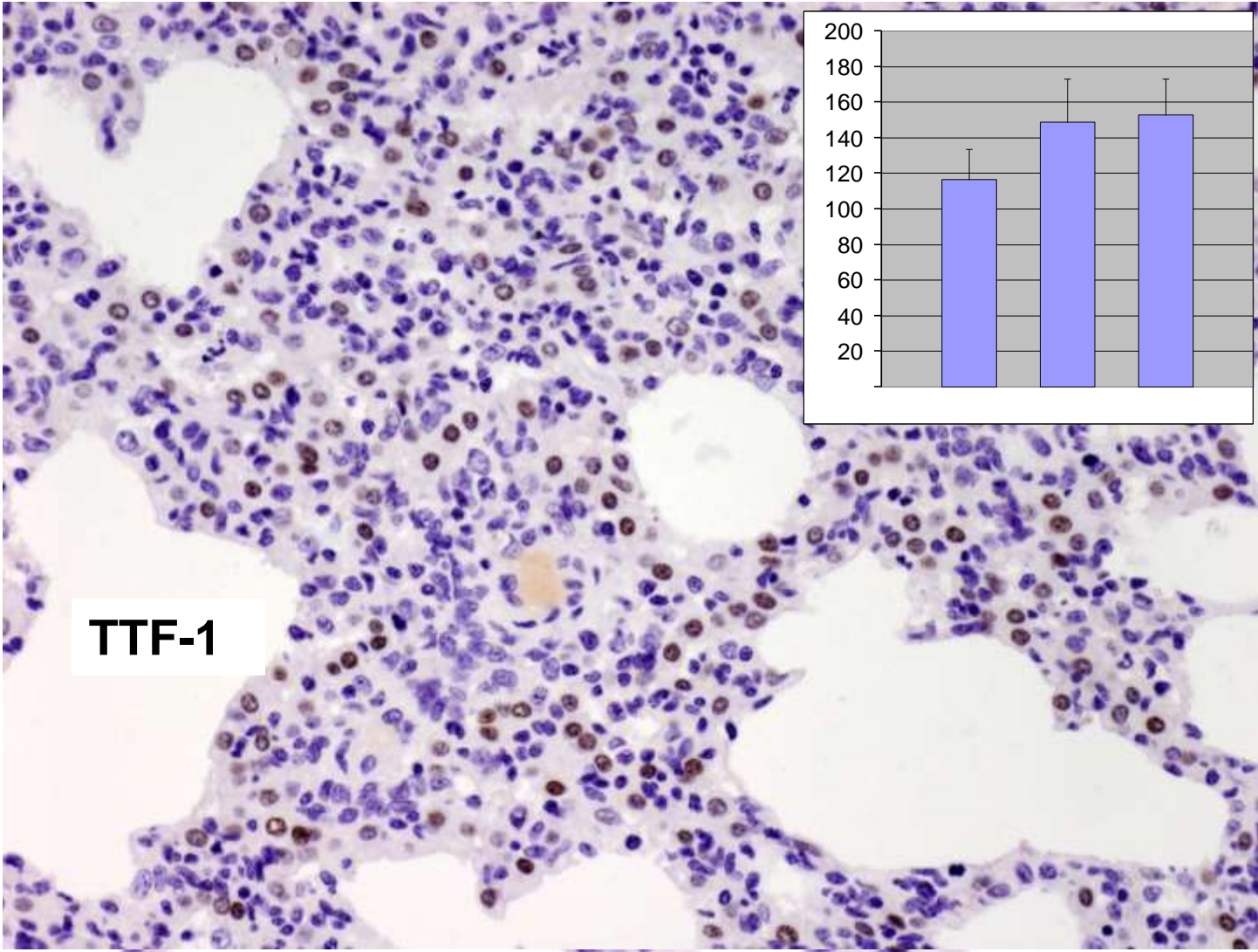


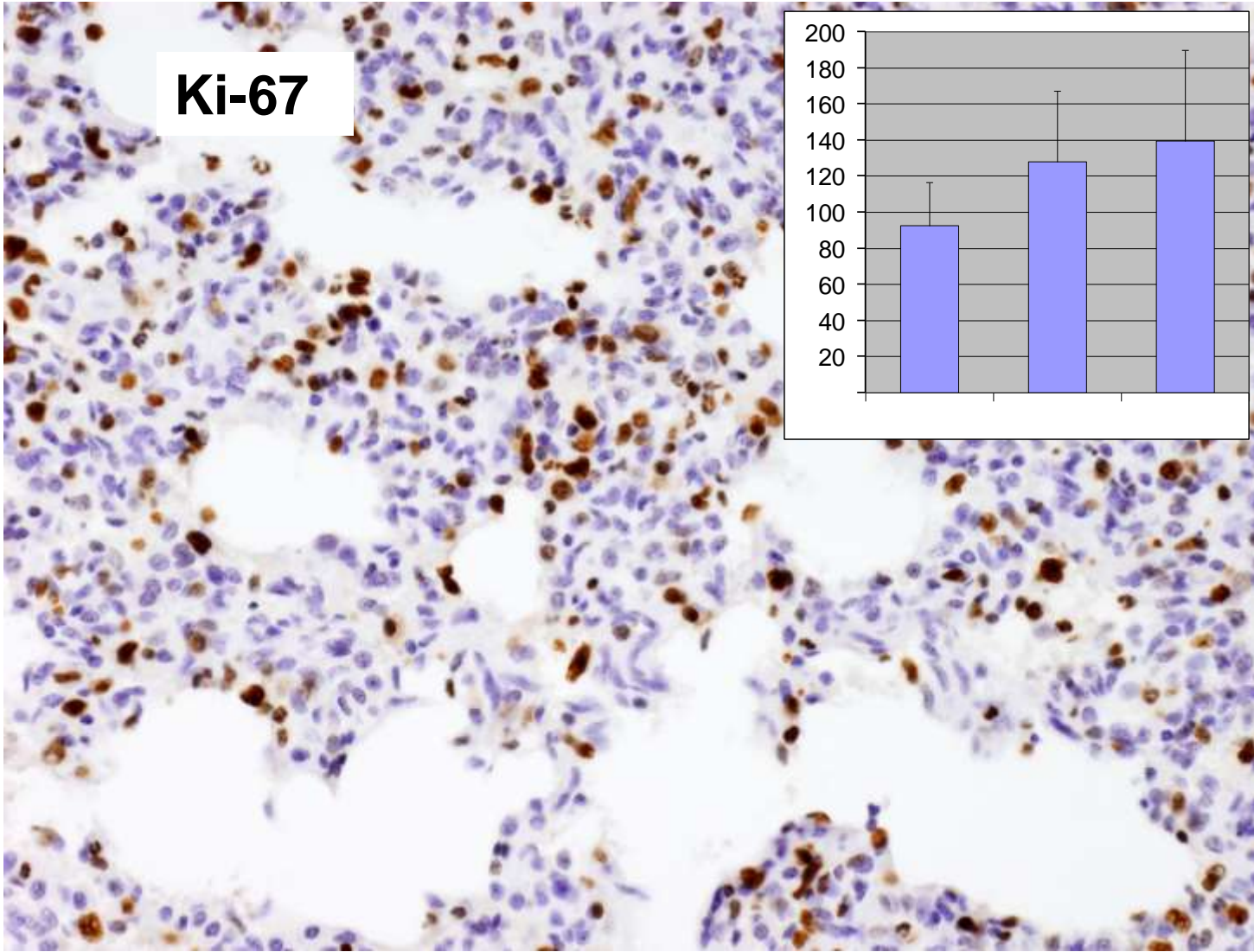
intraalveolar inflammatory cell infiltration

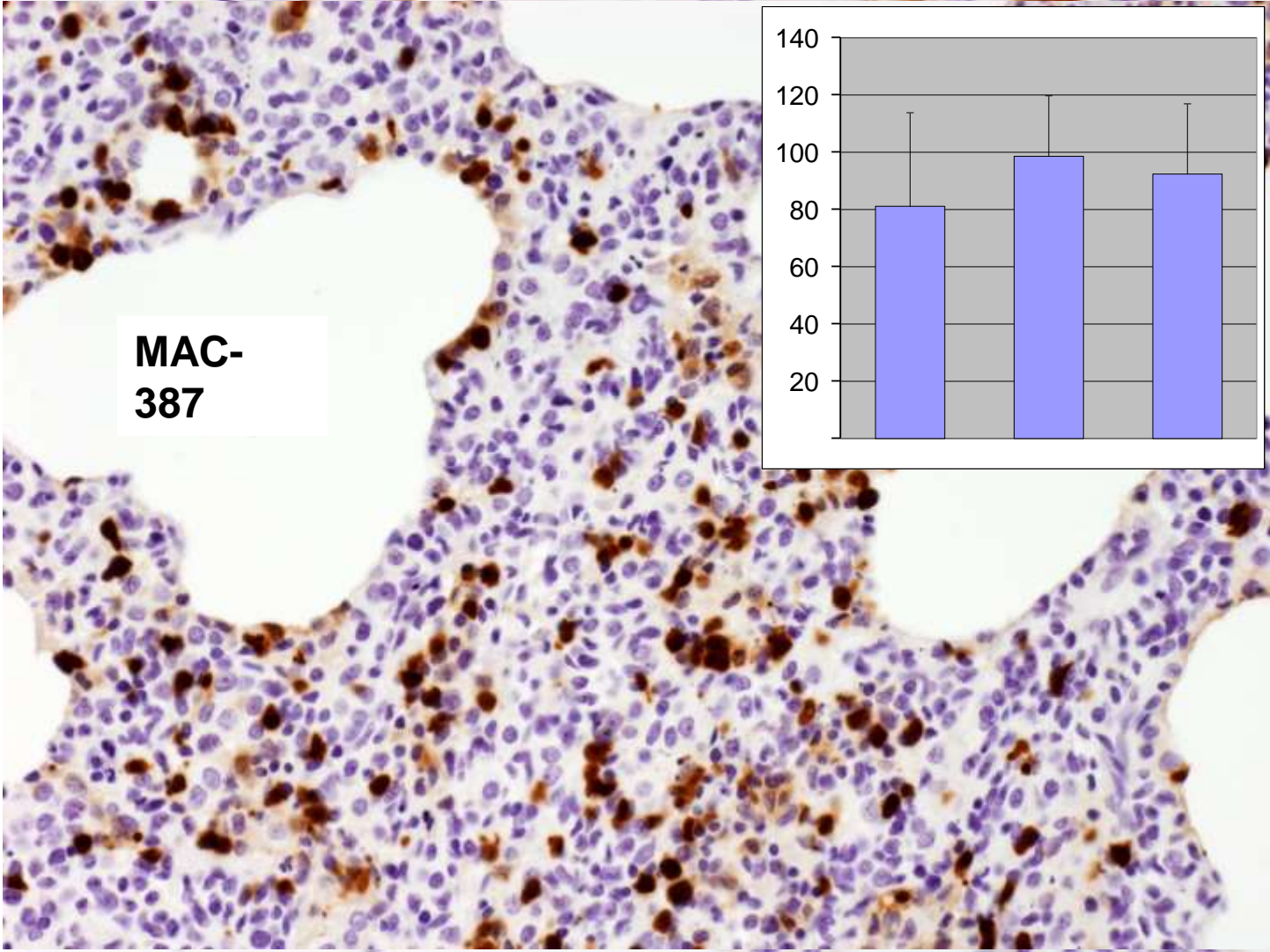


perivascular inflammatory cells

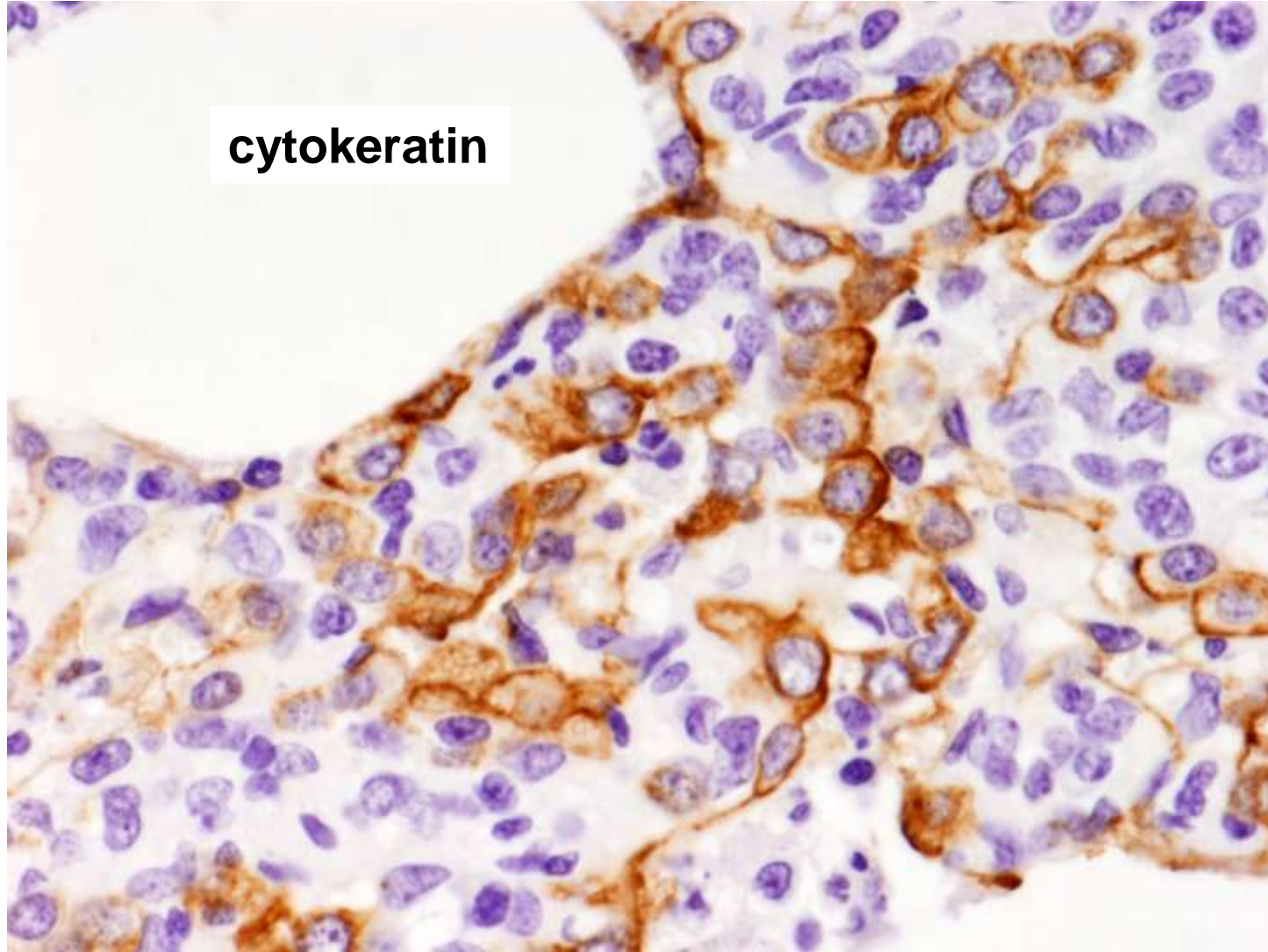




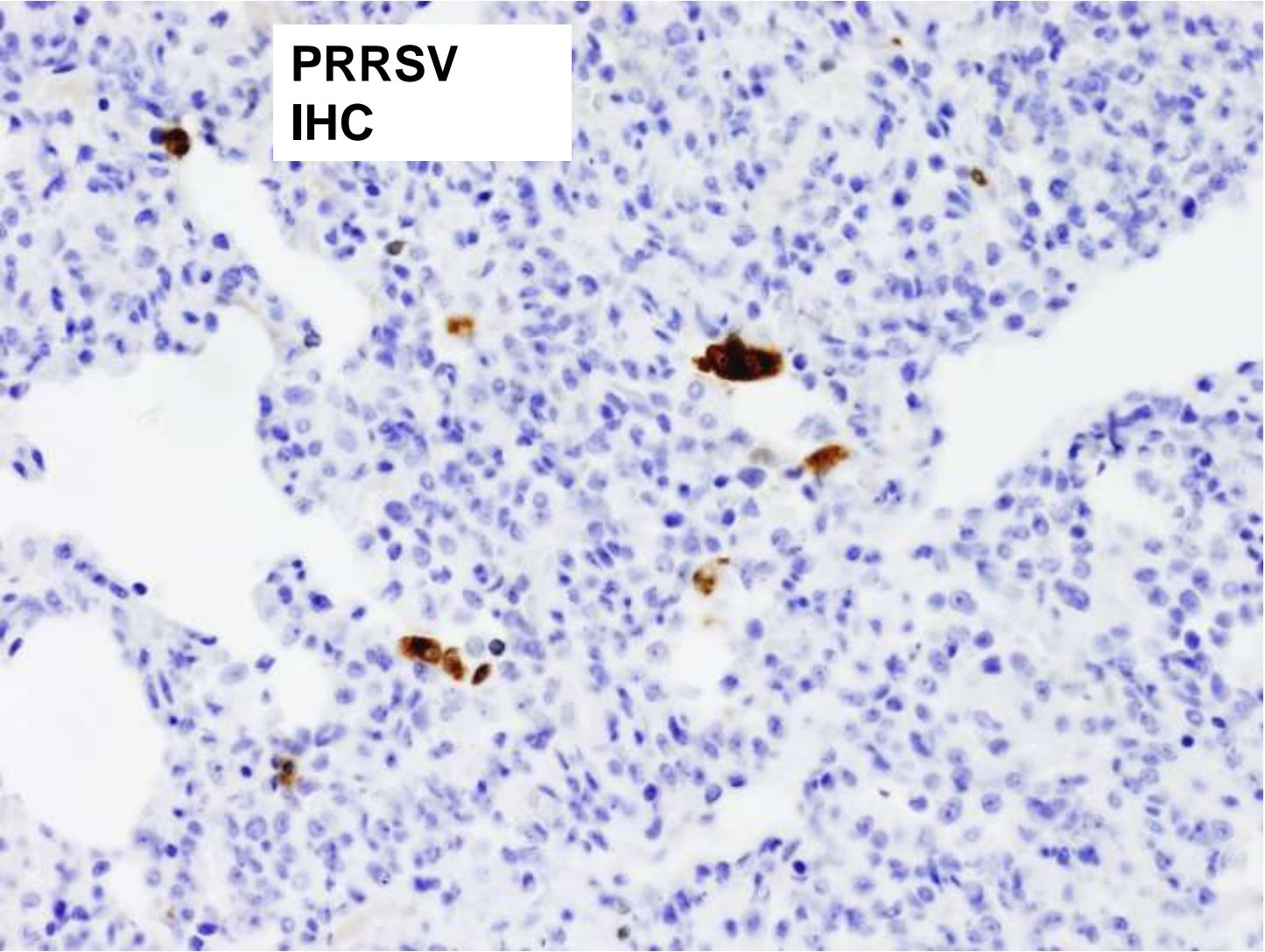




cytokeratin



**PRRSV
IHC**





EXPERIMENTALLY INDUCED DISEASE

**Immunohistochemical Characterization of Type II
Pneumocyte Proliferation after Challenge with
Type I Porcine Reproductive and Respiratory
Syndrome Virus**

**G. Balka^{*}, A. Ladinig[†], M. Ritzmann^{†,‡}, A. Saalmüller[§], W. Gerner[§],
T. Käser[§], C. Jakab^{*}, M. Rusvai^{*} and H. Weissenböck[¶]**

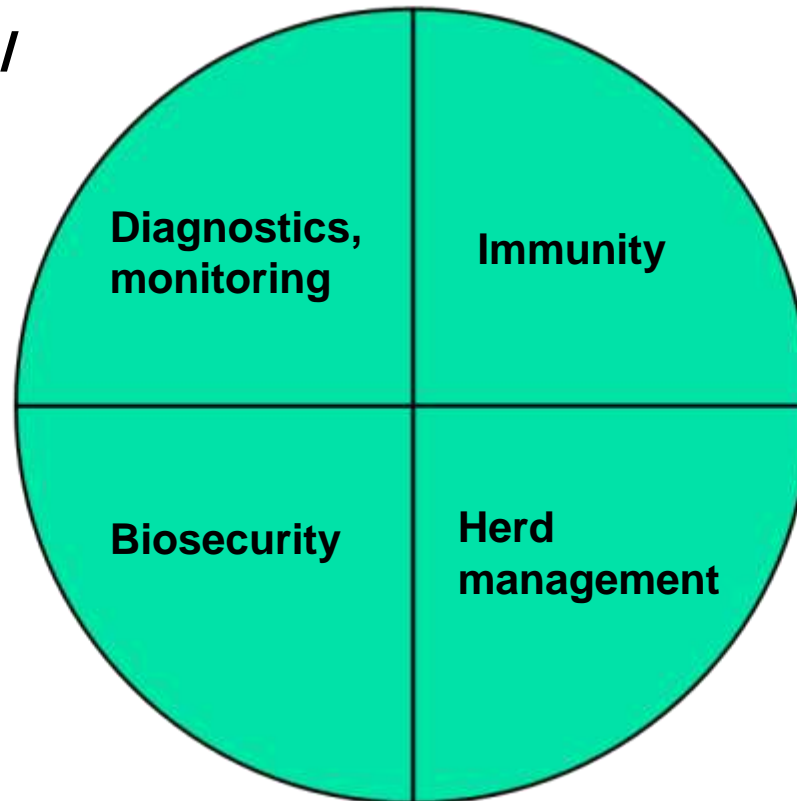
^{}Department of Pathology and Forensic Veterinary Medicine, Faculty of Veterinary Science, Szent István University, Budapest, Hungary, [†]Clinic for Swine, Department for Farm Animals and Veterinary Public Health, University of Veterinary Medicine, Vienna, Austria, [‡]Clinic for Swine, Ludwig-Maximilians University, Oberschleissheim, Germany, [§]Institute of Immunology, Department of Pathobiology and [¶]Institute of Pathology and Forensic Veterinary Medicine, University of Veterinary Medicine, Vienna, Austria*

Control of PRRS

Control – eradication

**Characterisation/
Early diagnosis**

**Gilt
acclimatization
Vaccination**



**External,
Internal**

**Reduction of
transmission,
reduction of the
economic impact**

After Mateu 2017

Control – eradication

- The key of control: decrease or stop virus excretion/circulation among the sows
 - homogenous immune status of the **breeding animals** (gilts introduction)
 - Piglet vaccination to reduce shedding
- **Eradication: LOAD-CLOSE-EXPOSE**
 - Min. 210 days, mass vaccination two times at least, then loading with naive gilts
 - Thorough examination of suckling piglets (PCR), then the sentinels (ELISA)
- **Prevent reinfection** – biosecurity (air filtration)
- **Vaccination + management!!!**

Linhares et al. 2013

- **TTPN** (time to produce negative piglets)
- **TTBP** (time to baseline production)

LVI (local/live virus inoculation) vs. **MLV**

- With LVI negative piglets earlier
- With LVI earlier herd stability

BUT!!!

The use of MLV was significantly more cost effective, less additional losses (abortions, medication, loss of income, etc.)

Vaccines

- **MLV**: strong reaction, horizontal and vertical spread
- **KV**: no AB reaction in naïve animals; booster effect after infection or MLV; harmless, no shedding, no spread
- Marker, subunit, peptide vaccines, genome shuffling

Vaccines

- Genetic heterogeneity has a negative impact on vaccine efficacy (Labarque et al. 2004), BUT
- Degree of (ORF5) similarity can **NOT** predict vaccine efficacy (Prieto et al. 2008)
- Different strains (vaccines) induce different cytokine production patterns (Díaz et al. 2006)
- Vaccines do not prevent infection and viraemia after heterologous challenge, BUT in a natural infection model they can provide 70% clinical protection (Martelli et al. 2009)

Suvaxyn PRRS challenge trial

- 41 piglets involved in the study born to 4 sows
- Piglets were randomly cross fostered after birth, then 2 litters got vaccinated

Treatment Group	Test Material	Dose Volume per Admin (mL)	Day of Admin	Challenge	Day of challenge	Day of necropsy**
T01	Saline solution (CP)	2 mL IM	D0	PRRSV-1 subtype 1 strain AUT15-33 by IN route at 1×10^6 cfu total in 5ml	D28	D41/42
T02	Suvaxyn PRRS MLV (IVP)					

- At weaning (D25) former littermates were put back together

Suvaxyn PRRS challenge trial

- Challenge at day 28 with highly pathogenic PRRSV-1, Subtype 1 „ACRO” strain AUT15-33
- Clinical observation, rectal temperature, body weight
- Serum samples, nasal swabs, oral swabs
 - ELISA and PCR
- Euthanasia at day 41/42
- Necropsy
 - Macroscopical evaluation (% affected)
 - Lung tissue samples for PCR and histopathology

Suvaxyn PRRS challenge trial

- Histopathology (Balka et al. 2013, J. Comp. Pathol.)

BLINDED ANALYSIS

- Pneumocytic hypertrophy and hyperplasia
- Septal infiltration with mononuclear cells
- Intraalveolar necrotic debris
- Intraalveolar accumulation of inflammatory cells
- Perivascular accumulation of inflammatory cells

Suvaxyn PRRS challenge trial

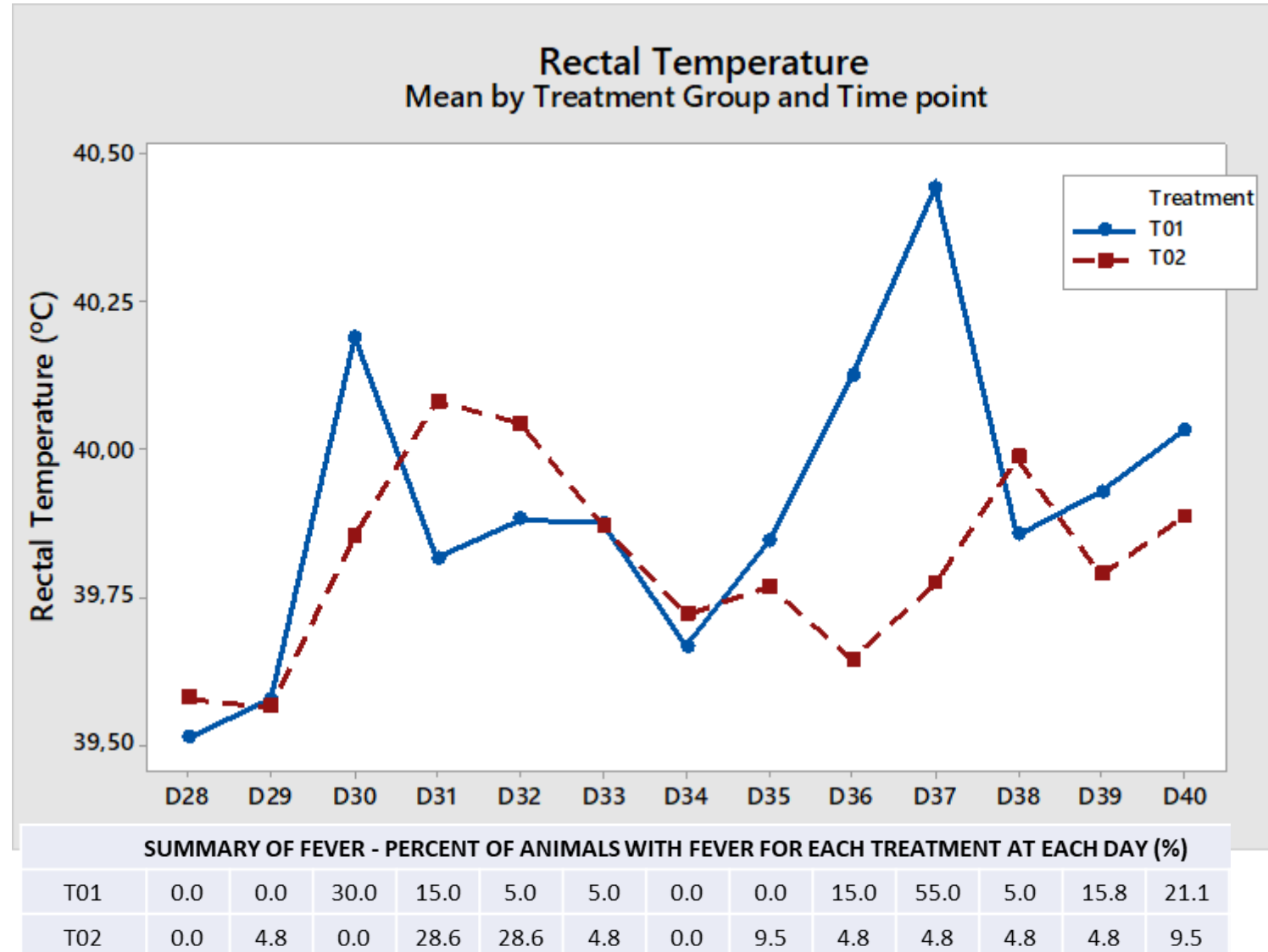
RESULTS

ANALYSIS OF **BODY WEIGHT** - COMPARISONS OF AVERAGE **DAILY GAIN** BETWEEN TREATMENTS

Label	Difference in average daily gain	std error of diff. in average daily gain	2-tailed p-value (1)	Significance of 2-tailed p-value (2)
28 to 35 ADG T01 v T02	0.07	0.02	0.0013	*
28 to 41/42 ADG T01 v T02	0.07	0.03	0.0173	*
35 to 41/42 ADG T01 v T02	0.07	0.04	0.0971	N.S.

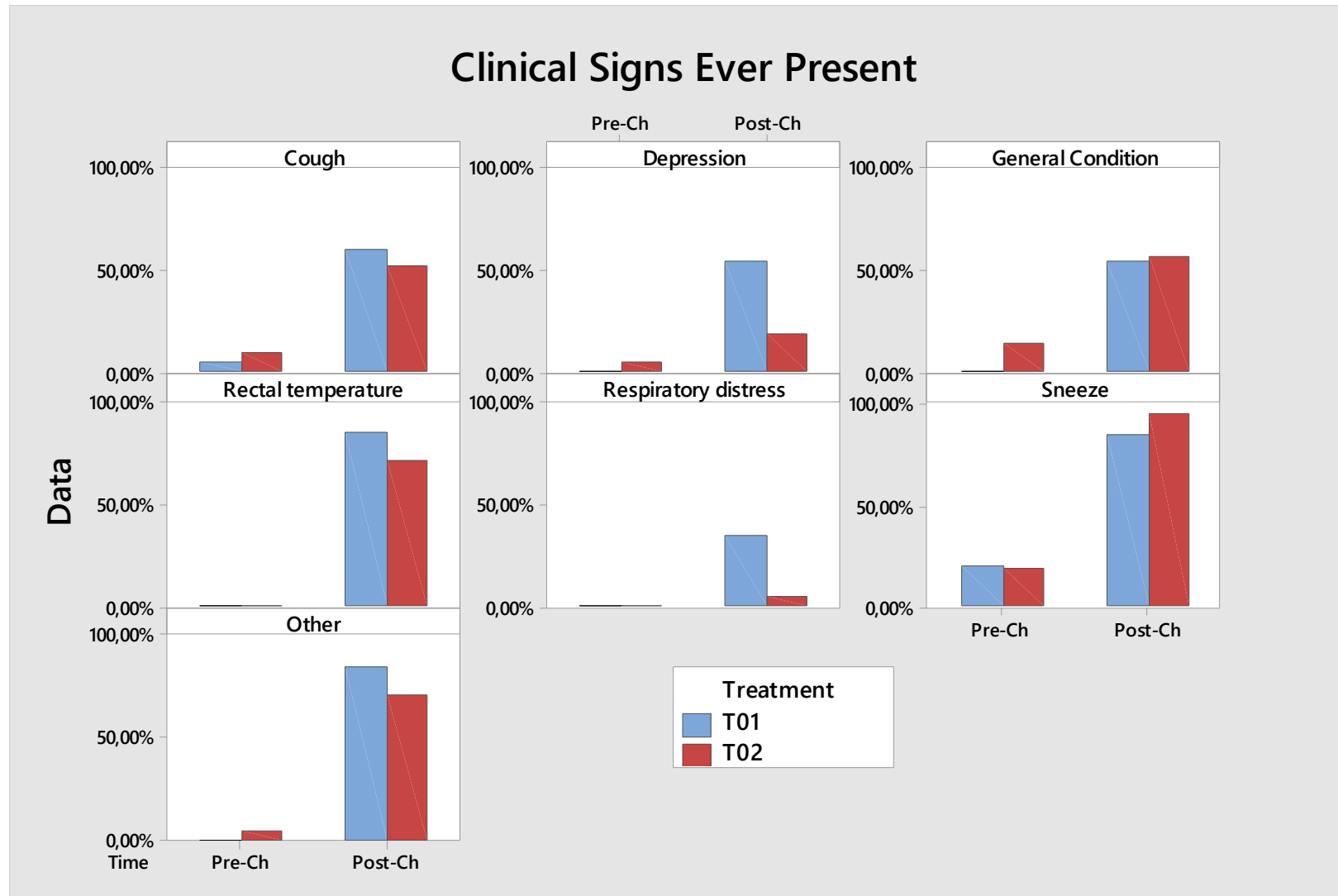
Suvaxyn PRRS challenge trial

RESULTS



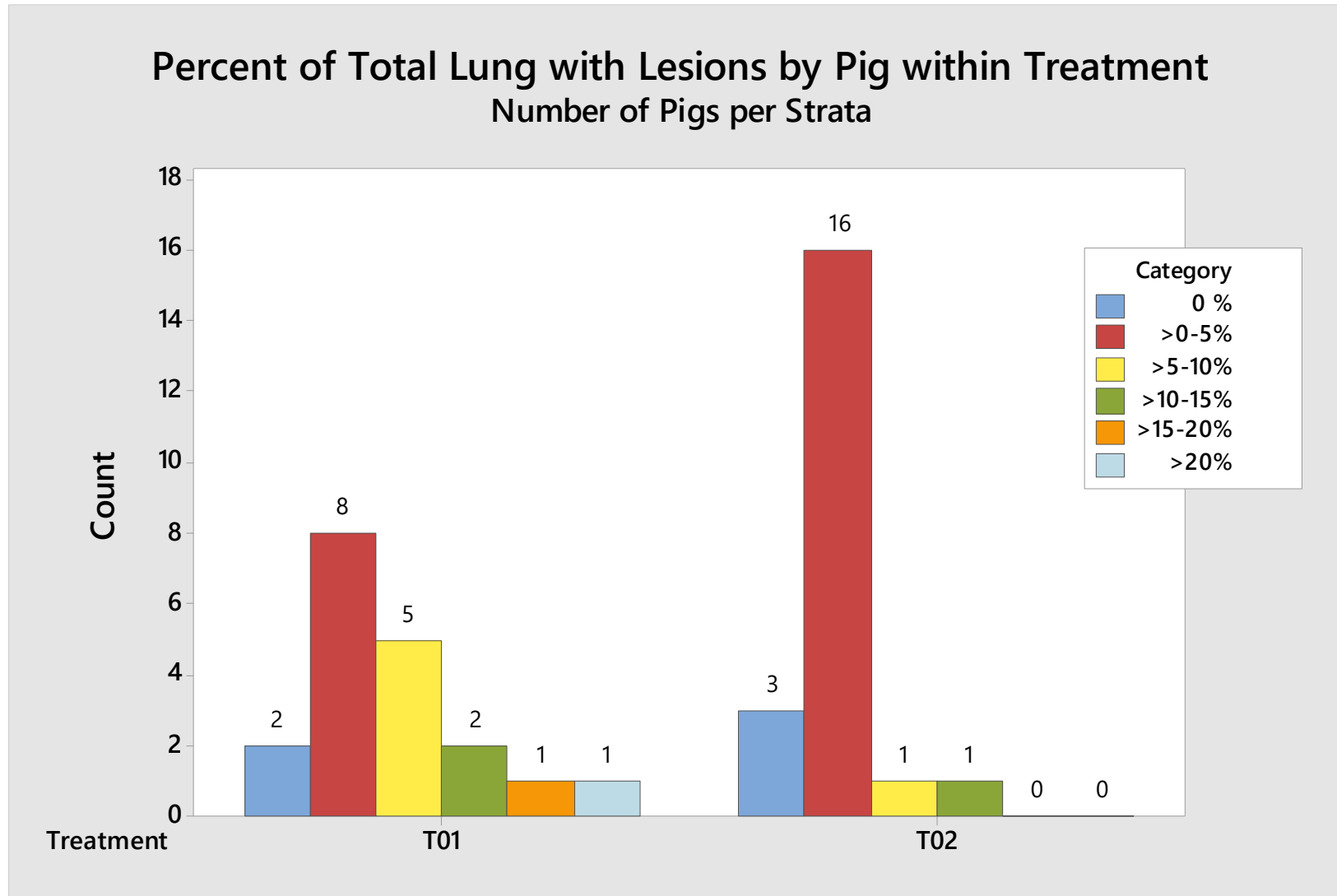
Suvaxyn PRRS challenge trial

RESULTS



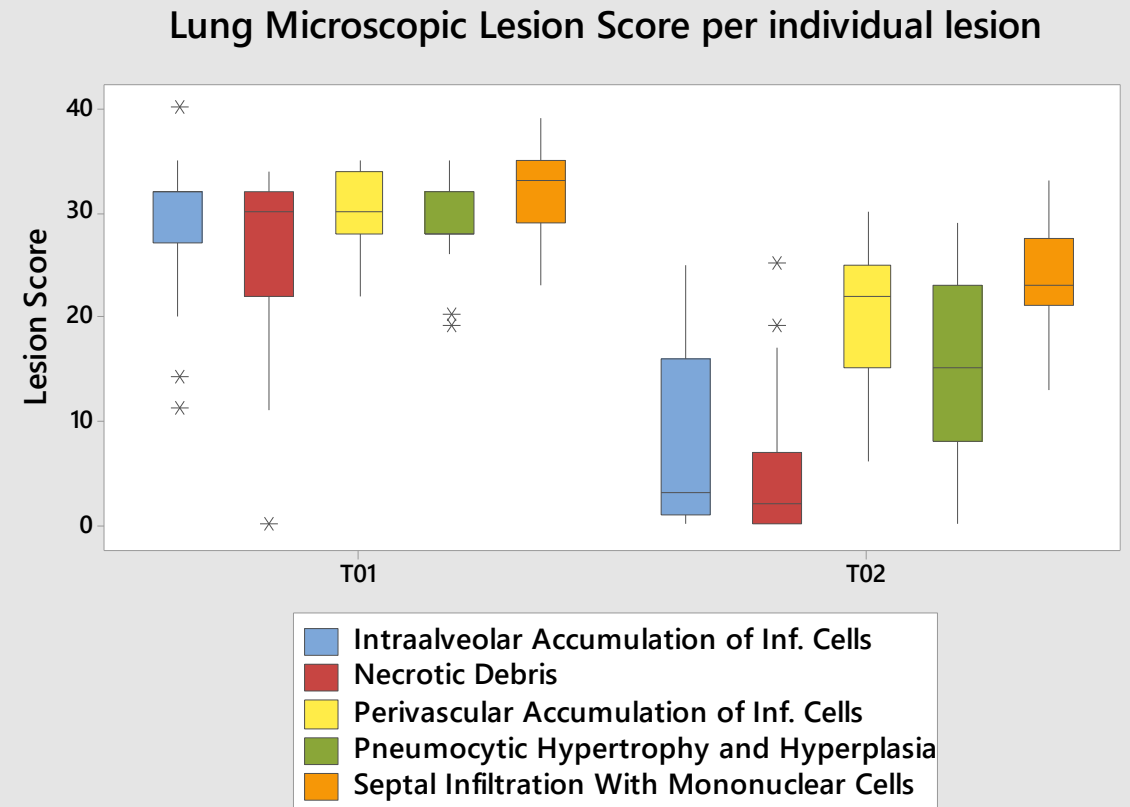
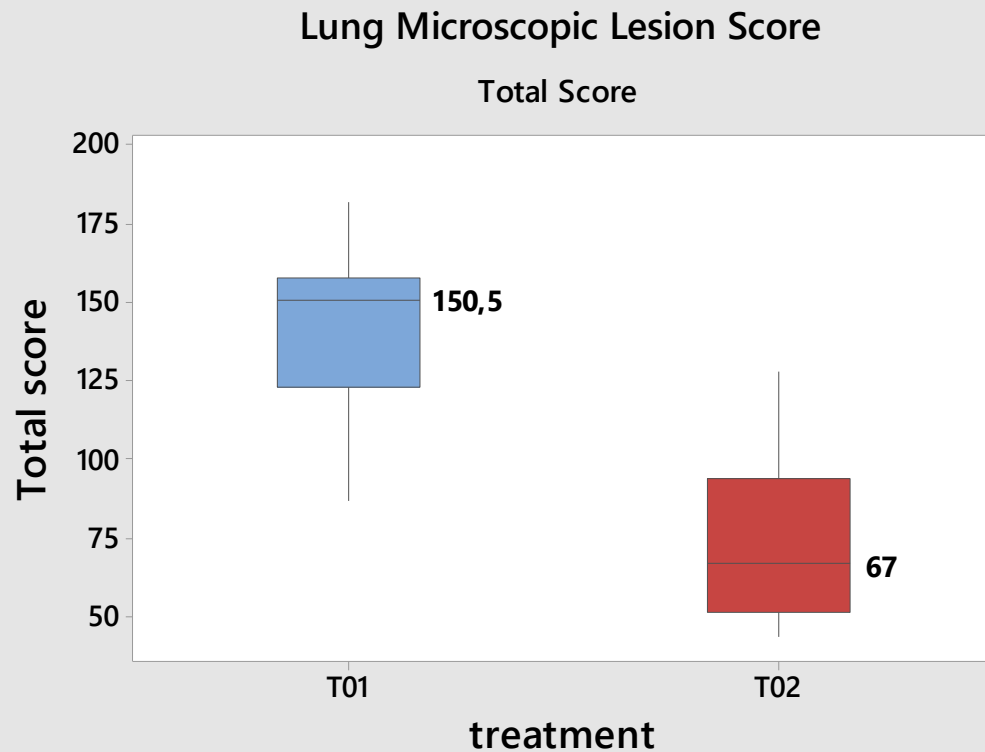
Suvaxyn PRRS challenge trial

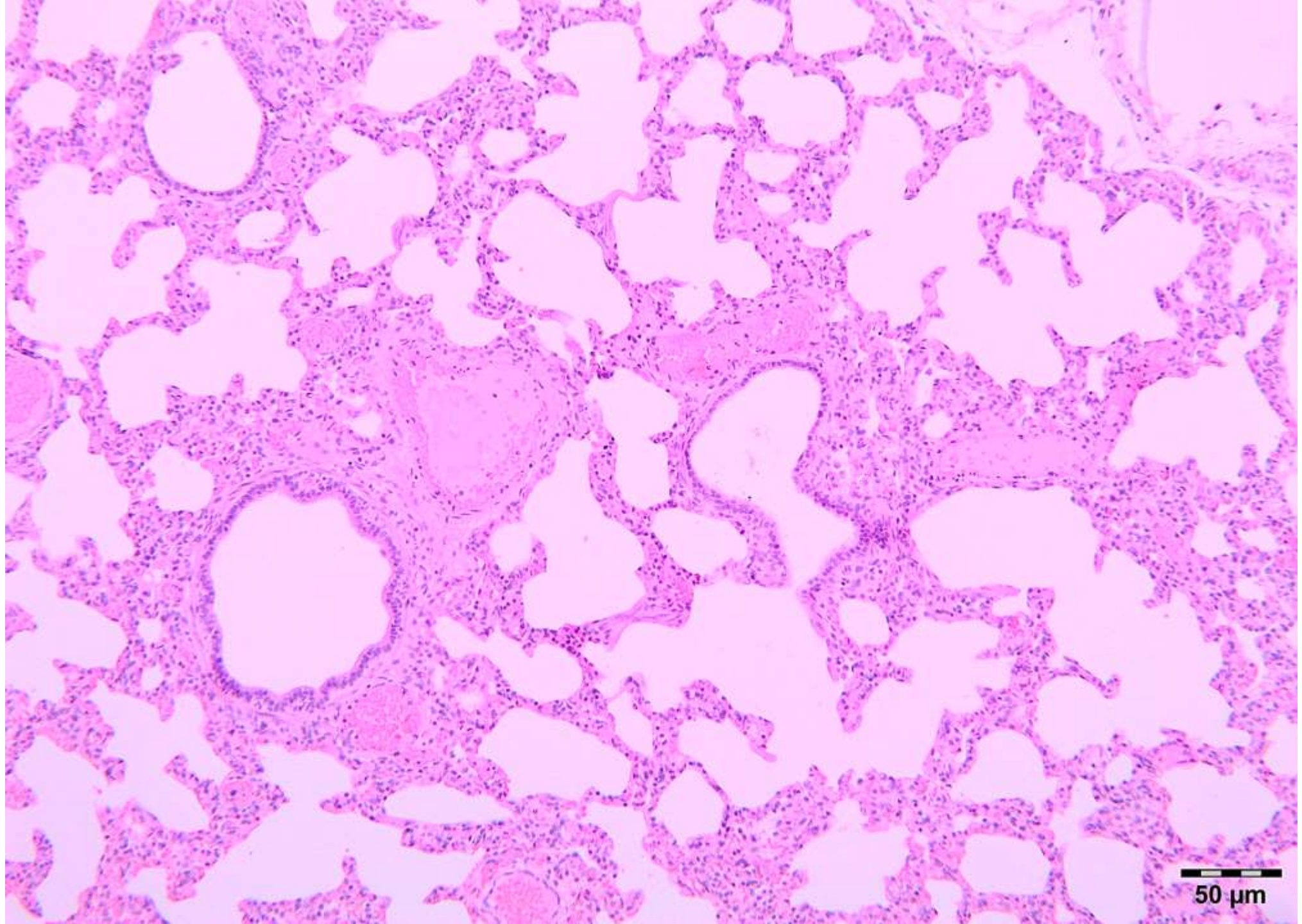
RESULTS



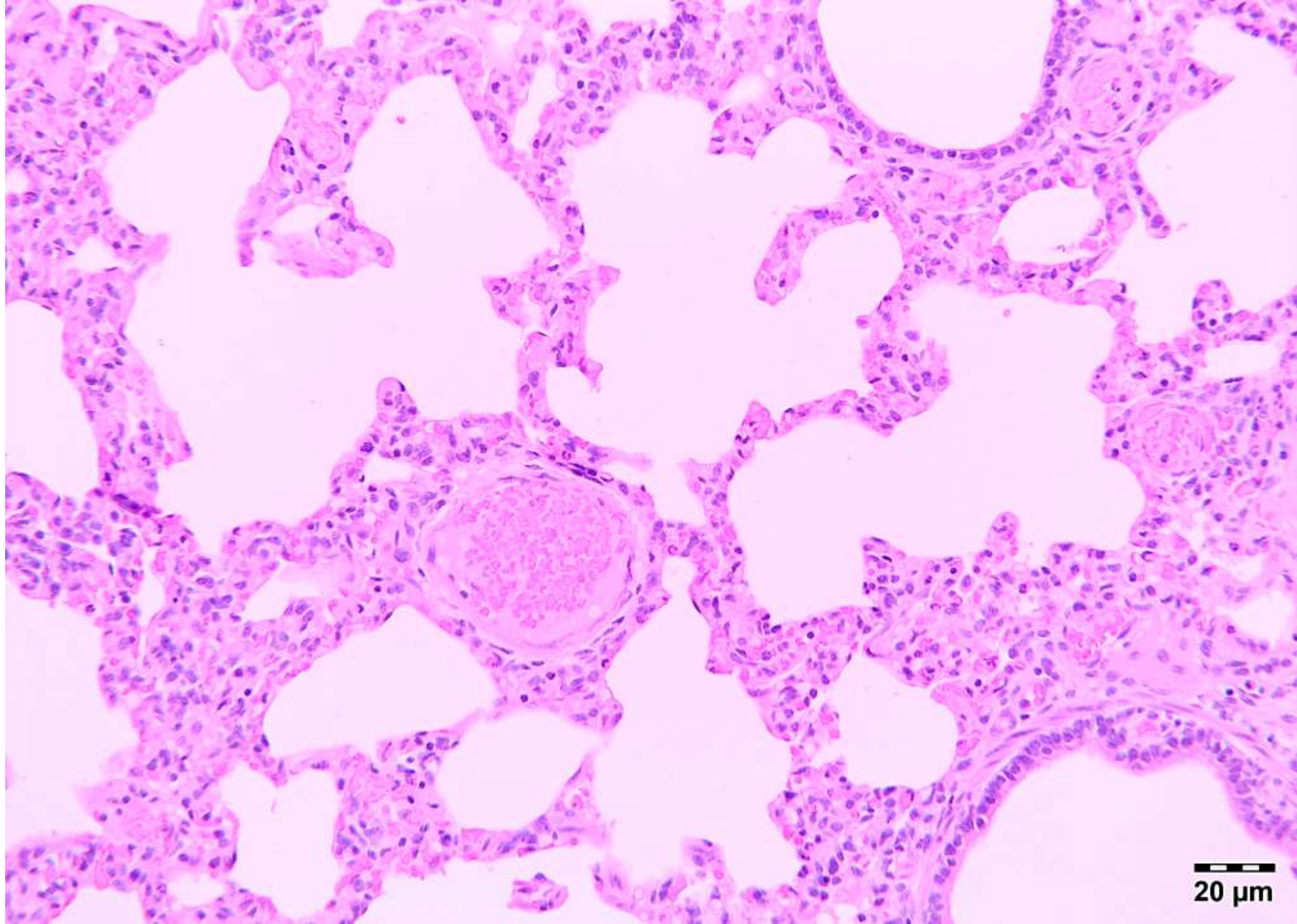
Suvaxyn PRRS challenge trial

RESULTS – summary of histological data

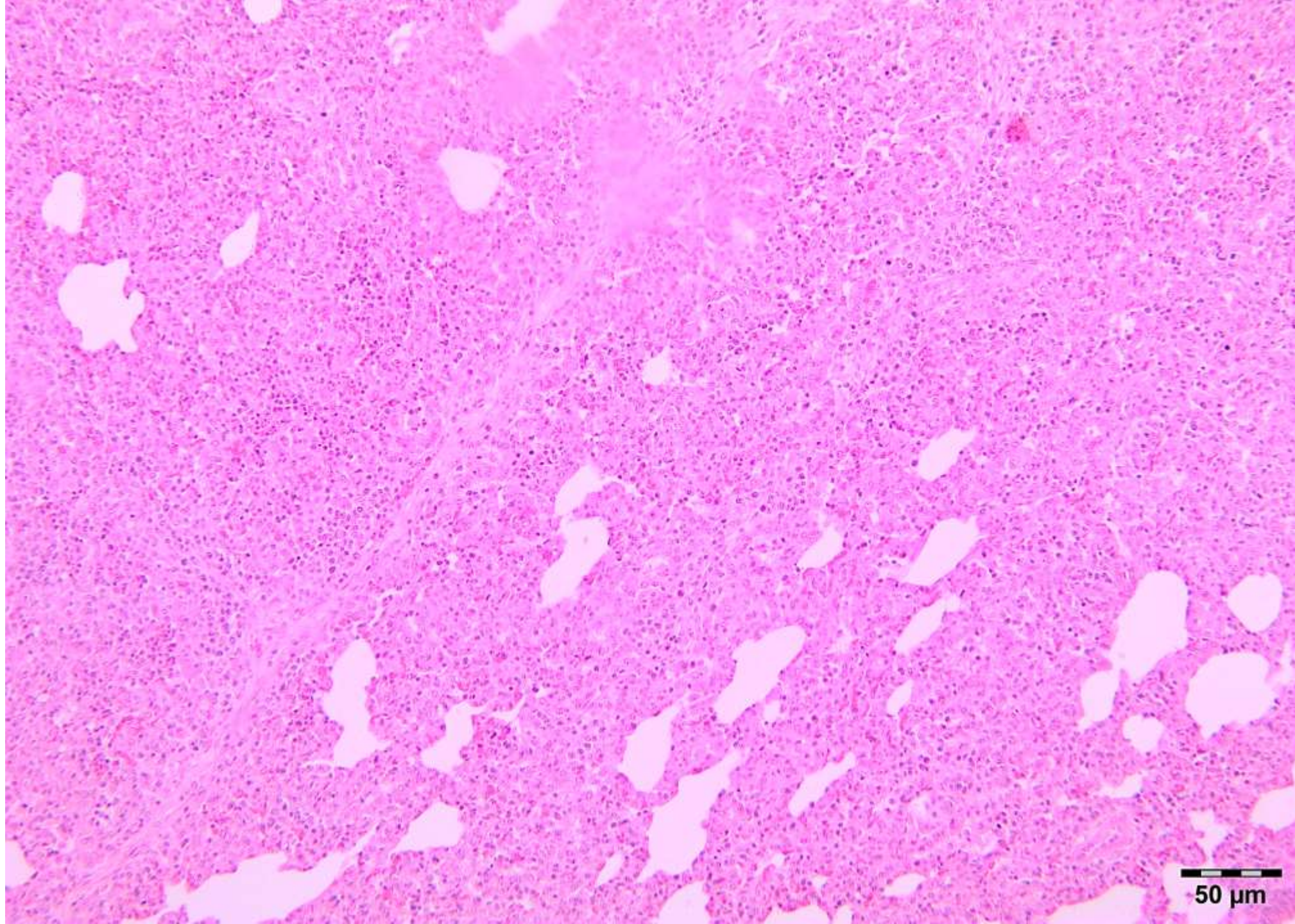




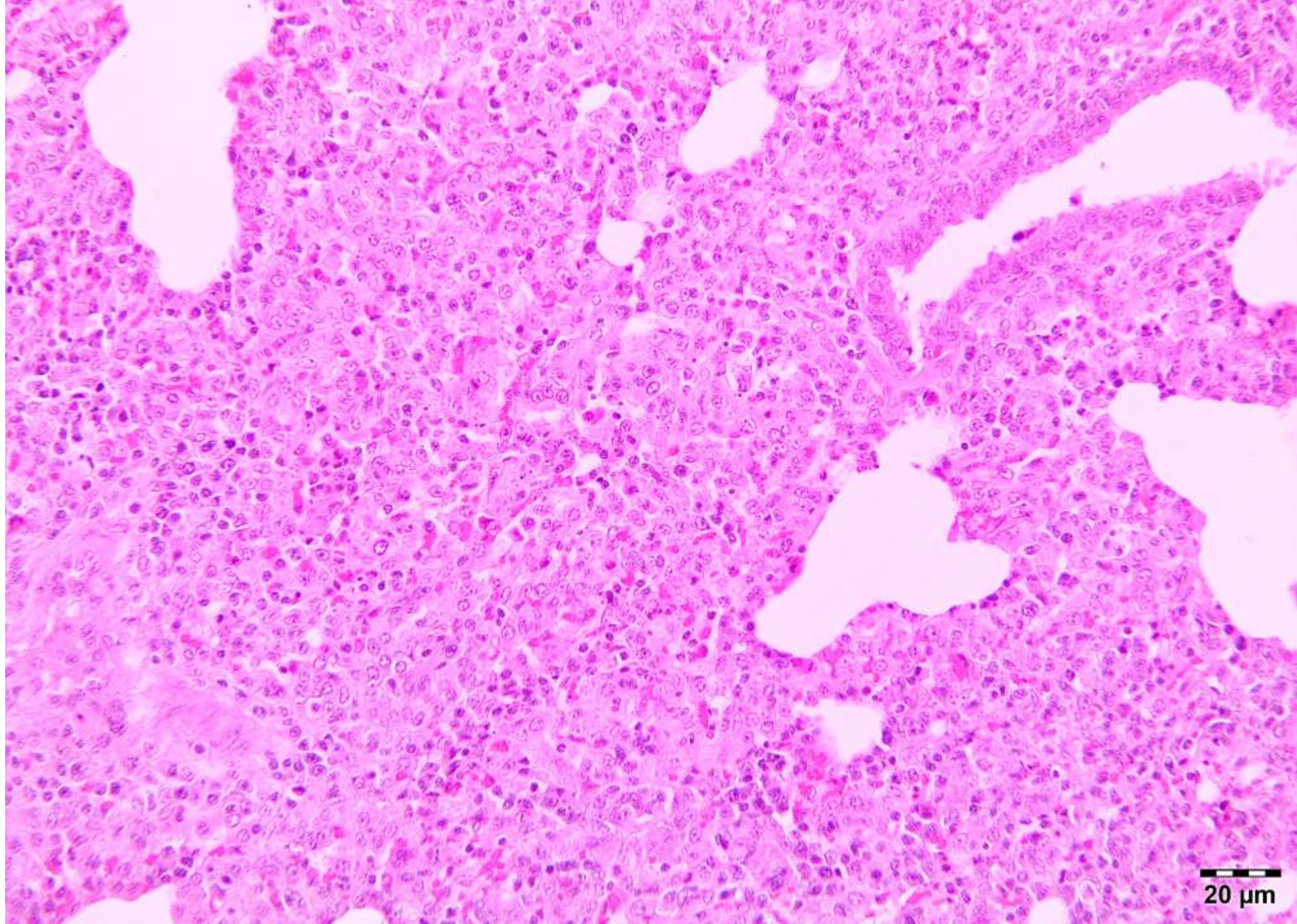
50 μ m

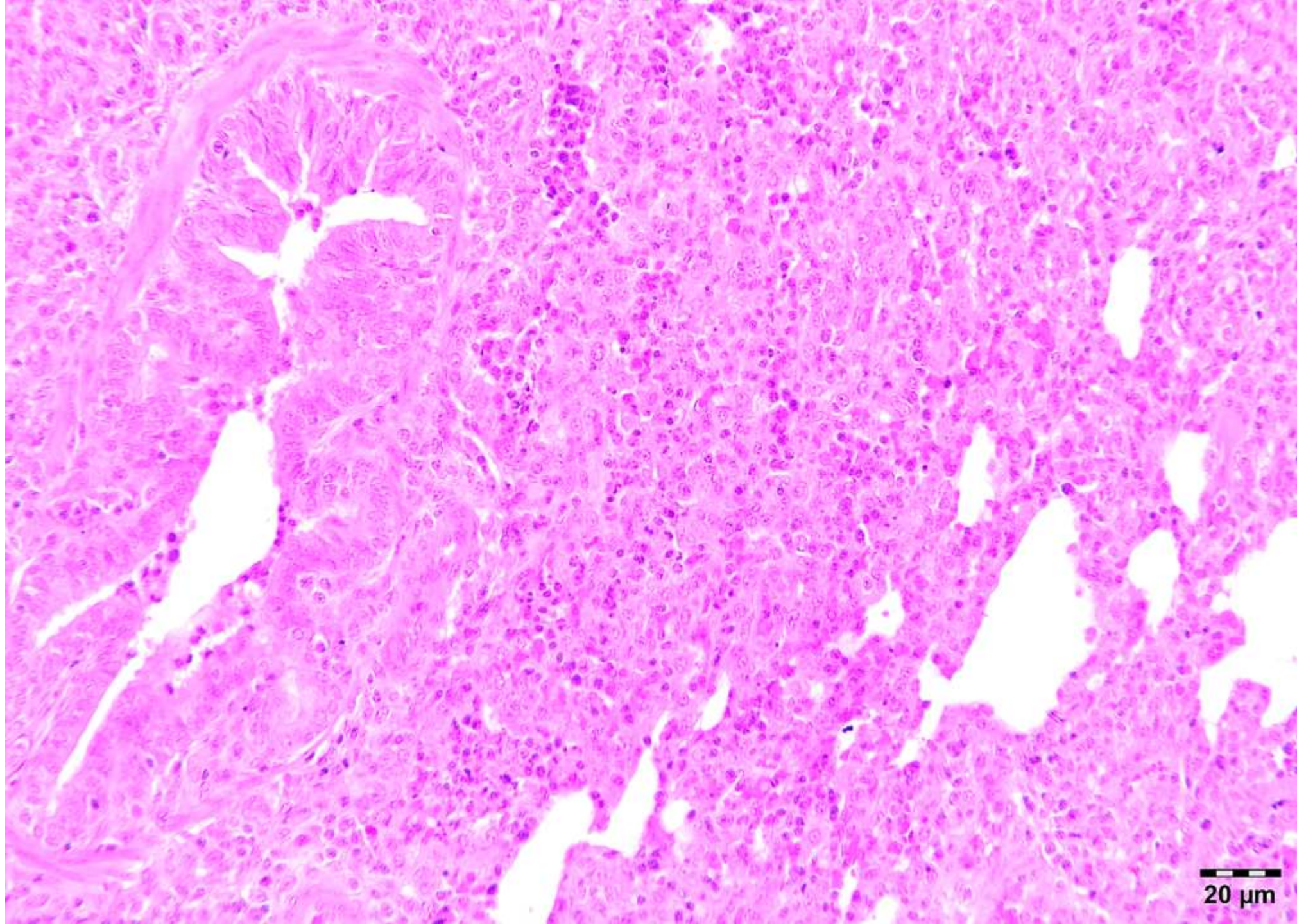


20 μ m

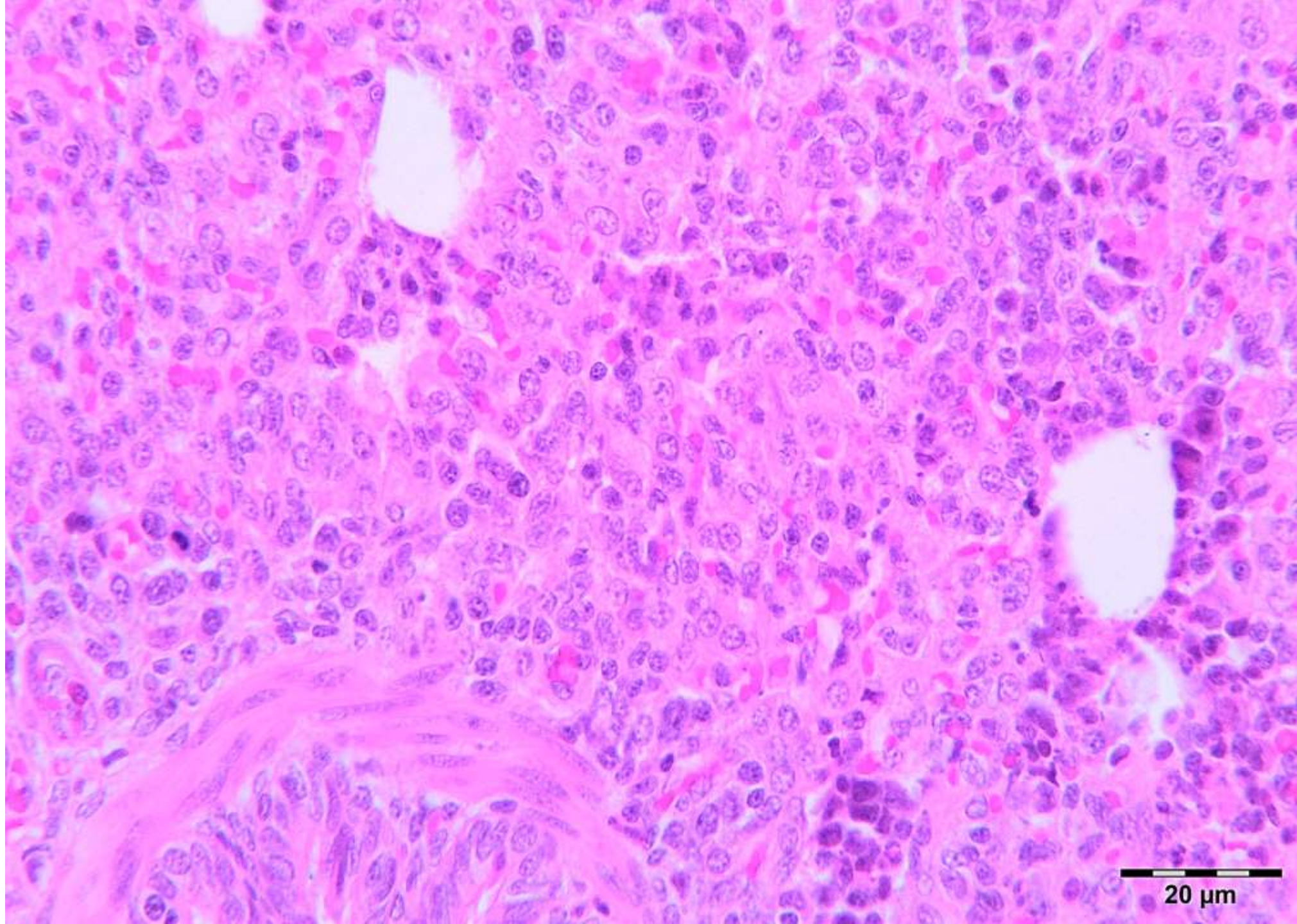


50 μm





20 μm



20 μm

Thank you for your attention!